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Anti-Nociceptive Synergism of Pregabalin and Xylopic acid Co-administration in Paclitaxel-induced Neuropathy: Isobolographic Analysis

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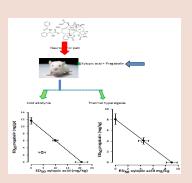
ABSTRACT

Background: Paclitaxel-induced neuropathy still remains a clinical problem for patients undergoing chemotherapy. Objective: The objective of the study was to determine the interaction between xylopic acid and pregabalinco-administration as well as their toxicity in paclitaxel-induced neuropathy. Materials and Methods: Neuropathic pain was induced in rats with 2 mg/ kg of paclitaxel on four alternate days (days 0, 2, 4 and 6). Equi effective doses of xylopic acid and pregabalin that produced 50% anti-nociception (ED₅₀) were determined from their log-dose response curves in the cold allodynia and thermal pain tests. Xylopic acid and pregabalin were again administered to rats in a fixed ratio combination (1:1) of their ED₅₀'s in order to determine the experimental $ED_{50}(Z_{exp})$ of the co-administered compounds. Isobolograms were constructed to compare the $Z_{\rm exp}$ to a theoretical ${\rm ED}_{\rm 50}$ (Z_{add}). Blood samples from the various treatment groups of rats were collected for toxicological assessment of the co-administered compounds. **Results:** The Z_{ava} lay below the Z_{add} on the isobologram of the cold allodnia test. The co-administration exhibited additivity in the thermal pain test. The co-administration did not produce significant (p>0.05) toxicity in rats. The co-administration may be beneficial in paclitaxel-induced neuropathy.

Key words: Isobolograms, Neuropathic pain, Paclitaxel, xylopic acid, Toxicity.

SUMMARY

- Paclitaxel-induced neuropathy is still a clinical problem.
- Pregabalin and Xylopic acid co-administration produced anti-nociceptive synergism against this pain type.
- The co-administration was fairly safe in animals.



PICTORIAL ABSTRACT

Abbreviations used: XA: Xylopic acid, PG: Pregabalin, Z_{add} : Theoritical ED_{50} , Z_{exp} : Experimental ED_{50} .

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INTRODUCTION

Pain has been associated with a wide range of diseases such as cancer and arthritis. Pain can sometimes be the disease itself seen in conditions of neuropathy. Recently, it is reported that more than 1.5 billion people worldwide suffer from chronic pain. Again, approximately 3-4.5% of the global population suffers from neuropathic pain, with incidence rate increasing in complementary to age.2 Neuropathic pain arising from the cancer chemotherapeutic agent, paclitaxel still remains a clinical problem for patients undergoing chemotherapy.³ Paclitaxel is one of the most effective and commonly used anticancer drugs for the treatment of solid tumours. Paclitaxel produces neuropathy by destroying the nerves via its binding to β -tubulin of microtubules. This impairs axoplasmic transport leading to a progressive dying-back axonopathy, hence the neuropathy. Evidence of swollen and vacuolated mitochondria in peripheral sensory axons, both C-fibers and myelinated axons has been reported.4 Its use is limited by Myelossupression and peripheral neurotoxicity.3 Clinically, the sensory neurotoxicity produced by paclitaxel include allodynia, numbness, tingling, and burning pain which often begin symmetrically in the feet and sometimes appear simultaneously in both hands and

feet. The neuropathic pain sometimes becomes a chronic problem even after cessation of paclitaxel administration.⁵ Despite this problem confronted by patients, there is currently no valid drug to prevent or treat the paclitaxel-induced pain.⁶ Pregabalin is the only drug currently approved in the USA for the treatment of spinal neuropathic pain, a type of pain condition affecting 40% of spinal cord injury patients.⁷ There is the likelihood for food-drug interaction occurring between xylopic acid and pregabalin especially in Africa. Patients prescribed with pregabalin for their disease conditions may ingest xylopic acid which is present in the fruits of *Xylopia aethiopica*. *Xylopia aethiopica* is consumed as a delicacy in most parts of the world.8 This presents important drug-food interaction because xylopic acid is the highest occurring compound in the fruits of X. aethiopica.9 Xylopic acid has demonstrated antinociceptive property in several models of pain. The relief of pain by xylopic acid has been attributed to its action on the opioidergic, adrenergic, bradykinin and prostaglandin nociceptive pathways. 10-12 Pregabalin, an antiepileptic drug has demonstrated analgesic property in paclitaxel-induced neuropathic pain in pre-clinical, clinical research and therapy.¹³

