Study of the Effect of Lampeni (*Ardisia humilis* Vahl.) Planting Condition toward the Alpha-glucosidase Inhibition Activity *In vitro*

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ABSTRACT

Background: The quality of a medicinal plant is influenced by agronomic conditions and harvesting time. Objective: This study aimed to evaluate the effect of planting method (openair (OA) and shedding house (SH)) and harvesting time (2, 4, 6 months) of Lampeni (Ardisia humilis Vahl.) toward the inhibitory activity of alpha-glucosidase. Methods: The Lampeni seedling were placed under controlled light conditions (SH) and on direct sun exposure (OA). Harvesting of the leaves was carried out at the age of 2, 4, and 6 months after plantation (2m, 4m, and 6m). Each leaves dry powder was refluxed with methanol 70% and followed by liquid-liquid partition using n-hexane, ethyl acetate (EtOAc), and water. All samples were evaluated toward inhibition of the alpha-glucosidase enzyme in vitro. Total phenol levels were determined using Folin-Ciocalteu reagent. Results: The results showed that EtOAc fractions of both plantation techniques exhibited the highest inhibition of alpha-glucosidase. The highest activity was demonstrated by the 4m-OA-EtOAc fraction (IC $_{\rm 50'}$ 93.50 ppm) and followed by the 6m-OA-EtOAc fraction (IC $_{\rm 50'}$ 98.13 ppm). Based on the kinetic study, the inhibition type of the two most active samples were categorized as a non-competitive type. Total phenolic contents were decreased in the following order: 6m-SH-EtOAc > 4m-OA-EtOAc > 6m-OA-EtOAc > 4m-SH-EtOAc fraction. It was shown that there was no positive correlation between the strength of inhibition with total phenolic content. Conclusions: This study concluded that Lampeni at open-air plantation harvested on fourth months demonstrated the highest alpha-glucosidase inhibitory activity, although there was no positive correlation between the inhibition activity and phenolic content.

Key words: Alpha-glucosidase, Ardisia humilis Vahl., Lampeni, Open-air, Shedding house, Total phenolic.

INTRODUCTION

Diabetes mellitus (DM) is a disease characterized by the rise of blood glucose levels beyond normal (hyperglycemia) caused by a defect of insulin production and/or insulin response. Postprandial hyperglycemia is the elevation of plasma glucose after a meal. There were some shreds of evidence that the loss of postprandial glucose control both in DM or non-DM people may be an independent risk factor and could be a potential cause of the cardiovascular disease.1 Absorption of starch hydrolysis product is the main source of increased glucose in the blood. A group of enzymes, including pancreatic alpha-amylase and intestinal alpha-glucosidase, are directly involved in starch hydrolysis. Inhibition of these hydrolytic enzymes is believed to be an attempt to control postprandial blood sugar levels.² Conventional DM management using chemical drugs such as biguanides, sulfonylureas, meglitinide, thiazolidinedione (TZD), dipeptidyl peptidase 4 (DPP-4) inhibitors, sodium-glucose cotransporter (SGLT2) inhibitors, and a-glucosidase inhibitors are known to cause some side effects. Some evidence of the side effects of synthetic hypoglycemic drugs such as

hypoglycemia, digestive, and central nervous system disorders had been reported.³ Finally, finding new drugs from natural resources was a strategic effort for solving these problems.

Indonesia is one of the tropical countries that have abundant natural resources from plants, animals, or minerals. Lampeni known as Ardisia humilis Vahl. belongs to Magnoliatae family is a woody plant that is commonly found in the Indonesian forest. This plant had been used as folk medicine with empirical activities such as stimulant, carminative, antidiarrheal, treat rheumatism, skin sore, and vertigo⁽⁴⁾ besides as a decorative plant. Previous study reported that the methanolic extract of A. humilis demonstrated cytotoxic, thrombolytic, and antioxidant activities.4 Mice treated with ethanolic extract of Lampeni at the dose ranged from 100 to 300 mg/kg bw demonstrated antiplatelet activity by prolonged bleeding time.5 Lampeni also has blood glucose-lowering activity in which alpha-amiryn compound was responsible for these biological properties.6 The other chemical compound from the other species Ardisia elliptica named beta-amiryn was known for having more potent antiplatelet activity than aspirin.7

