GC-MS Analysis of Anti-Enterobacterial Dichloromethane Fraction of Mandukaparni (Hydrocotyle javanica Thunb.) – A plant from Ayurveda

Manab Mandal¹, Debabrata Misra¹, Narendra Nath Ghosh¹, Sukhendu Mandal², Vivekananda Mandal*¹

ABSTRACT

Background: Mandukaparni (Hydrocotyle javanica Thunb.) is a well known medicinal herb used as folklore medicine in many chronic and infectious gastric and other diseases by the people of Eastern Himalayan regions. However, the therapeutic active principles of this plant remained unknown. Objective: The main objective of the study was to characterize anti-enterobacterial dichloromethane fraction of the volatile oils of it by GC-MS. Materials and Methods: In the present study, dichloromethane (DCM) fraction (MP-DCMf) of Mandukaparni was collected by phase separation of the methanol extract and tested for anti-enterobacterial potentiality against human pathogenic gastrointestinal and food poisoning bacteria by agar well diffusion assay, viability assay and LDH assay and SEM studies. Characterization of the active MP-DCMf fraction was performed by TLC and GC-MS analysis. Results: The MP-DCMf possessed bio-active compounds that have antibacterial potentiality against both the Gram-positive and Gram-negative bacteria. The MIC and MBC values were in the range from 1.56 mg/ml to 0.78 mg/ml and 6.25 to 1.56 mg/ml, respectively. The time kill assay showed that at a dose of 3.12 mg/ml of MP-DCMf was lethal to the E. coli MTCC 723 at the 18th hr of treatment. LDH release had moderate positive correlation with the activity index and time of treatment whereas strong negative correlation with CFU count. It caused highest cell disruption in S. mutans. The principal compounds were D-carvon (30.949%); 1H-Isoindole-1,3(2H)-dione; 2-(2-chlorophenyl) (28.483%); Cyclohexanone; 2-methyl-5-(1-methylethen) trans (10.04%); D-Limnone (9.256%); 2,6-Octadien-1-ol, 3,7-dimethyl-acetate (6.684%); p-Cresol (0.551%); and Thymol (0.118%). Pub-chem database search also supports that these compounds have very strong bactericidal activity by membrane damage as evidenced by LDH and SEM studies. Conclusions: MP-DCMf contains many potential antibacterial compounds that can be used to combat the gastrointestinal and food poisoning bacterial pathogens. Key words: Anti-enterobacterial, Chemical profiling, Mandukaparni, Time kill assay, Volatile oils.

INTRODUCTION

Hydrocotyle javanica Thunb. (H. javanica) and Centella asiatica (L.) Urban. (C. asiatica) which are popularly known as Java pennywort and Indian pennywort respectively are commonly known as ‘Mandukaparni’ in Ayurveda for resemblance in their leaf shape and used for the ailments of indigestion, dysentery, nervousness and blood impurity as mutual substitute in herbal preparations.¹ C. asiatica (known as Gotu kola in Hindi) belonging to the Apiaceae family is an important medicinal plant of the Ayurveda for improving the brain function and delaying ageing process since thousands of years back and listed in the historic ‘Sushruta Samhita’, an ancient Indian medical text.² Fresh Gotu kola juice is taken along with cow’s milk in cough and asthma and for rejuvenation, decoction is taken in fever and dysentery and leaf is used as a vegetable also.³ The Hydrocotyle genus has almost 100 species that grow in tropical and temperate regions worldwide and belongs to the family Araliaceae, previously Apiaceae. H. javanica grows abundantly in Himalaya and southern part of India known as ‘Golpatta’ in Nepali, and ‘Mahagotukola in Hindi.’ The leaves of the plant are being used to treat several diseases like gastritis, throat sore, eye and ear infection by various ethnic communities like Bhutia, Damai, Kami, Lepcha, Limbu, Rai, Sherpa and Tamang of Darjeeling Hills (27°3'15.88" N and 88°15'28.10" E, elevation 2034 m) of West Bengal, India.⁴ It is also used to treat jaundice. Different biomedical aspects of the species like antibacterial activity and physicochemical parameters of methanol extract had been reported.⁵ In Kaushikasutra, Mandukaparni is described as an Tiktà (Bitter in taste) Aushadhi (Medicinal herb). Other ancient Ayurvedic texts described the medicinal properties of the herb as follows: It has a Kasaya (Astringent) after taste and a cooling effect. The Guna (Main quality) of the herb is Laghu (Light to digest); and its Virya (Potency) is Sheeta (Cold). Its Prabhava (Action) is Nootropic and neuroprotective. The Dhatu (Tissue) effect is through Rasa (Fluid), Rakta (Blood) and Mamsa (Muscle).³ Gastro-intestinal ailments like diarrhea and typhoid are one of the major public health problems in developing