To improve the analgesic potency and efficacy of xylopic acid and pregabalin against paclitaxel-induced neuropathy in rats, isobolographic study was performed on these two agents. Because the effects of co-administered drugs are often highly variable, such variability necessitates the use of statistical robust methodology. Isobolographic study is a tool that provides unambiguous terminologies of drug interactions and a statistical framework for the analysis of drug combinations or co-administration. Drug combination regimens have been utilised in areas of HIV-AIDS, cancer and tuberculosis with the aim of achieving a therapeutic efficacy greater than that achieved with monotherapy with some success. Other benefits include decreased toxicity—antagonistic interaction, the delay or prevention of drug resistance development, and the favourable effects of synergistic drug interactions. Synergism is especially important in clinical situations with drugs because it allows the use of smaller amounts of the constituent drugs. An adverse effect may also synergise, presenting a phenomenon of special importance in clinical situations which often leads to reduction of doses of the individual drugs for therapy. The detection of synergism may also be useful in illuminating mechanism of drug action and in the development of new theories. 14,15 It is therefore possible to upsurge the analgesic efficacy and potency of xylopic acid and pregabalin and correspondingly decrease their side effects via combination therapy.

METHODS AND MATERIALS

Extraction of xylopic acid (15 β -Acetoxy(-)-kaur-16-en-19-oic acid)

Xylopic acid was extracted according to the process described elsewhere. 9,10 Briefly, 360 g of dried fruit of *Xylopia aethiopica* was macerated with 5 L of petroleum ether (40–60°C). This was and allowed to stand for three days. The petroleum ether was drained and concentrated with rotary evaporator at a temperature of 50°C. Crude xylopic acid formed after three days was washed with petroleum ether at 40–60°C and purified with 96% ethanol. The yield of the xylopic acid was 1.41%, melting point of 261-262°C. The purity of the extracted xylopic acid as determined with High Performance Liquid Chromatography was 95% as described previously. 9

Animals and husbandry

Sprague-Dawley rats (150–200 g) of both sexes were housed in the animal facility of the Department of Biomedical and Forensic Sciences. The animals were housed in groups of six in stainless steel cages (34×47×18 cm) with soft wood shavings as bedding. They were fed with normal commercial pellet diet (AGRICARE, Kumasi, Ghana), given water *ad libitum* and maintained under standard laboratory conditions. All procedures and techniques used in these studies were in accordance with the National Institute of Health Guidelines for the Care and Use of Laboratory Animals. All protocols used were approved by the Departmental Ethics Committee.

Drugs and reagents

Pregabalin (Lyrica®) was purchased from Pfizer Pharmaceuticals, Arzneimittelwerk Godecke, Freiburg, Germany, cremophor from Sigma-Aldrich Inc., St. Louis, MO, USA and paclitaxel (Intaxel®) from Fresinius Kabi Oncology, Badi, India.

Paclitaxel administration

Baseline measurements of thermal and cold stimuli were performed after allowing the rats to acclimatise to the behavioural testing environment. Neuropathic pain was induced in the rats by intraperitoneal (i.p.) injection of paclitaxel (2 mg/kg) dissolved in saline on four alternate days (days 0, 2, 4 and 6) as described. On day 16 post-paclitaxel administra-

tion, the various groups of animals were treated with vehicle (cremophore solution) xylopic acid (10-100 mg/kg in cremophore) and pregabalin (10-100 mg/kg) for five days.

Cold allodynia

The analgesic effect of orally co-administered xylopic acid and pregabalin on cold allodynia was assessed by immersing the rat's hind paw into cold water (4.5°C). The latency for a rat to withdrawal its paw was measured with a digital timer.¹⁷ Just one hind paw was assessed during each immersion at a time with a cut-off time of 20 s. For each animal, two recordings were made for each hind paw, and the withdrawal responses were reported as the mean of both hind paw values. The treatment was continued with vehicle, xylopic acid and pregabalin for 5 days.

Thermal Hyperalgesia

The tail immersion test was done by immersing the distal portion of the tail (3-4 cm) of the various treated groups of rats in hot water maintained at 52°C until the tail was withdrawn. The duration of immersion was recorded and a cut-off time of 10 s was used. ¹⁸ The treatment was continued with vehicle, xylopic acid and pregabalin for 5 days.