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It has been a common knowledge that one of the main roles in the production of plant phytochemicals is the growth conditions. The content of chemical compounds in a medicinal plant affects the pharmacological properties. Blum-Silva, et al.8 reported that the level of polyphenol and methylxanthine content of Ilex paraguariensis was found higher in the older leaves than the young one and exhibited a difference in the strength of pharmacological activity. Meanwhile, it was also stated that the quantitative analysis of some phytochemical compounds from Ilex paraguariensis were affected by light intensity and the age of the leaves.9 Implementation of a good agricultural process (GAP) will produce high quality medicinal plants.¹⁰ The purpose of this study was to evaluate the effect of agronomic (light intensity and harvesting time) to obtain Lampeni leaves that could demonstrate alpha-glucosidase inhibitory properties. The parameters tested were the light intensity (open-air/direct sun exposure and shedding house) and the harvest time (2, 4, and 6 months).

MATERIAL AND METHODS

Experimental design

The scheme of experiments was depicted as in the Figure 1.

Materials

Chemicals used in this experiment were methanol (Merck), chloroform (Merck), n-hexane (Merck), ethyl acetate (Merck), chloride acid (Merck), aquadest (local market), phosphate buffer solution pH 7.0 (Merck), p-nitrophenyl- α -D-glucopyranoside (Sigma), alpha-glucosidase (Sigma), bovine serum albumin (Sigma), dimetyl sulfoxide (Merck), sodium carbonate (Merck), acarbose tablet, folin ciocalteu reagent (Merck), gallic acid (Sigma), toluene (Merck), formic acid (Merck), growing medium consist of soil, sand, compost, Bioroot^{*}, Antracol^{*} fungicide (Bayer), Agrept^{*} bactericide, Biopex^{*}.

Fully grown 15-20-year-old tree of Lampeni from Ujung Kulon National Park Banten Indonesia was selected as the mother plant. Determination of the plant sample was conducted at LIPI Biology Center, Cibinong, Bogor, Indonesia. Plant seeds were prepared based on the protocol developed by Laboratory of Biotechnology, Agency for The Assessment and Application of Technology. Briefly, the stems containing shoots of the mother plant were cut and soaked in a bactericidal-fungicide solution for 15 minutes and then drained by bloating to tissue paper. Root inductions were carried out by applying Bioroot' paste contained hormones at the base of the bottom of the bud. The seedling were then planted in media consisting of the proportionate sand and soil in a polybag. In order to stimulate the shoot growth, the shoots were sprayed with the Biopex' hormone solution. Polybags were placed in two conditions, namely, in a shedding house (at a temperature of 25- $30 \pm 1^{\circ}$ C for 16 hours photoperiod and 50-70% relative humidity) and direct exposure to the sun (open-air, OA) with 45 polybags in each treatment. At this time, the age of leaves was set to zero months. The leaves were collected on 2, 4 and 6 months from each 15 polybags after planting without the top 3-4 leaves, dried under 50°C using an oven and then crushed until powder mass obtained for the next process.

Extraction and fractionation process

The crude extract of each planting method was prepared by reflux technique. Each Lampeni leaves dry powder was refluxed using 70% methanol for 3 hours. The filtrate was concentrated by a rotary evaporator until semi-solid mass obtained. Toward each semisolid crude extract was added to distilled water and shaken until a homogenous mass obtained. The suspension was then partitioned using n-hexane in a glass separating funnel, followed with ethyl acetate. Each filtrate was concentrated by vacuum evaporator, as stated above. From these extraction processes, there were some samples obtained, namely, a crude extract, n-hexane fraction (Fr.n-hexane), ethyl acetate fraction (Fr.EtOAc), aqueous fraction (Fr.Aquos) of both SH and OA plant that collected at 2, 4, and 6 months.