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and developed countries. WHO estimates the diarrhea and typhoid fever diseases among infants and young children caused by various enteric pathogens, responsible for approximately 2.2 million and 0.6 million deaths each year. Some important pathogenic bacteria viz. Escherichia coli, Salmonella spp., Shigella sp., Vibrio cholera and V. parahaemolyticus are responsible for diarrhea causing more than two million deaths every year. In Indian hospitals, one-third of the total pediatric patients admitted are diarrheal and 17% mortality in indoor pediatric patients are diarrhea related.2 To combat such fatal disease many potent antibiotics have been used for the last few decades like Rifaximin, Metronidazole, Ciprofloxacin but the recurrent and rampant use of certain drugs lead to development of resistance among these pathogens. Antibiotics also have been associated with adverse effects which include hypersensitivity, allergic reactions and immune suppression. Many pathogens show multidrug resistance (MDR) against antibiotics. In these circumstances, plants such potential drug, the scaffold of the molecules of natural product is to treat these drug resistant pathogens. Most of the cases, to develop a need to develop potent alternative anti-enterobacterial compounds evaluate its mechanism of antibacterial action.

**MATERIALS AND METHODS**

**Plant material**

The Mandukaparni plant (H. javanica) was collected from Darjeeling hill areas of West Bengal, India. The voucher specimen (vide reference no. DGC/SP-02) was identified and authenticated at Central National Herbarium, Indian Botanic Garden, Shibpur, Howrah, India.

**Extraction and fractionation of bioactive components**

The bioactive components of the Mandukaparni powder were extracted by Soxhlet extractor using 80% aqueous methanol solvent and concentrated by rotary vacuum evaporator following the methodology of Mandal et al.13 The MP-DCMf fraction from the methanol extract concentrated by rotary vacuum evaporator following the methodology of Mandal et al.13 The MP-DCMf fraction from the methanol extract was recovered with 1/4 th volume of DCM using orbital shaker for 48 hours. The recovered fraction was evaporated to dryness and used for further study.

**Anti-bacterial activity assay**

The gastrointestinal pathogenic bacterial strains used in this study were procured from MTCC, Chandigarh, India, and MCC, Pune, India and were cultured in respective culture conditions. Antibacterial activity of the MP-DCMf (25 mg/ml) was evaluated by the agar wells diffusion method. Ciprofloxacin (50 μg/ml) and DCM (100%) were used as positive and negative control, respectively. The potentiality was assessed by activity index by calculating the ratio of diameter of zone inhibition of the extract and diameter of inhibition zone of antibiotic.

**Determination of MIC and MBC and time kill assay**

MIC was determined using 1.3% (v/v) Mueller Hinton agar (MHA) medium with the conc. range of MP-DCMf (25 mg/ml to 0.75 mg/ml).13 MBC and bactericidal kinetic assay were done by applying greater than the MIC dose on the actively grown bacterial strain (log phase growth) of B. cereus MTCC 1272, E. coli MTCC 723, E. faecalis MCC 2041T and S. mutans MTCC 497 and the viability was evaluated by plate count method after incubating at 37°C for 96 hrs. Time kill measurement was done at ½, 1, 2, 4, 8, 16 and 20hrs. The culture without treatment was considered as negative control.

**RESULTS**

**Antibacterial activity of the MP-DCMf**

The in-vitro antibacterial assay of the MP-DCMf demonstrated broad antibacterial activity against both Gram-negative and Gram-positive bacterial strains. The activity index of the bioactive fraction showed the highest value (0.866) against E. coli MTCC 723 and the lowest (0.822) in case of E. faecalis MCC 2041T. Apart from antibacterial activity against the gastrointestinal pathogens it had also potential activity index (0.675) against the topical pathogen, S. aureus MTCC 96, and food toxigenic B. cereus as shown in Table 1.

