

In vitro Assay of Alpha-Glucosidase Inhibitor Activities of Three Seagrasses from Banten Bay, Indonesia

Agung Widiyanto¹, Effionora Anwar^{1*}, Tati Nurhayati²

ABSTRACT

Introduction: Diabetes mellitus (DM) is an endocrine disorder characterized by high blood glucose levels. One of the approaches for treating DM is by regulating the postprandial blood glucose via inhibition of α -glucosidase enzymes. Seagrass is known as a plant containing bioactive compounds, especially for therapy antidiabetic. This research aims to evaluate the α -glucosidase inhibition activity of three species seagrasses (*Enhalus acoroides*, *Thalassia hemprichii* and *Cymodocea rotundata*) from the Banten Bay, Indonesia. **Methods:** The extracts of all parts (root, leaf, and stem) from three seagrasses were prepared with maceration method using 80% ethanol solvents. The α -glucosidase inhibitory activity was conducted by microplate reader at 400 nm using acarbose as a positive control. Furthermore, the kinetics of α -glucosidase inhibition, phytochemical screening, and total phenolics content were evaluated against extract which has the most potential α -glucosidase inhibition. **Results:** The result showed that the 80% ethanol extracts of *Enhalus acoroides* (IC₅₀ values 168.15 ± 2.71 μ g/mL) had the most potential α -glucosidase inhibitors activity compared with the positive control acarbose, *Thalassia hemprichii* and *Cymodocea rotundata* (IC₅₀ values 197.27 ± 3.07 μ g/mL, 425.86 ± 5.15 μ g/mL and 429.28 ± 8.89 μ g/mL). The kinetic type of inhibition against α -glucosidase was noncompetitive inhibition. The phytochemical compounds were phenols, flavonoid, terpenes, and tannin with the total phenolic content was 28.76 ± 2.46 mgGAE/g. **Conclusion:** *Enhalus acoroides* has the strongest inhibitor of α -glucosidase and can be further developed for DM therapy agents.

Key words: *Cymodocea rotundata*, Diabetes mellitus, *Enhalus acoroides*, Phytochemical compound, *Thalassia hemprichii*.

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INTRODUCTION

Handling of diabetes mellitus can be done through inhibition of the α -glucosidase enzyme that catalyzes the final step of starch digestion and abundant disaccharides in the human diet on the border of the small intestine mucosal brush.¹⁻² Seagrasses are marine angiosperms growing abundantly in the sea that produce secondary metabolites as an antidiabetic,³ antioxidant³⁻⁴ and antibacterial⁵⁻⁶ activities with the chemical compound such as phenol, flavonoids, tannin, terpene, sterol.⁶⁻⁷ The aim of the present work is to investigate the α -glucosidase inhibitory activities of three seagrasses in Banten Bay, Indonesia i.e. *Enhalus acoroides*, *Thalassia hemprichii* and *Cymodocea rotundata* extracts.

MATERIALS AND METHODS

Materials

The plant parts used in this study were all plant parts (leaves, stems, and roots) collected in October 2017 from the Bay of Banten, Indonesia and identified by the Department of Biology, FMIPA University of Indonesia.

The chemical materials such as an α -glucosidase enzyme which was obtained from *Saccharomyces cerevisiae* (Sigma Aldrich, USA), *p*-nitrophenyl α -D-glucopyranoside (pNPG) (Sigma Aldrich, USA), Acarbose (Huadong Medicine, China), Quercetin (Sigma Aldrich, German), bovine serum albumin (Merck, German). All reagents used in this research were analytical grade.

Preparation of the extract

All parts of the seagrass were washed with fresh water to remove epiphytes and other impurities. The sample was dried for 5 days in the shade at room temperature. The dried sample was homogenized to obtain the fine powder.

Extraction

Each dried powdered of seagrasses (100 g) were homogenized by Ultraturrax with a rotation speed 15.000 rpm for 10 min and extracted by maceration for 2 days with 80% ethanol then evaporated.

