Insight into Screening of Secondary Metabolites, Phenolic and Flavonoid Contents and Antioxidant Activity of *Raphanus sativus* L. Cultivated in Eastern Province of Saudi Arabia

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**ABSTRACT**

Medicinal plants and vegetables are promising source of antioxidant products. The purpose of this study is to evaluate the phytochemicals and antioxidant activity of leaves and roots of *Raphanus sativus* of Saudi origin. Various phytochemicals were screened in n-hexane, chloroform, ethyl acetate and aqueous fractions of leaves and roots of *Raphanus sativus* using standard protocols and IR screening method. Total phenolic (TPC) and flavonoid (TFC) contents were assessed by Folin-Ciocalteau and aluminium chloride methods respectively. The antioxidant activity was evaluated by DPPH antioxidant protocol, using trolox as standard. Results demonstrated that *Raphanus sativus* chemically characterized by the availability of various constituents such as flavonoids, steroids, saponins, tannins and carbohydrates at different levels in fractions of leaves and roots of *Raphanus sativus* and the absence of cardiac glycosides, anthraquinones and alkaloids which was further confirmed using FTIR analysis. TPC was ranged from (8.92±1.01) and (211.80±1.57) mg GAE/g extract and TFC was ranged from (0.036±0.03) and (11.57±0.60) mg QE/g extract for leaves and roots extracts respectively. Due to the high phenolic and flavonoid content in aqueous and ethyl acetate fractions of leaves and roots extracts, results demonstrated high antioxidant activity with IC50: (56.3±1.3) and (69.7±1.8) for aqueous fractions and (47.2±1.5) and (58.7±0.7) for ethyl acetate fractions of leaves and roots extracts respectively. Study revealed that ethyl acetate and aqueous fractions of leaves and roots of *Raphanus sativus* could develop a potential natural antioxidant herbal remedy. The study recommends future investigation to isolate and identify the bioactive secondary metabolites in *Raphanus sativus*.

**Key words:** *Raphanus Sativus*, Folin-Ciocalteau, DPPH, Total phenolic, Total flavonoid.

**INTRODUCTION**

Reactive oxygen species (ROS) are highly reactive substances which may bring about deep harms to tissues and consequently lead to serious degenerative conditions, such as ageing disorders, cancer, cardiovascular damage and diabetes.

Obviously, living organisms are gifted with defense mechanisms but unquestionably it is needed for additional support of external antioxidants to express more arsenals to fight. Antioxidants constitute are reputed to inhibit the harmful effects of ROS. However, synthetic antioxidants are available to help but they may have toxic and adverse effects. Hence, the development and exploitation of natural antioxidants are anticipated. Recently, medicinal plants including vegetables are promising source of antioxidant agents because of their potency and diversity in chemical structure as compared to synthetic compounds. The area of Arabian Peninsula (AP) comprises hundreds of species of medicinal plants and vegetables, The Kingdom of Saudi Arabia as part of AP, is rich biodiversity flora. Most of those native and cultivated plants are reputed for their higher contents of various bio-constituents and wide range of therapeutic benefits against different infectious diseases. *Raphanus sativus* L. (Brassicaceae) commonly known as radish, is an annual herb with white or brightly pink colored edible roots. It is commonly and traditionally used as a vegetable or appetizer in salads. It grows and cultivated worldwide including Saudi Arabia. Leaves and peel of root are always discarded which may possess potent biological properties. A literature survey indicated that, many studies revealed the identification of phytochemicals and biological activities including; anticancer, antimicrobial, antidiabetic and diuretic activities of roots and leaves of *Raphanus sativus* of many origins especially Indian origin. Different habitats and environmental conditions could alter the biosynthesis and machinery pathways in plant species. Hence, and based on the concept of different origins of the same plant may lead to different constituents or at least different concentrations of compounds. The present work is to investigate and highlight the phytochemical profiling including determination of total phenolic and flavonoid contents and radical scavenger activity of various fractions of different polarities of leaves and roots of white radish grown in Eastern Province of Saudi Arabia to categorize the most active fractions for future investigation to isolate and identify the bioactive secondary metabolites.

**MATERIALS AND METHODS**

**Materials**

Radish material (*Raphanus sativus*) was collected from local farm, Al-hasa, Eastern Province, Saudi...
Arabia. Leaves and roots were separated and subjected to air-drying according to standard herbarium procedures. A voucher sample (RS-2021) was kept in Department of Pharmaceutical Sciences, College of Clinical Pharmacy, King Faisal University. All reagents and chemicals were of the finest grade available.