**MIC, MBC and bactericidal kinetic of the MP-DCMf**

The MICs were in the range from 3.12 mg/ml to 0.78 mg/ml. The MIC value for B. cereus, E. coli, S. typhimurium was 0.78 mg/ml while for B. subtilis, L. monocytogenes, S. mutans, was 1.56 mg/ml for the antibiotic activity was evaluated by lactate dehydrogenase (LDH) assay. Sonicated (100% amplitude, at 0.9 cycles/sec) and cell morphology

The effect of MP-DCMf on cellular integrity was evaluated by lactate dehydrogenase (LDH) assay. Sonicated (100% amplitude, at 0.9 cycles/sec) and the viability was evaluated by plate count method and linear regression model using MS – Excel 2007.

**Thin layer chromatographic analysis**

TLC analysis was done in Silica Gel 60 F254 precoated aluminium sheet (Merck, Germany) and developed in n-hexane-ethyl acetate (3:2, v/v) solvent system as mobile phase and detected under UV254 nm. The chemical group of the active fraction was evaluated by comparative TLC with standard flavonoids (Quercetin) and phenols (Gallic acid) using two different solvent systems, viz. chloroform: methanol (96:4, v/v) and toluene: ethyl acetate: methanol: formic acid (3:3:0.8:0.2, v/v/v), respectively, and detected under UV 254 nm. For alkaloids, chloroform: ethanol (9:1, v/v) was used as the solvent system and visualized by spraying with Dragendorff’s reagent. The Rf values were measured and compared with the standards.

**Statistical analysis**

Antimicrobial activities were calculated as mean ± SE. The correlation between LDH release and activity index, LDH release and time of treatment, and LDH release and Log CFU count were analyzed by Spearman’s 2-tailed bi-variate method and linear regression model using MS – Excel 2007.

**Prominent antibacterial activity of the MP-DCMf**

**GC-MS analysis**

The active compounds present in the MP-DCMf was analysed by using the GC-MS instrument (Model 7890B GC-240 ION TRAP MS, Agilent Technologies, USA) equipped with a capillary column, VF-5MS (Length-30m, ID-0.25 mm, Film-0.25 μm, Temp-max 325 °C) with the mass detection range of m/z 10-1000. The instrument was operated in electron impact mode at an emission current of 25 μAmps, maximum ion time of 65000 μSeconds, target TIC of 20000 counts, with injector temperature at 250 °C, and detected at 300 °C. The identification of compounds was done by matching the mass spectral records of NIST library and the Mole % of the compounds was determined by the formula: Mole % = A i/A c × 100, where, A i = Peak area count of individual compound and A c = Cumulative peak area count.

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E. faecalis, S. aureus and V. parahaemolyticus it was 3.12 mg/ml. The MBC values were in the range from 6.25 to 1.56 mg/ml. MBC value for B. cereus, E. coli, S. typhimurium was 1.56 mg/ml while for B. subtilis, L. monocytogenes, S. mutans it was 3.12 mg/ml and for E. faecalis, S. aureus and V. parahaemolyticus it was 6.25 mg/ml. The time kill assay showed that at a dose of 3.12 mg/ml, the MP-DCMf was lethal to the E. coli MTCC 723 at 18th hrs of treatment (Figure 1) which indicated that the active fraction had a potential bactericidal effect on the treated bacterial strain.

**Mode of action of the MP-DCMf**

**LDH activity and its correlation**

The LDH activity of MP-DCMf on the test strains was shown in Figure 2. A gradual increase in the LDH activity was recorded after 3 hrs and reaching the highest after 5 hrs of treatment. This indicated the MP-DCMf causes damage to the bacterial cell membrane. The statistical analysis revealed that LDH release had moderate positive correlation with the activity index and time of treatment whereas strong negative correlation with CFU count. The regression analysis also strongly supported the correlation results as represented in Figure 3.

**Effect on the cell morphology**

MP-DCMf caused cellular damage as confirmed by SEM photomicrographs (Figure 4A-D). It showed changes in the morphology and shape of the bacterial cells, like the formation of notch (4B), blebbing (4C) and decay of the cell walls (4D) which is comparable to the untreated bacteria (4A).

**TLC analysis**

The comparative TLC analysis of the MP-DCMf showed the presence of alkaloids, flavonoids, and phenols in the active fraction as shown in Figure 5. The presence of alkaloid in the fraction was confirmed as orange spots after spraying with the Dragendoff’s reagent which had Rf value of 0.55. In flavonoids test (Fig. 5B) quercetin (spot 1), MP-DCMf (spot 2) and MP-DCMf + quercetin (spot 3) showed the same Rf values i.e. 0.254 (Fig. 5B, spot 1). In phenol (Fig. 5A) gallic acid (spot 1), MP-DCMf (spot 2) and MP-DCMf + gallic acid (spot 3) showed the same Rf values of 0.690.