Alpha-glucosidase inhibitory evaluation in vitro

Evaluation of alpha-glucosidase inhibitory activity was conducted using p-nitrophenyl-alpha-D-glucopyranoside/alpha-glucosidase system based on the previous study¹¹ with modification. Briefly, the enzyme stock solution was prepared by adding 0.125 g of the intestinal rat α -glucosidase enzyme with 5 mL of a cold phosphate buffer pH 7.0, sonicated and centrifuged at 5000 rpm, 40°C for 5-10 minutes, and then the supernatant was collected into a disposable plastic tube. Enzyme stock was diluted 2.5 times using a phosphate buffer pH 7.0. The tested samples were prepared by dissolving 20 mg of crude extract/fraction with 200 uL of DMSO homogeneously and diluting with phosphate buffer pH 7.0 to the final concentration of 100 ppm. The measurement of alpha-glucosidase inhibition activity was carried out as follow. In the disposable plastic cuvette, 70 μ L each sample solution, 100 μ L the p-nitrophenyl-alpha-D-glucopyranoside 10 mM, and 80 µL phosphate buffer solution pH 7.0 were mixed together and incubated for 5 minutes at 37°C. The reaction was then started by adding 100 µL the enzyme working solution and re-incubated for 15 minutes at 37°C. The reaction was stopped by adding 400 µL 0.2 M Na₂CO₃, and the absorbance was measured at 400 nm using a UV-Vis spectrophotometer. Sample blank was prepared by replacing the enzyme solution with phosphate buffer pH 7.0. Enzyme control, which states an enzyme without inhibition, was made by replacing the test sample with phosphate buffer pH 7.0. Phosphate buffer pH 7.0 was as an enzyme control blank. As positive control was used acarbose. Measurements were carried out triplicate. Percentage of inhibition was calculated by the equation as follow.

$$\% Inhibition = \left[1 - \frac{[Abs \ sample \ -Abs \ sample \ blank]}{[Abs \ control \ -Abs \ control \ blank]}\right] \times 100\%$$

Determination of IC₅₀ value was conducted with the similar procedure stated above using a range concentration of each sample from 250-2000 ppm. IC₅₀ was derived from the regression curve by plotting each final concentration and % inhibition of each sample using the *Microsoft Excel* program. Experiments were carried out triplicate. The type of inhibition was determined by measuring the percent inhibition of a range concentrations of the tested sample toward a series concentration substrate of p-nitrophenyl-alpha-D-glucopyranoside. A Lineweaver-Burk curve which stated the relationship between 1/V to 1/[S] (substrate concentration) was plotted and the type of inhibition was derived from the intersection between curves obtained.

Total phenolic content determination

Total phenolic compounds were determined using gallic acid as positive control based on the previous study¹² with minor modifications. Briefly, twenty mg gallic acid was dissolved in 20 ml of methanol and then diluted to make a serial concentration of working solutions (25, 50, 100, 150 and 200 ppm) using methanol. Into the test tube, 200 µL of each working solution was added with 750 µL of Folin-Ciocalteau reagent, gently shaken until homogenous and then incubated at room temperature for 5 minutes. To the reaction, 750 µL of 6% sodium carbonate solution was added, gently stirred and incubated in the same way for 90 minutes. Furthermore, absorbance was measured by UV-Vis spectrophotometer at 725 nm. Methanol was used as a blank solution. Measurements were performed in triplicate. A calibration curve was made by extrapolating absorbance against to final concentration and the regression equation, y=ax+b, was calculated using the Microsoft Excel program. Total phenolic content of the tested samples were measured by dissolving 20 mg of each sample in 200 μL dimetyl sulfoxide and then diluted with methanol until the final concentration 100 ppm obtained. Each sample solution was carried out as positive control stated above. Total phenolic

content was measured using the regression equation, y=ax+b, of gallic acid and stated as gallic acid equivalent (GAE). Percent of total phenolic content was calculated using this equation.

% Total phenolic content = $\left[\frac{GAE \ (ppm)}{final \ concentration \ (ppm)}\right] x \ 100\%$

GC-MS analysis

Ethyl acetate fractions of *Ardisia humilis* Vahl. leaves were analyzed by GC-MS technique for the detection of the active components present in the extract. GC analysis was conducted using a GC-MS (Agilent Technologies 7890) equipped with auto-injector and an HP Ultra 2 Capillary Column ((5%-phenyl)-methylpolysiloxane) of 0.20 mm diameter, 30 m length, and 0.11 µm film thickness. Ten mg of the semisolid sample were dissolved in 1 mL of methanol solvent, sonicated and centrifuged. The sample size of 5 µl was injected through the injector. The inert gas helium was used as the carrier gas. Moreover, the MS chromatogram was taken at 70 eV of ionization energy with 1.2 mL/minute column flow. The column mode used was constant flow. The initial temperature of the oven was 80°C hold for 0 minutes, rising at 3°C/min to 150°C hold for 1 minute and finally rising 20°C/min to 280°C hold for 26 minutes. The relative percent amount of each component was expressed as a percentage with the peak area.