**Extraction**

The air-dried powdered leaves and roots (300 g and 200 g, respectively) were thoroughly extracted three times (for 5 days) using 3 L of 70% methanol (Sigma Aldrich, Germany), applying cold maceration method to avoid destruction of secondary metabolites. Various extracts were concentrated using rotovap and then freeze-dried to yield the total dry extracts of leaves and roots, 24 g and 10 g respectively, which were kept in freezer for the next steps. The total extracts of leaves and roots, (20 g and 7 g, respectively) were mixed with distilled water (500 mL) and fractionated with n-hexane (4×500 mL). The n-hexane fractions were combined and concentrated to yield 9.5 g and 1.3 g, for leaves and roots respectively. The remaining aqueous fraction was subjected to further fractionation with chloroform (4×500 mL). The chloroform fractions were collected and concentrated to give 3.5 g and 1.2 g, for leaves and roots respectively. Similarly, the ethyl acetate fraction was developed to give 4.3 g and 2.4 g for leaves and roots respectively. The remaining aqueous fraction was lyophilized to give 2.4 g and 1.8 g for leaves and roots respectively.

**Screening of secondary metabolites**

Qualitative screening of the secondary metabolites was performed according to the standard protocols. 16,17

**Flavonoid**

Extract of 2 ml mixture was mixed with of 1 ml 2 % NaOH. Deep yellow color was developed, which turned colorless on addition of 2 drops of dilute acetic acid. This result indicates the presence of flavonoids.

**Alkaloid**

Extract of 0.5 ml was dissolved in 1.5 ml dilute Hydrochloric acid and filtered. The obtained filtrate was mixed with 2–3 drops of Dragendorf’s reagent and observed for five minutes. Dragendorf’s reagent produces an orange or orange-red precipitate indicates the presence of alkaloids.

**Saponins**

Extract of 1 ml was dissolved in 20 ml distilled water and mixture was shaken vigorously. The appearance of 1 cm froth for 15 minutes indicates the presence of saponins.

**Steroid**

Extract of 0.5 ml was dissolved in 5 ml chloroform and few drops of acetic anhydride and concentrated sulphuric acid were added from the side of the test tube. The upper yellow layer with green/blue color indicates the presence of steroids.

**Tannins/Phenolics**

About 2–3 drops of 5% ferric chloride solution was added to 0.5 ml of extract gives intense blue-greenish indicating the presence of tannins/phenolics.

**Glycosides**

Extract of 2 ml was mixed Fehling’s solution (A) and (B) and mixture was heated on a water bath for about two minutes. After heating, it gives a brick-red color that indicates the presence of glycosides.

**Anthraquinones**

Extract 0.5 ml was boiled with dilute sulphuric acid then filtered and cooled. The filtrate was extracted with chloroform and dilute ammonia solution was added to it. The aqueous ammonia layer becomes pink to red due to the presence of anthraquinones derivatives.

**Cardiac glycosides**

Extract of 10 ml was mixed with (4 ml) of solution of glacial acetic acid and 1 drop of 2% ferric chloride followed by 1 ml of concentrated sulphuric acid. A brown ring formed between the layers demonstrates the presence of cardiac glycosides.

**Carbohydrates**

Plant extract of 2 ml was mixed with a 10 ml Molisch reagent. Then, 2 ml concentrated sulphuric acid was added from the side of the test tube. The formation of a violet ring at the junction of two phases indicates the presence of carbohydrates.

**Fourier transform infrared spectroscopy (FTIR) characterization**

Characterization of various functional groups of various phyto-constituents was inspected employing FTIR using FTIR spectrophotometer by means of potassium bromide pellet technique. Potassium bromide discs were developed and about 5μL of different fractions were dropped on the discs and dried. The prepared discs of different fractions were inspected for FTIR (spectra ranged between 4000 and 400 cm⁻¹). 18

**Determination of total phenolic content (TPC)**

TPC was evaluated using the Folin-Ciocalteu index protocol. 19 Stock solutions with a concentration of 1 mg/ml of different fractions were arranged in methanol. Folin-Ciocalteu reagent (0.5 ml) and 6 ml of double distilled deionized water were sequentially added to 0.1 ml of stock solution of each fraction. Later, 1.5 ml of a 20% sodium carbonate solution was added to obtain final volume of 10 ml. A reaction was completed for 2 h at 25°C. Then, the absorbance of different mixtures was measured at 760 nm. Calibration curve of gallic acid (standard was prepared using serial dilution (0.5, 0.4, 0.3, 0.2 and 0.1 mg/ml in distilled water).

**Determination of total flavonoid content (TFC)**

TFC was investigated in accordance to Khalil et al method. 20 Various fractions (10 mg) were diluted in (100 ml) of mixture with ratio of (1:1 v/v) consisting of deionized water and acetonitrile. A solution of 0.25 ml of the serially diluted sample was added to 0.75 ml of a sodium nitrite (5% w/v) solution, followed by (0.15 ml) of a freshly prepared 10% aluminum chloride solution. The previously developed mixture was mixed with 0.5 ml of 1 M sodium hydroxide solution. Then, the volume of developed medium was adjusted to 10 ml with deionized double distilled water. The subsequent developed mixtures were set aside for 5 min and the absorbance was observed at 510 nm against the same components lacking of the sample. Calibration curve was prepared using quercetin as reference (0.0, 0.1, 0.5, 1, 5 and 10 μg/ml).