<table>
<thead>
<tr>
<th>S. No.</th>
<th>Enteric Strains</th>
<th>Strains properties</th>
<th>Growth condition</th>
<th>DCM Fraction</th>
<th>Antibiotics (50 μg/mL)</th>
<th>Activity Index</th>
</tr>
</thead>
<tbody>
<tr>
<td>1.</td>
<td>B. cereus</td>
<td>Pathogen of food borne illnesses</td>
<td>TSB, 37°C</td>
<td>15.6 ± 0.78</td>
<td>18.0 ± 0.9</td>
<td>0.866</td>
</tr>
<tr>
<td>2.</td>
<td>B. subtilis</td>
<td>Food borne illnesses</td>
<td>TSB, 37°C</td>
<td>15.0 ± 0.75</td>
<td>18.0 ± 0.9</td>
<td>0.833</td>
</tr>
<tr>
<td>3.</td>
<td>E. coli</td>
<td>GI, colicin indicator strain</td>
<td>TSB, 37°C</td>
<td>18.0 ± 0.9</td>
<td>20.0 ± 1</td>
<td>0.9</td>
</tr>
<tr>
<td>4.</td>
<td>E. faecalis</td>
<td>Gastrointestinal causing abdominal infections</td>
<td>NB, 37°C</td>
<td>14.8 ± 0.74</td>
<td>18.0 ± 0.9</td>
<td>0.822</td>
</tr>
<tr>
<td>5.</td>
<td>L. monocytogenes</td>
<td>HPF borne pathogen causing listeriosis</td>
<td>BHI, 37 °C</td>
<td>15.0 ± 0.75</td>
<td>18.0 ± 0.9</td>
<td>0.833</td>
</tr>
<tr>
<td>6.</td>
<td>S. aureus</td>
<td>Pathogenic, antibiotic sensitive strain</td>
<td>TSB, 37°C</td>
<td>12.5 ± 0.63</td>
<td>18.5 ± 0.93</td>
<td>0.675</td>
</tr>
<tr>
<td>7.</td>
<td>S. mutans</td>
<td>Dental caries pathogen</td>
<td>BHI, 37 °C</td>
<td>17.0 ± 0.85</td>
<td>19.5 ± 0.98</td>
<td>0.871</td>
</tr>
<tr>
<td>8.</td>
<td>S. typhimurium</td>
<td>Gastroenteritis pathogenic prototrophic strain</td>
<td>TSB, 37°C</td>
<td>16.4 ± 0.82</td>
<td>19.0 ± 0.95</td>
<td>0.863</td>
</tr>
<tr>
<td>9.</td>
<td>V. parahaemolyticus</td>
<td>Gastroenteritis halophilic organism</td>
<td>NB, 37°C</td>
<td>14.6 ± 0.46</td>
<td>17.0 ± 0.85</td>
<td>0.858</td>
</tr>
</tbody>
</table>

Table 1: Spectrum of antimicrobial activity of HJ-DCMf against the enterobacterial strains.

Here, DCM solvent shows negative inhibition zone against all the enteric pathogens.
Figure 2: LDH activity of *B. cereus*, *E. coli*, *E. faecalis*, *S. mutans* induced by MP-DCMf (3.12mg/mL). Each point represented the mean of triplicate trials and error bars represented standard errors.

Figure 3: Correlation of LDH release with different related parameters: (A) LDH and Activity index; (B) LDH and Treatment time; (C) LDH and log CFU/ml.
GC-MS analysis

The GC-MS analysis showed that D-Carvone occupied the maximum peak area (30.949%) and Cyclopentane, 1,1-dimethyl occupied the lowest peak area (0.024%). Other compounds present in significant proportions were 1H-Isodoindole-1, 3(2H)-dione, 2-(2-chlorophenyl) (28.483%), D-Limonene (9.256%), Phosphonic acid (1.723%), and p-Cresol (0.551%). About twenty two terpenoid compounds that occupied 66.881% of total moles were identified. The first compound was Fampridine with the least retention time i.e., 4.773 min. and the last compound was 1H-Isodoindole-1,3(2H)-dione, 2-(2-chlorophenyl) with the highest retention time i.e., 19.189 min. as detected in the spectrum (Figure 6, Table 2).