Data analysis

Data obtained from these experiments were presented as mean \pm SD. Statistical analysis was conducted using the ANAVA (parametric data) or Kruskal-Wallis (non-parametric data) method and followed by the LSD (least square deference) for parametric data or Mann Whitney method non-parametric data to determine the further differences between samples. The analysis was carried out with SPSS 11 program at

a 95% confidence level (p = 0.05). P value < 0.05 was considered to be statistically significant.

RESULTS

Sample preparation

The experimental scheme and the sixth months of age Lampeni that planted in two different conditions were showed in Figures 1 and 2. The tested samples were prepared with the reflux method followed by liquid-liquid partition of crude extracts until some fractions obtained. The yield of all crude extracts and fractions of Lampeni was presented in Table 1. Crude extracts still contain various types of compounds ranging from non-polar to polar. To separate compounds based on their level of polarity a liquid-liquid partition using a type of solvent was carried out with different levels of polarity. The partition was began by separating the non-polar, semipolar and finally polar compounds using n-hexane, ethyl acetate, and water, respectively.

Alpha-glucosidase inhibitory evaluation in vitro

The inhibitory activities of each tested sample toward alpha glucosidase enzyme were presented in Figure 3. The value showed was an average of triplicate test results which conducted at 100 ppm final concentration. The result displayed that ethyl acetate fraction (Fr. EtOAc) harvested at 4 and 6 months age demonstrated the highest activity. However, their activities were significantly different to positive control acarbose (p < 0.05). Both activity Fr. EtOAc of 4 and 6 months were almost the same (p = 0.056).

Figure 3 demonstrated that the harvesting age influenced alphaglucosidase inhibition activity in which the older the plant age, the higher the inhibition activity was demonstrated. Sun exposure also

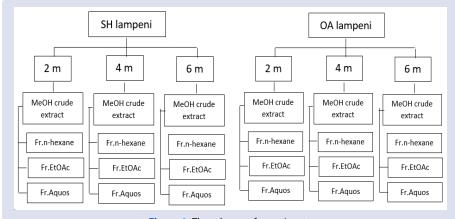


Figure 1: The scheme of experiment.

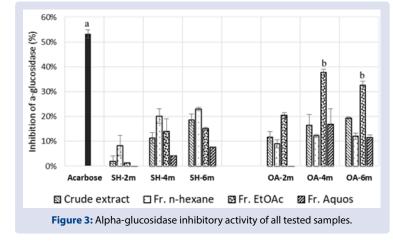
Evaluation of alpha glucosidase inhibitory activity and total phenolic determination



Figure 2: The sixth months of age Lampeni (*Ardisia humilis* Vahl.) at shedding house (left) and open air (right) location.

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Location	The age of plant	Extraction yield (g extract/ 100 g leaf powder)					
		Crude extract	Fr. n-hexane	Fr. EtOAc	Fr. Aquos		
Shedding House	2m	10.38	1.42	0.66	9.34		
	4m	15.13	1.30	1.58	12.63		
	6m	9.71	1.93	1.54	7.46		
Open air	2m	13.48	1.14	1.24	15.38		
	4m	20.67	1.35	0.88	14.34		
	6m	18.47	0.48	1.27	14.29		



contributed to the inhibitory activity. Plants which placed in the open-air provided better activities than those one maintained in shedding house. It was an interesting phenomenon that the activity of each fraction from the two planting conditions showed a different pattern. On the plants that cultivated in shedding house, the n-hexane fraction showed higher activity than the other two fractions. While on plant cultivated direct to sun exposure, the highest activity was shown in EtOAc fractions, especially from 4 and 6 months harvested leaves. The inhibitory strength of both EtOAc fraction was significantly higher than the other fractions. However, when they were compared to control acarbose, it was still significantly lower (p <0.05). The IC₅₀ values for the two most active fraction were presented in Table 2.