**Determination of antioxidant activity (DPPH assay)**

This protocol is based primarily on the reduction of reagent (2,2-diphenyl-1-picrylhydrazyl (DPPH)) dissolved in methanol and in presence of a donated protons from available molecules such as free radical scavenger substance, led to formation of non-radical form DPPH-H. The reduction in DPPH radicals was monitored by the reduction in its absorbance at 515 nm caused by antioxidant molecules.
RESULTS AND DISCUSSIONS

Phytochemical screening of different fractions

The preliminary phytochemical screening of various fractions demonstrated the presence of different phyto-constituents such as flavonoids, saponins, steroids, tannins/phenols, glycosides and carbohydrates at different levels in different fractions and the absence of cardiac glycosides, anthraquinones and alkaloids as shown in Table 1.

Fourier transform infrared spectroscopy (FTIR) screening

The spectra of various fractions showed bands approximately at 3400–3420 cm⁻¹, related to stretching vibration of hydroxyl groups (OH of phenolic compounds) Other bands were exhibited at 2900–3000 cm⁻¹ which could be assigned to stretching vibration of C-H bonds of aromatic skeletons like flavonoids or aromatic acids bands at 1650–1710 cm⁻¹, probably related to stretching vibration of carboxyl groups; stretching vibration of C=C and C=O groups of aromatic ring deformations (flavonoids and phenolic acids. Bands at 1350–1550 cm⁻¹ could be related to bending vibration of CH3, CH2, stretching vibration of flavonoids and aromatic rings. The bands at 1100–1260 cm⁻¹ would be due to vibration of C-O group of polyhydroxy constituents, such as hydroxyflavonoids. Bands at 1010–1090 cm⁻¹ would be related to alcohols and/or to C-O stretching ester groups. Bands at ~878 cm⁻¹, probably related to aromatic ring vibration (Figure 1).²² These results also confirmed the results of phytochemical screening.

TPC assessment

TPC in various fractions was calculated using the regression equation: y = 0.0357x + 0.0051, R² = 0.9702. The results were presented as the equivalence of milligrams of quercetin per gram of dried plant extract (mg GAE/g) (Figure 2A) Results depicted that the amount of total flavonoid constituents differed from organ to organ and varies from (0.036±0.03) to (11.57±0.60) mg GAE/g of dry extract. Similarly, to TPC, ethyl acetate fractions were the richest fractions in TFC then aqueous fractions. Chloroform fractions demonstrated the least amount of TFC compared to other fractions. On the other hand, n-hexane fractions hardly contained any flavonoid constituents (Figure 2B). The results demonstrated that leaves fractions are richer in contents compared to roots fractions.

Antioxidant activity (DPPH)

Besides, various fractions were inspected for their radical scavenger potential using the DPPH free radical scavenging assay (Figure 4). Ethyl acetate fraction showed marked scavenging activities (IC₅₀: 47.2±1.5 and 58.7±0.7 µg/ml, for leaves and roots fractions respectively), aqueous fraction showed moderate activity (IC₅₀: 56.3±1.3 and 69.7±1.8 µg/ml for leaves and roots fractions respectively) and n-hexane and chloroform fractions demonstrated much weaker potentials with IC₅₀ above 100 µg/ml, all results were comparable to the standard trolox (IC₅₀: 22.7 µM).

CONCLUSION

The significance of herbal extracts as antioxidant agents gained merits being of low risks and higher efficiency compared with synthetic agents. The present work demonstrated that Raphanus sativus of Saudi origin have several secondary metabolites which have significant antioxidant activity specially the ethyl acetate and aqueous fractions. Results could be credited to the higher TPC and TFC in these fractions. This was confirmed from the elevated GAE/g and QE/g values. These distinguished results serve as evidence to recommend future investigation to isolate and identify the bioactive secondary metabolites in Raphanus sativus. Hence, to open the gate for further developing of some of Saudi origin food into natural antioxidant herbal remedy in the pharmaceutical industry.

Table 1: Preliminary phytochemical screening of various fractions of Raphanus sativus.

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<th>LEA</th>
<th>LAQ</th>
<th>RnH</th>
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**Figure 1:** FTIR spectra of; ethyl acetate fraction of leaves (A), ethyl acetate fraction of roots (B), aqueous fraction of leaves (C) and aqueous fraction of roots (D).

**Figure 2:** Calibration curve for gallic acid (A) and total phenolic contents (B) of various fractions of leaves and roots of Raphanus sativus. nH; n-hexane fraction, CH; chloroform fraction, EA; ethyl acetate fraction, AQ; remaining aqueous fraction, GAE/g; Gallic acid equivalence per gram of dried plant extract.

**Figure 3:** Calibration curve for quercetin (A) and total flavonoid contents (B) of various fractions of leaves and roots of Raphanus sativus. nH; n-hexane fraction, CH; chloroform fraction, EA; ethyl acetate fraction, AQ; remaining aqueous fraction, QE/g; quercetin equivalence per gram of dried plant extract.
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GRAPHICAL ABSTRACT

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