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The preliminary test of α -glucosidase

The preliminary test performed was to determine the maximum wavelength of pNPG, optimum incubation time and substrate concentration using microplate reader.

The α -glucosidase inhibition activity assay

The α -glucosidase inhibition of the extracts was assayed according to the published method with modification.² One mg of α -glucosidase was dissolved in 10 mL of phosphate buffer (pH 6.8) containing bovine serum albumin (0.2% w/v). Sample (10 μ L) was mixed with 66 μ L phosphate buffer (pH 6.8) and 17 μ L of 5mM pNPG then preincubated at 37°C for 5 min. 7 μ L α -glucosidase (0.07 unit/mL) was added then incubated at 37°C for 15 min.

Then 100 μ L Na₂CO₃ 200 mM was added to stop the reaction. The α -glucosidase activity was read at 400 nm in the microplate reader by measuring p-nitrophenol released from pNPG. As a positive control of the α -glucosidase inhibitor used acarbose. An activity of α -glucosidase standard and extract could be defined in % inhibition which obtained using the following formula:

$$\% \text{ inhibition} = \frac{(A - B) - (C - D)}{A - B} \times 100$$

A = blank solution absorption

B = blank control solution absorption

C = sample solution absorption

D = sample control solution absorption

The inhibitory concentrations of standard and extracts required to inhibit the action of α -glucosidase by 50% (IC₅₀) were calculated by linear regression.

Type of inhibition α -glucosidase

Inhibition types analysis were conducted only for the extract that has the best α -glucosidase inhibition activity. This analysis measured a correlation between the concentration of pNPG as a substrate in the existence or absence of extract at different concentrations. The type of inhibition was described by the Lineweaver-Burk data plot analysis, which was calculated from the Michaelis-Menten kinetics results.

Phytochemical screening

The extract which has the best α -glucosidase inhibition activity were evaluated for its bioactive compounds by Thin Layer Chromatography (TLC). The developed TLC plates were air dried and observed under ultraviolet light at 254 nm and 366 nm. Then *Retention factor* (Rf) was calculated by dividing the distance of compound by the distance of solvent migration from the original position.

Total phenolic content

The total phenolic content of the extracts was measured by Folin-Ciocalteu method according to Vanitha et al. (2017)⁷ with modification. An aliquot of the extract (0.5 mL) was mixed with 1 mL of Folin-Ciocalteu reagent (10% v/v). The tubes were vortexed for 20 sec and allowed to stand for 5 min. Two mL of sodium carbonate (7.5% w/v) was added. The tubes were vortexed again for 20 sec and allowed to stand for 75 min. Absorbance was then measured at 750 nm using the spectrophotometer UV-Vis. The total phenolic content was expressed as mg/g Gallic acid equivalent.

RESULTS

Extraction

The percentages of each extract yield were shown below in Table 1.

Table 1: % yield extract value.

| Sample | Simplicial weight extracted (gram) | Crude extract weight (gram) | Yield (%) |
|----------------------|------------------------------------|-----------------------------|-----------|
| <i>E. acoroides</i> | 100 | 6.48 | 6.48 |
| <i>T. hemprichii</i> | 100 | 7.12 | 7.12 |
| <i>C. rotundata</i> | 100 | 7.09 | 7.09 |

The preliminary test of α -glucosidase activity

The determination of maximum wavelength, incubation time and also substrate optimization was performed in enzyme solutions 0.07 U/mL. Temperature and pH used in the product information from Sigma Aldrich was 37°C and pH 6.8. Then the substrate concentration was optimized to determine the appropriate substrate concentration to react with enzyme unit used. Substrate concentration was said optimum if all active site in the enzyme had been bound to the substrate so that there were no free enzyme which would produce products. In substrate optimization, enzyme unit was used 0.07 U/mL with substrate variations were 1,25; 2,5; 5; 10; 15; 20; and 30 mM. The result showed that the optimum concentration of pNPG was 5 mM. The optimum conditions for the enzyme were 400 nm, 15 min incubation time and 5mM substrate concentration.