Table 2 showed that the EtOAc fraction provided a higher resistance than the other fractions, especially for plants harvested at the age of 4 and 6 months. IC_{50} values of both EtOAc fractions were 93.50 ppm (Fr EtOAc OA-4m) and 98.13 ppm (Fr EtOAc OA-6m), respectively, and those both values were not statistically significantly different (*p*>0.05). The inhibition type of the two most active samples (Fr EtOAc OA-4m) and Fr EtOAc OA-6m) was presented in Figure 4.

Figure 4 displayed the kinetic study toward alpha glucosidase inhibition of 2 samples that had the highest activity, in which there was a speed decreasing when the inhibitor (samples) concentrations were increased. It also showed that the Km value remained the same even though the inhibitor concentrations were added. According to the inhibition type criteria, the curve as Figure 4 was non-competitive category. Km is the Michaelis Menten constant which expresses the affinity of the enzyme to the substrate. The value of Km is shown by the intersection between the curve and the x-axis.¹³ The each Km value of the two EtOAc fractions were almost the same at the range between 0.3-0.4. This means that the possibility of chemical compounds composition that provided alpha-glucosidase inhibition from both fractions originating from different the age of harvest ages was almost similar. In non-competitive type inhibitions, inhibitors (chemical compounds in the fraction) bound to either E (enzyme) or ES (enzyme-substrate) complex and produced EI

(enzyme-inhibitor) and ESI (enzyme-substrate-inhibitor) complexes. The compounds contained in the Fr.EtOAc, in these studies, may not bind at the same side as the substrate, this was possibility due to the structure of the compounds might possess no resemblance to the substrate structure. Complexes that are formed either EI or ESI cause the changing of enzyme conformation and then the catalytic reaction can not work perfectly.^{13,14}

Total phenolic content determination

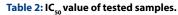
The results of total phenolic levels measurement were shown in Figure 5. Total phenolic content was the average from 3 times measurements expressed as mg galic acid equivalent (GAE) /final concentration x 100%. Total phenolic was determined using Folin-Ciocalteau reagent spectrophotometerically at 725 nm.

Figure 5 demonstrated that the levels of total phenolic compounds in methanol crude extract did not differ significantly between the age of planting (2, 4, and 6 months) and between planting conditions (openair and shedding house). However, in the fractioned samples, EtOAc fraction of each planting condition showed the highest total phenol compound level compared to the other fractions, especially in the leaves that harvested at the age of 4-OA and 6-OA months. Statistically, all of EtOAc fractions did not differ significantly (p=0.118).

GC-MS analysis

In this study, all of EtOAc fractions of *Ardisia humilis* Vahl. leaves planted in the open area (OA) and shedding house (SH) conditions were analyzed using GC-MS (the spectra of each EtOAc fraction were presented in Figure 6). GC-MS analysis was conducted with the HP ultra 2 capillary that had non-polar properties. Based on the chromatogram, it showed that the most major peaks chromatogram appeared in the middle area. It indicated that the majority of the dominant compounds contained in Fr.EtOAc of Lampeni leaves were semi-polar. After being matched with the database, pyrogallol (38.52 % with RT 17.8, Figure 7)

Fr. EtOAc OA-4m			Fr. EtOAc OA-6m		
Final concentration	Inhibition	IC ₅₀	Final concentration	Inhibition	IC ₅₀ (ppm)
(ppm)	(%)	(ppm)	(ppm)	(%)	-50 (1-1
186.67	97.4%		186.67	99.6%	
140.00	71.1%		140.00	74.3%	
93.33	39.3%	93.50	93.33	37.2%	98.13
46.67	31.1%		46.67	28.4%	
23.33	21.7%		23.33	9.4%	