Isobolographic analysis of analgesic property and toxicological assessment of xylopic acid and pregabalin co-administration

The potencies of xylopic acid and pregabalin estimated previously in the cold allodynia and thermal hyperalgesia tests were used for the isobolographic studies. The estimated potencies (ED_{50} 's) of xylopic acid and pregabalin in both tests were also used to compute the theoretical potency (Zadd) as follows;

To obtain the combination potency of co-administered xylopic acid

 $Zadd = f(ED_{50})$ of pregabalin + (1-f) ED_{50} of xylopic acid Where f is the fraction of the each component in the mixture. ¹⁵

(XA) and pregabalin (PG), the two agents were orally administered to paclitaxel-induced neuropathic pain suffering rats daily at doses of their respective ED50's and in fixed ratio combinations of fractions of their respective ED₅₀ values (equieffctive doses) of 1/2, 1/4, 1/8, 1/16. The treatment regime continued for 5 days in the cold allodynia and thermal hyperalgesia tests. The experimental potency (Z_{exp}) of the co-administered test agents (XA+PG) was determined by least square method of regression. To determine the drug-drug interaction in the living system, isobologram (a cartesian plot of pairs of doses that, in combination, yield a specified level of effect) was constructed by connecting the theoretical ED₅₀ of xylopic acid plotted on the abscissa and pregabalin plotted on the ordinate to obtain the line of additivity. The ED₅₀ (experimental) was compared to the theoretical additive ED₅₀ using a't'-test. Synergistic analgesic effect is obtained when the effect of a drug combination is higher and statistically different (ED₅₀ significantly lower) than the theoretically calculated equieffective dose of a drug combination with the same proportions. If the ED50's are not statistically different, the effect of the combination is additive which implies each constituent contributes with its own potency to the total effect. The interaction index was also computed as Z_{op}/Z_{old} . If the value is close to one, the interaction is additive. Values lower than one indicates the magnitude of synergistic interactions and values higher than 1 represent antagonistic interactions. Blood samples were also collected for haematology and biochemical assay for signs of toxicity on the fifth day post-treatment. The haematological parameters were determined with an automatic analyzer (Sysmex XT-2000lCELL-DYN 1700, Abbot Diagnostics Division, Abbot Laboratories, Abbot Park, Illinois, USA). Haematological parameters estimated included haemoglobin (Hb), erythrocyte count (RBC), haematocrit (HCT), mean corpuscular volume (MCV), mean corpuscular haemoglobin (MCH), mean corpuscular haemoglobin concentration (MCHC), platelet count

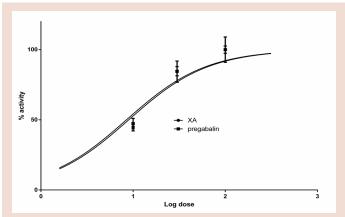


Figure 1: Dose-response curves of (a) xylopic acid and (b) pregabalin in the tail immersion test Data is presented as mean \pm S.E.M

(PLT), and total white blood cells (WBCs). The serum biochemistry parameters studied were alanine aminotransferase (ALT), aspartate aminotransferase (AST), alkaline phosphatase (ALP), gamma glutamyl transferase (GGT), bilirubin, blood urea nitrogen (BUN), creatinine, total protein (protein) and albumin.

Statistical analysis

The data were analysed with GraphPad Prism Version 5 (GraphPad Software, San Diego, CA, USA). The results are presented as mean \pm S.E.M. The difference between the means of co-treated groups and negative control group was analysed with One-way analysis of variance followed by Tukey's post *hoc* test. Doses for 50% of the maximal effect (ED $_{50}$) for each drug were determined using an iterative computer least squares method, with the following nonlinear regression (three-parameter logistic) equation:

$$Y = \frac{a + (b - a)}{\left(1 + 10^{(LogED_{50} - X)}\right)}$$

Where, X is the logarithm of dose and Y is the response. Y starts at a (the bottom) and goes to b (the top) with a sigmoid shape. P<0.05 was considered statistically significant. The fitted midpoints (ED $_{50}$ s) of the curves, i.e., Z $_{\rm exp}$ and Z $_{\rm add}$ were compared statistically using t-test. In Isobolographic calculations were performed with the program Pharm Tools Pro (version 1.27, the Mc Cary Group Inc.). Results are presented as mean \pm S.E.M.