DISCUSSION

MP-DCMf has potential broad spectrum antibacterial activity at a dose of 25 mg/ml against Gram-negative and Gram-positive bacteria with the average diameter of zone inhibition of 17.00 mm and 15.00 mm, respectively. The minimum and maximum MBC values (6.25 mg/ml and 1.56 mg/ml, respectively) were significantly lower than the MBC value of other species of Hydrocotyle. Bactericidal activity of the MP-DCMf was the highest against the enteric pathogen E. coli, S. enterica serovar. typhimurium and another highly entero-pathogenic strain, E. faecalis. B. cereus, B. subtilis, L. monocytogenes, and S. mutans were also very susceptible to the active fraction at a lethal dose of 3.12mg/ml. The most of the bacterial strains showed their growth inhibition within the minimum concentration of 1.56mg/ml. Hence, the MP-DCMf fraction contains some potential bactericidal compounds that can be used against these pathogenic bacteria. The MP-DCMf also exhibited significant bactericidal activity against the dental carries pathogen, S. mutans. The antibacterial efficacy, MIC and MBC of the MP-DCMf was greater than the methanol extract as shown in Table 3. LDH activity results indicated that the MP-DCMf causes extracellular leakage due to bacterial cell membrane damage. Time kill assay has the implication in the bactericidal kinetics of the antibacterial agent against specific bacterial strain. It demonstrates the degree of lethality of the particular antibacterial agent. The comparative TLC analysis shows the presence of some potent antibacterial alkaloids, phenols and flavonoids in the MP-DCMf (Figure 5).

GC-MS analysis of the MP-DCMf showed eight alkaloids, six phenolics, one flavonoid and one glycoside compounds present at a proportion of 30.126%, 3.579%, 0.055% and 0.075% of total moles, respectively as shown in the Table 2. A GC-MS study of Hydrocotyle bonariensis also reported the presence of monoterpines (53.6% mol) and sesquiterpines (10.5% mol) as the major phytochemical classes. The monoterpenes α-pinene, β-pinene, and limonene represent the main components of the oil. These indicate the genus Hydrocotyle possesses the biosynthetic machinery of novel antimicrobials. For large scale isolation, purification and pharmaceutical application this plant may provide a valuable tool. Further in-vitro propagation and metabolic engineering could provide life supporting drugs for industrial uses.

In the MP-DCMf p-Cresol was detected as the major phenolic compound which possesses significant antibacterial potentiality and was also available in the methanol extract of the same plant. It might also increase the plasma membrane permeability that results in higher leakage of fluid material from bacterial cells and inhibit microbial respiration. Iso-indole alkaloids possess good activity against the Gram-negative bacteria (E. coli, K. pneumonia and P. aeruginosa, and yeasts (C. albicans, S. cerevisiae) with MIC ranging from 50-200µg/ml, while it was less susceptible against the Gram-positive bacteria (B. megaterium, B. subtilis and S. aureus). Iso-indole alkaloids act as nucleic acid synthesis inhibitor. Terpenoid shows antibacterial activity against E. coli, E. faecalis, K. pneumonia, P. aeruginosa and S.
Figure 5: Comparative TLC analysis of the MP-DCMf. (A) Phenols and (B) Flavonoids.

Figure 6: Gas chromatogram of the MP-DCMf.
**Dosha Karma (Effect on Humor) of Mandukaparni**: It overcomes with LDH release through the cell membrane of the bacterial cell. The bacterial zone inhibition by the active fraction is positively correlated with the activity index and LDH correlation analysis proved that the active compounds present in the DCM fraction was visualized by the GC-MS study for drug development, and vivid mode of action might explore the better application of this novel repository herb for anti-enteric drug development.