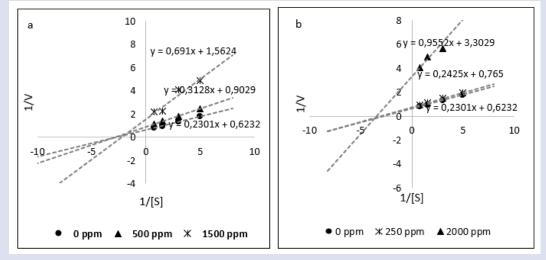
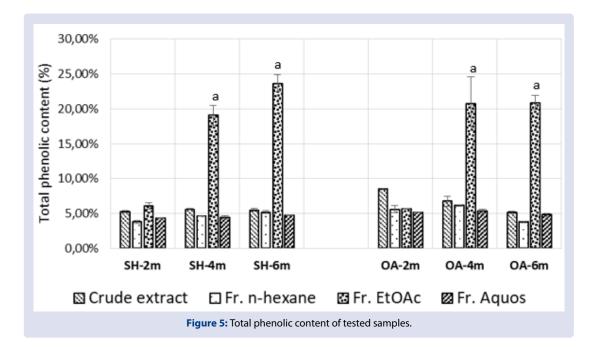


Figure 4: Lineweaver-Burk plot curve (a) Fr. EtOAc OA-4m (b) Fr. EtOAc OA-6m.



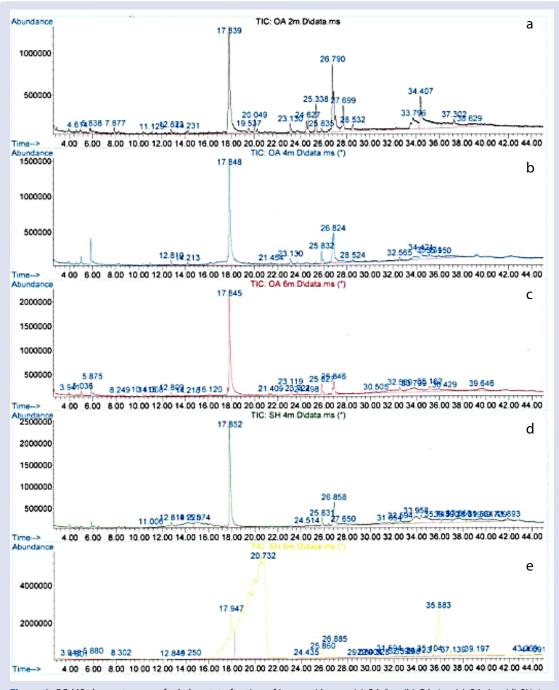
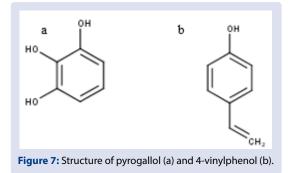


Figure 6: GC-MS chromatogram of ethyl acetate fraction of Lampeni leaves. (a) OA-2m; (b) OA-4m; (c) OA-6m; (d) SH-4m; (e) SH-6m.



could be categorized as the major compound in which the compound appeared with the biggest total area in all tested fraction. Additionally, some peaks of the OA-4m and OA-6m fraction with RT 4 and 8 minutes appeared more abundance. One of the most apparent peak of both fractions was 4-vinylphenol (4.13 % with RT 5.876) (Figure 7).

DISCUSSION

Glucose in the body can be categorized into two, namely exogenous glucose (derived from food intake) and endogenous glucose, which derived dominantly from the liver with glycogenolysis (conversion of glycogen to glucose) and gluconeogenesis (glucose formation), which about ~15% is produced by the kidneys. The imbalance of production and usage of glucose could be the cause of hyperglycemia.¹⁵ Intake of starch-rich foods (a chain glucose molecule) is one of the leading causes of the postprandial glucose increase. Starch hydrolysis process to produce glucose monomer compounds which can be absorbed from gastrointestinal tract involves various enzymes ranging from ptyalin in the oral cavity to hydrolytic enzymes such as alpha-amylase and alphaglucosidase.¹⁶ These enzymes that contributed to hydrolyzing amylum to be oligo and monosaccharide form are called amylolytic enzymes.¹⁷ Monosaccharides absorbed will be transported through the portal vessels to the liver. The efforts to inhibit these enzymes will be beneficial in suppressing postprandial glucose which plays a main role in DM type 2 treatment.¹⁸ Postprandial hyperglycemia is an elevation of blood glucose level after a meal. There are some evidences that the chronically postprandial hyperglycemia associated with macro and microvascular complication through a complex oxidative stress generation, vascular inflammation and platelet activation pathways.19