RESULTS

Estimation of the potencies of xylopic acid and pregabalin in the cold allodynia and thermal hyperalgesia tests

The potencies of xylopic acid and pregabalin were 8.97 \pm 0.96 mg/kg and 8.1 \pm 0.85 mg/kg in the thermal pain tests respectively (Figure 1). In the cold allodynia test, the ED $_{50}$ of xylopic acid and pregabalin were 20.64 \pm 0.99 mg/kg and 11.65 \pm 095 mg/kg respectively (Figure 2). The fractions of the ED $_{50}$ of xylopic acid and pregabalin used for the isobolographic studies are shown on Table 1.

Isobolographic assessment of the analgesic effect of xylopic acid and pregabalin co-administration in the cold allodynia and thermal hyperalgesia tests

The co-administration of XA and PG resulted in significant analysis effect compared to the vehicle treated animals. The theoretical ${\rm ED}_{50}$'s in the

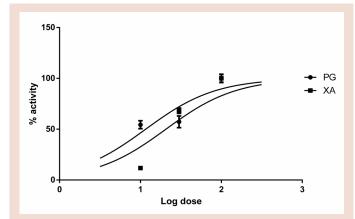


Figure 2: Dose-response curves of xylopic acid and pregabalin in the cold allodynia test
Data is presented as mean ± S.E.M

cold allodynia and thermal hyperalgesia tests were 16.15 \pm 1.2 and 8.54 \pm 1.4 respectively. The experimental ED $_{50}$ (Zexp) for the mixture (XA+PG) in the cold allodynia and thermal pain tests were 7.25 \pm 0.23 and 8.16 \pm 1.6 respectively. The Zexp (open circle) lay significantly below the line of additivity and the Zadd (closed circles) on the isobologram of the cold allodynia but not the thermal hyperalgesia test (Figure 3a and 3b) indicating synergism of analgesic effect (P<0.001) in the cold allodynia test but not the thermal pain test. The degree of interaction calculated as the interaction indices were 0.45 and 0.96 for the cold and thermal tests respectively (Table 2).

Effect of xylopic acid and pregabalin co-administration on haematological parameters and heart, liver and kidney enzymes

The vehicle treated group and XA+PG co-treated groups had reduced but not significant white blood cells concentration compared to sham control groups (Table 3). The liver and bile enzymes, AST, ALT, GGT and ALP from the animals co-administered with various concentrations of XA and PA combination were not statistically different from the negative control groups. Albumin and total protein concentrations were also not different from the negative control group. The ALT, ALP and AST concentrations of the vehicle treated group were slightly elevated but not statistically significant from that of the sham group (Table 3). The kidney

Table 1: The ED $_{\rm so}$'s \pm S.E.M. and fractions of Xylopic acid and Pregabalin used for determining the Z $_{\rm exp}$ in the cold allodynia and thermal hyperalgesia tests

	Doses used (mg/kg)						
Tests	Xylopic acid	Pregabalin					
Thermalpain (Tail immersion test)	8.97 ± 0.96	8.1 ± 0.85					
	4.49	4.05					
	2.24	2.025					
	1.12	1.013					
	0.56	0.506					
Cold allodynia	20.64 ± 0.99	11.65 ± 0.95					
	10.32	5.825					
	5.16	2.913					
	2.85	1.456					
	1.29	0.728					

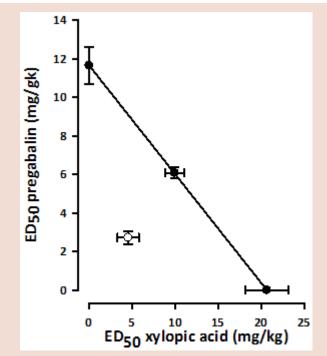


Figure 3a: Isobologram for the combination of xylopic acid and pregabalin in (a) cold allodynia test in rats. Filled circles (●) are the theoretical ED50's ± S.E.M. and open circles (o), the experimental ED50's ± S.E.M

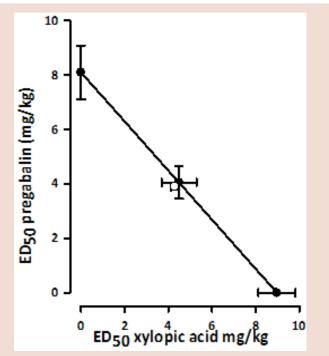


Figure 3b: Isobologram for the combination of xylopic acid and pregabalin in (b) thermal hyperalgesia tests in rats. Filled circles (•) are the theoretical ED50's ± S.E.M. and open circles (o), the experimental ED50's ± S.E.M