α -glucosidase inhibition activity assay

Inhibition α -glucosidase activity of extracts was determined using pNPG as a substrate and compared with acarbose (Figure 1). The IC₅₀ values of the three types of seagrass extract and acarbose as controls have been obtained. The sequence inhibition α -glucosidase activity from the strong potential to the weak is the extracts of *Enhalus acoroides* (168.15 \pm 2.71 μ g/mL), acarbose (197.27 \pm 3.07 μ g/mL), *Thalassia hemprichii* (425.86 \pm 5.15 μ g/mL) and *Cymodocea rotundata* (429.28 \pm 8.89 μ g/mL). The type of inhibition of 80% ethanol extract of *Enhalus acoroides* was investigated and showed a noncompetitive inhibition (Figure 2).

Phytochemical screening

The 80% ethanol extract from *Enhalus acoroides* was identified with TLC showing the presence of phenol, flavonoid, terpene and tannin compounds (Table 2).⁹

Total phenolic content

The total phenols were expressed as μ g/mL gallic acid equivalent. Standard curve equation was used: $y = 0,00037x + 0,135$, R²=0,995, where y is the absorbance value at 750 nm. The total phenolic content of the 80% ethanol extract *Enhalus acoroides* was 28.76 \pm 2.46 mgGAE/g.

DISCUSSION

The 80% ethanol extract *Enhalus acoroides* had the highest α -glucosidase inhibition activity compare with *Thalassia hemprichii* and *Cymodocea rotundata* and more potential than acarbose. The α -glucosidase inhibition activity of the three types of seagrass extracts is more likely due to more than one compounds containing in the extracts such as phenolic and terpenoid compounds in each extract. Their chemical compounds may have a synergic effect to inhibit the enzyme α -glucosidase.⁸ The type of enzyme inhibition mechanism of 80% ethanol extract *Enhalus acoroides* was a noncompetitive inhibitor. Noncompetitive inhibition may have been due to the chemical compound extract as an inhibitor binds to the enzyme at a location other than the active site.² The phytochemical compound of the extract is phenols, flavonoid, tannin and terpene. While the total phenolic content of the 80% ethanol extract *Enhalus acoroides* was 28.76 \pm 2.46 mgGAE/g. Phenolics compounds are reported as anti-oxidant, anti-inflammatory, antidiabetic etc.^{3,5,10} Phenolic compounds

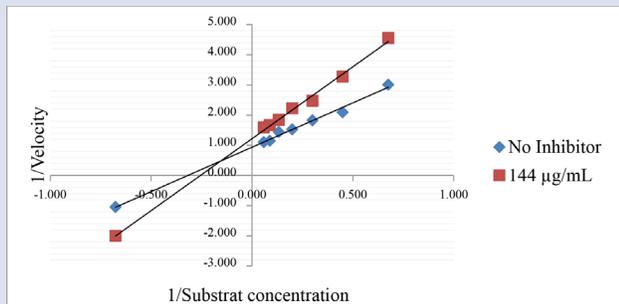


Figure 2: Lineweaver-Burk plot of 80% ethanol extract of *Enhalus acoroides* with concentration of 144 µg/mL.

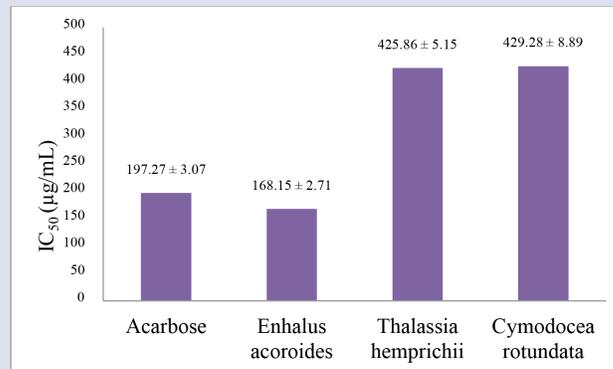


Figure 1: IC₅₀ value standard and extract against α-glucosidase.