Some plants demonstrated the inhibitory activity in the starch metabolizing enzyme.³ This activity was caused by the content of chemical compounds such as polyphenol or phenolic compounds in which the concentrations were influenced by several agronomic factors such as the place to grow, the way and age of harvest, and post-harvest processing.¹⁰ The inhibitory alpha-glucosidase activity of these *Ardisia humilis* extracts shown in this study were in accordance with the previous studies. It had been reported previously that the alpha-amyrin compound isolated from 70% methanolic extract of *Ardisia elliptica* leaves exhibited activity in decreasing blood glucose level.²⁰ The other research found that alpha-amyrin decreased blood glucose about 17% at 24 h after sucrose induction in STZ-induced diabetic rat that induced by sucrose at dose 100 mg/kg bw per oral.⁶

The agricultural study states that the quality and quantity of compounds in the plant-derived extract are significantly affected by many factors such as climate, seasons of the year, phenological stage, genetic load, temperature, altitude, cultivation conditions and humidity.²¹ The secondary metabolite compounds determine the strength and type of biological activity of a natural product. The research presented that the levels of phenolic compounds were influenced by planting conditions and the age of harvest. The leaves harvested at the age of 2 months were not fully grown, so the secondary metabolites were not completely produced. It gave the consequence that the inhibition strength was the lowest due to the total phenol content too low. The older the plant, the stronger the alpha glucosidase inhibitory activity was produced. The inhibition strength of alpha-glucosidase enzyme was proportionate with the total phenolic content of the extract in which it was influenced by some factors such as the age of the plant and the method of planting. This result was in accordance with the previous study that conducted in Yerba mate⁸ in which the antioxidant activity and total phenolic content were significantly affected by leaves age. The other study²² also demonstrated that there was a correlation between the phenolic compound level and antidiabetic activity such as in Sansevieria cylindrica extract, a plant that traditionally used for the treatment of various ailments in African countries. Phenolic compounds contained in plants had been recognized to have some pharmacological activities, such as antimicrobial, antioxidant, antitumor, and antidiabetic activities.²³ This result was supported by GC-MS analysis, in which after being matched with database, it showed that the phenolic compound pyrogallol was found as the most dominant compound in all tested fractions. Additionally, there were several compounds found only in ethyl acetate fractions (OA-4m and OA-6m fraction) with high inhibitory activity, one of which was 4-vinyl phenol. The presence of this compound may cause both the OA-4m and OA-6m EtOAc fractions demonstrated the highest alpha-glucosidase inhibitory activity besides others which previously reported.

CONCLUSION

The effect of some agronomic variable (the light intensity and the age of harvesting) and extract preparation of *Ardisia humilis* Vahl. (Lampeni) toward the inhibitory activity of alpha-glucosidase enzyme and polyphenol content had been investigated. The results showed that the plant cultivated under direct sun exposure harvested at the age of 4-6 months and prepared as ethyl acetate fraction gave the highest alpha-glucosidase inhibition activity although there was no positive correlation between the strength of inhibition and total phenolic content.

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AUTHORS CONTRIBUTION

Sri Ningsih (SN). Fifit Juniarti (FJ). Idah Rosidah (IR). Syofi Rosmalawati (SR). Agung Eru Wibowo (AEW). Adam Arditya Fajriawan (AAF). Kurnia Agustini (KA). Wahono Sumaryono (WS). Erliana Sasikirana (ES). SN, AEW, and WS conceived and planned the experiments. SN, FJ, IR, SR, KA, ES carried out the experiments. SN and ES analyzed data. SN, ES and WS contributed to the interpretation of the results. SN and AAF prepared, corrected and submited the manuscript.

CONFLICTS OF INTEREST

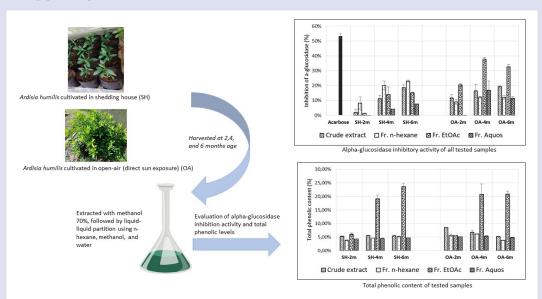
No potential conflicts of interest were reported by the authors.

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