Table 2: Theoretical (Z_{add}) and experimental (Z_{exp}) ED₅₀ \pm S.E.M. values of Xylopic acid and Pregabalin co-administration in the cold allodynia and thermal hyperalgesia tests

Tests							
ED ₅₀ 's (XA+PG: 1:1)	Thermal pain (Tail immersion test)	Cold allodynia					
Zadd (mg/kg)	8.54 ± 1.4	16.15 ± 1.2					
Zexp (mg/kg)	8.16 ± 1.6	7.25 ± 0.23***					
Interaction index	0.96	0.45					

Values are presented as Mean \pm S.E.M. ***P<0.001compared to their respective Z_{adds}

enzymes and function marker, creatinine and BUN were insignificantly altered in the XA+PG co-treated groups compared to the vehicle treated group (Table 3). The various co-treatments did not produce toxicity on the erythrocytes and their contents (Table 3).

DISCUSSION

Oral co-administration of pregabalin and xylopic acid produced synergistic analgesic properties and no significant toxic effects in rat model of paclitaxel-induced neuropathic pain. The potency of co-administered agents, xylopic acid and pregabalin was significantly higher, and lay below the line of additivity and the theoretical potency of the two agents in the cold allodynia test. The additivity line represents the level of effect for the individual agents and the closeness of the experimental potency of the mixture to this line in the thermal pain test demonstrates additivity.²⁰ Both compounds were more potent in the thermal pain test compared to the cold allodynia test. On the contrary, the magnitude of potentiation, with reference to the interaction indices was greater in the cold allodynia test than the thermal pain test. The difference in efficacy of the co-administered agents in the two tests may be due to the different pathophysiology associated with the two pain types. It is obvious that the efficacy of the two agents as well as the co-administration vary with respect to type of noxious stimulus.

Paclitaxel, after its administration accumulates in the dorsal root ganglia and the brain at very low concentrations. Accumulation has also been reported in the sciatic nerve and spinal cord at intermediate concentrations.21 Morphological changes such as swollen and vacuolated mitochondria as well as reduced respiration and energy production in axonal mitochondria that results from paclitaxel administration initiates apoptosis pathways by altering intracellular calcium levels.^{4,22} Therefore, the neuropathy in this study after low dose paclitaxel administration was due to atypical (swollen and vacuolated) mitochondria in peripheral sensory axons, both C-fibers and myelinated axons as well as the loss of intraepidermal nerve fibres. Both agents may have acted centrally to produce analgesic synergism. This is supported by the tail immersion test. The tail immersion thermal pain test investigates pain primarily of spinal origin.²³ Pregabalin is a well-established anticonvulsant and analgesic agent that has shown efficacy and dose dependent effects either as monotherapy or in combination with analgesics in relieving pain and related symptoms. It is noteworthy that pregabalin has received Food and Drug Association (FDA) approval for the treatment of diabetic neuropathy and post-herpetic neuralgia. Pregabalin produces its analgesic effect through high-affinity alpha2-delta-1 subunit of voltage-gated calcium channels. Both xylopic acid and pregabalin are calcium channel blockers

Table 3: Effect of Xylopic acid and Pregablin co-administration on blood biochemistry and haematological parameters