Table 2: Phytochemical Analysis of 80% ethanol extract from *Enhalus acoroides* by Thin Layer Chromatography.

| Chemical name | Solvent system | Rf values | Detection/ Spray reagent | Color of day light | UV 254 | UV 366 | Colour with spray reagent | Summary |
|---------------|---|------------|--------------------------|--------------------|-----------|-----------|---------------------------|---------|
| Phenol | cloroform: methanol (9:1) | 0.25; 0.38 | FeCl ₃ | yellow | black | black | black | + |
| Terpene | n-hexane: etil acetat (1:1) | 0.31 | Vanilin-sulfic acid | yellow | black | dark blue | purple | + |
| Tannin | methanol: aquadest (6:4) | 0.19; 0.69 | FeCl ₃ | yellow | dark blue | black | black | + |
| Flavonoid | n-butanol:acetid acid: aquadest (4:1:5) | 0.87 | Quercetin standard | yellow | brown | purple | (no spray) | + |

(+) : present.

affect responses that are related to diabetes by inhibiting α-glucosidase enzyme thereby decreasing starch digestion.¹¹⁻¹³

CONCLUSION

The 80% ethanol extract *Enhalus acoroides* (IC₅₀ values :168.15 ± 2.71 µg/mL) had the highest α-glucosidase inhibition activity compare with *Thalassia hemprichii*, and *Cymodocea rotundata* (IC₅₀ values : 425.86 ± 5.15 µg/mL; 429.28 ± 8.89 µg/mL) and more potential than acarbose (IC₅₀ values :197.27 ± 3.07 µg/mL). While the type of enzyme inhibition mechanism of 80% ethanol extract *Enhalus acoroides* was a noncompetitive inhibitor. The phytochemical compound of the extract is phenols, flavonoid, tannin and terpene. The total phenolic content of the 80% ethanol extract *Enhalus acoroides* was 28.76 ± 2.46 mgGAE/g. *Enhalus acoroides* potential inhibits the α-glucosidase enzyme and further research about the isolation of active compounds is possible.

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CONFLICT OF INTEREST

Nil (The author declares no conflict of interest.

ABBREVIATIONS

DM: Diabetes Mellitus; **α-glucosidase:** Alpha-glucosidase; **IC₅₀:** Half Maximal Inhibitory Activity; **TLC:** Thin Layer Chromatography; **pNPG:** p-Nitrophenyl-α-D-glucopyranoside; **GAE:** Gallic Acid Equivalent.

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SUMMARY

- Alpha-glucosidase inhibition activity of 80% ethanol extract of three seagrasses were determined.
- The 80% ethanol extract of *Enhalus acoroides* has the best α -glucosidase inhibition activity.
- IC_{50} value extract of *Enhalus acoroides*: $168.15 \pm 2.71 \mu\text{g/mL}$, *Thalassia hemprichii*: $425.86 \pm 5.15 \mu\text{g/mL}$ and *Cymodocea rotundata*: $429.28 \pm 8.89 \mu\text{g/mL}$.
- Inhibition types analysis, Phytochemical compounds, and total phenolic content were conducted only for *Enhalus acoroides*.
- The type of enzyme inhibition mechanism of 80% ethanol extract *Enhalus acoroides* was a noncompetitive inhibitor.
- Phytochemical compounds of 80% ethanol extract *Enhalus acoroides* i.e. phenols, flavonoid, tannin, and terpene were analyzed using Thin Layer Chromatography.
- The total phenolic content of the 80% ethanol extract *Enhalus acoroides* was $28.76 \pm 2.46 \text{ mgGAE/g}$.

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