D	CI.	NC		Doses of XA and PG (mg/kg)			
Parameter	Sham	NC	XA+PG	XA+PG/2	XA+PG/4	XA+PG/8	XA+P/16
BUN (mg %)	38.4 ± 0.5	39.1 ± 0.2	39.3 ± 0.6	38.9 ± 0.2	39.2 ± 0.5	38.8 ± 0.4	38.5 ± 0.5
Creatinine (mg/dl)	29.4 ± 1.1	30.5 ± 2.5	31.4 ± 0.7	30.7 ± 1.5	29.5 ± 1.8	30.5 ± 1.1	30.2 ± 1.8
Total bilirubin (mg/dl)	0.96 ± 0.1	1.1 ± 0.2	1.1 ± 0.9	1.1 ± 0.1	1.1 ± 0.2	1.0 ± 0.2	1.0 ± 0.5
ALP (IU/l)	0.7 ± 0.8	0.8 ± 0.1	0.9 ± 0.4	0.9 ± 0.2	0.8 ± 0.1	0.9 ± 0.3	0.8 ± 0.1
ALT(IU/l)	195 ± 3.4	193 ± 1.8	200 ± 2.5	163 ± 9.7	187 ± 4.3	189 ± 1.2	191 ± 2.4
AST/(IU/l)	59.5 ± 1.2	60.5 ± 0.5	68.2 ± 0.2	66.5 ± 1.4	59.8 ± 0.1	59.9 ± 1.5	59.7 ± 1.2
GGT(IU/l)	150 ± 1.3	153 ± 1.4	152 ± 1.2	150 ± 1.8	145 ± 0.5	144 ± 0.5	150 ± 1.3
Protein (g/dl)	11.4 ± 1.4	12.4 ± 0.3	12.1 ± 1.5	10.1 ± 1.3	7.2 ± 0.5	9.4 ± 1.5	10.5 ± 0.2
Albumin (g/l)	75.4 ± 0.5	77.5 ± 0.8	74.2 ± 0.4	75.3 ± 1.5	71.2 ± 1.6	77.4 ± 0.9	74.3 ± 0.8
WBC (×10³/μL)	40.2 ± 0.8	42.3 ± 0.4	39.5 ± 08	42.1 ± 0.4	43.2 ± 1.2	42.1 ± 0.9	40.3 ± 0.5
Platelets (×10³/μl)	13.2 ± 0.9	11.9 ± 0.9	11.1 ± 0.4	12.2 ± 0.1	13.5 ± 1.1	$14.95 \pm 0.$	$14.95 \pm 0.$
RBC ($\times 10^6/\mu L$)	301 ± 1.3	298 ± 1.1	305 ± 0.9	308 ± 1.5	298 ± 1.1	304 ± 0.8	309 ± 01.3
HCT (%)	8.5 ± 2.5	8.9 ± 1.3	8.7 ± 2.3	9.2 ± 0.7	8.1 ± 1.2	8.5 ± 0.65	8.8 ± 1.3
Hb (%)	34.1 ± 1.2	32.2 ± 1.5	30.1 ± 1.5	32.8 ± 0.8	35.5 ± 0.9	30.9 ± 1.3	33.8 ± 1.2
MCH (pg)	12.2 ± 0.4	12.7 ± 0.5	13.7 ± 1.1	10.3 ± 0.9	12.5 ± 0.2	12.4 ± 0.9	10.9 ± 1.2
MCV (μm³)	30.4 ± 0.5	28.4 ± 1.2	26.4 ± 0.8	28.4 ± 1.2	25.6 ± 0.5	29.4 ± 0.9	29.1 ± 1.4
MCHC (%)	54.8 ± 1.2	53.4 ± 0.9	50.2 ± 0.8	54.3 ± 0.2	51.8 ± 1.2	56.1 ± 0.9	56.1 ± 1.4

NC=Negative Control.

with the ability to stabilise injured nerve membrane. ^{24,25} This mechanism may underline the analgesic effect of the co-administered agents in the thermal pain test because additivity is at best observed when two agents activate the same pathway. ¹⁵

Xylopic acid produces analgesic effect in neuropathic pain by acting on NMDA and adrenergic nociceptive pathways. 12 Any of these nociceptive pathways might have been modulated by xylopic acid in addition to the inhibition of Ca2+ channel-mediated neurotransmitter release, activation of excitatory amino acid transporters, modulation of potassium channels and inhibition of pathways involving inflammatory mediators by pregabalin to produce the observed synergism in the cold allodynia test.²⁴ Although xylopic acid and pregabalin co-administration produced analgesic synergism and additivity, the combination may not be without toxic effects, especially on prolong usage. Paclitaxel slightly reduced the red blood cells and platelets count and these effects were reversed by the co-administered agents. Paclitaxel as well as the coadministered agents may have the potential to induce hepato-biliary diseases on prolong usage. Generally, a slight elevation of ALP and total bilirubin was observed for paclitaxel and the relative high dose combination treatments. The liver enzymes were not significantly affected by the co-administration.

It is also likely that the long term use of the co-administered agents will have deleterious effect on the kidneys. Blood urea nitrogen and serum creatinine were slightly but not significantly elevated in the paclitaxel and co-administered agents treated groups. This may indicate the diminished excretory efficiency of the kidneys of these animals. This implies the co-administration of xylopic acid and pregabalin in neuropathic pain states associated with paclitaxel cancer chemotherapy is fairly safe. However, further studies to determine the long term safety of the co-administration is needed to exclude toxicity from cumulative doses.

CONCLUSION

Co-administration of xylopic acid and pregabalin may be beneficial in some types of painful neuropathy associated with paclitaxel administration. The co-administration is also fairly safe.

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CONFLICT OF INTEREST

The authors declare no conflict of interest.

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