**Chemical Nature of the Phytocompounds Detected in GC-MS**

- **Alkaloids**
  - (i) Fampridine; (ii) 3-Furaldehyde; (iii) Pyridine-2,6-dimethyl; (iv) Pyrazine-2,3-dimethyl; (v) Pyrazole-4-carboxaldehyde; 1-methyl; (vi) Pyrazine-2-ethyl-6-methyl; (vii) 3,6-Dimethyl-2,3,3a,4,5,7a-hexahydrobenzofuran; (viii) 1H-Indoline-1,3(2H)-dione; 2-(2-chlorophenyl).
  - Total Mole %: 30.126

- **Flavonoids**
  - (i) 2-Cyclopetent-1-one,2-methyl; (ii) Trans-beta-Ocimene; (iii) 2-Cyclopetent-1-one,3-methyl; (iv) Alpha-Phellandrene; (v) 2,4,4,6-Tetramethyl-6-phenylheptane; (vi) D-Limonene; (vii) 2-Cyclopetent-1-one,2,3-dimethyl; (viii) 3-Carene; (ix) Ethanone, 1-(1H-pyrrol-2yl); (x) Cyclopentane,1,1dimethyl; (xi) Benzene,(butoxymethyl), (xii) 4-(1,2-Dimethyl-cyclopent-2-enyl)-butan-2-one; (xii) 2Cyclopentene-1-butanol, gamma2,3tetramethyl; (xv) Cyclohexanone ,2-methyl-5-(1-methylethen) trans; (xvi) Cylocboxanol, 2-methyl-5-(1-methylethenyl)-trans; (xvii) D-Carvone, (xviii) 2-Cyclohexen-1-one,2-methyl-5-(1-methylethyl); (xix) Thymol; (xx) 1,6-Octadien-3-ol,3,7-dimethyl,formate; (xxi) 2,6-Octadien-1-ol,3,7-dimethyl,acetate; (xxii) Benzene, 1,2,3-trimethoxy-5-(2-propenyl).
  - Total Mole %: 66.881

- **Terpenoids**
  - (i) Phosphoric acid, (p-hydroxyphenyl); (ii) p-Cresol; (iii) Methinol; (iv) 2,4-Dimethyl phenol; (v) p-Menth-1(7)-en-9-ol; (vi) 1,6-Benzene, 1,2,3-butanal,gamma2,3tetramethyl; (xxv) 1,6-Octadien-3-ol,3,7-dimethyl,formate; (xxvi) 2,6-Octadien-1-ol,3,7-dimethyl,acetate; (xxvii) Benzene, 1,2,3-trimethoxy-5-(2-propenyl).
  - Total Mole %: 3.579

- **Phenol**
  - Total Mole %: 0.075

- **Glycosides**
  - Total Mole %: 0.675

**Table 2**: Chemical nature of the phytocompounds detected in GC-MS.

<table>
<thead>
<tr>
<th>Class of compounds</th>
<th>Name of the compounds</th>
<th>Total Mole %</th>
</tr>
</thead>
<tbody>
<tr>
<td>Alkaloids</td>
<td>(i) Fampridine; (ii) 3-Furaldehyde; (iii) Pyridine-2,6-dimethyl; (iv) Pyrazine-2,3-dimethyl; (v) Pyrazole-4-carboxaldehyde; 1-methyl; (vi) Pyrazine-2-ethyl-6-methyl; (vii) 3,6-Dimethyl-2,3,3a,4,5,7a-hexahydrobenzofuran; (viii) 1H-Indoline-1,3(2H)-dione; 2-(2-chlorophenyl).</td>
<td>30.126</td>
</tr>
<tr>
<td></td>
<td>Flavonoids</td>
<td>0.655</td>
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<td></td>
<td>Terpenoids</td>
<td>66.881</td>
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<td></td>
<td>Phenol</td>
<td>3.579</td>
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<tr>
<td></td>
<td>Glycosides</td>
<td>0.075</td>
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</tbody>
</table>

**Table 3**: Comparative analysis of antibacterial potentiality of both DCM and Methanol extracts.

<table>
<thead>
<tr>
<th>S. No</th>
<th>Name of microbial strains</th>
<th>Diameter of zone inhibition</th>
<th>MIC value</th>
<th>MBC value</th>
</tr>
</thead>
<tbody>
<tr>
<td>1.</td>
<td>B. cereus</td>
<td>15.6 ± 0.78</td>
<td>0.78 ± 0.04</td>
<td>0.78 ± 0.04</td>
</tr>
<tr>
<td>2.</td>
<td>E. coli</td>
<td>18.00 ± 0.9</td>
<td>0.78 ± 0.04</td>
<td>0.78 ± 0.04</td>
</tr>
<tr>
<td>3.</td>
<td>L. monocytogenes</td>
<td>15.00 ± 0.75</td>
<td>8.60 ± 0.43</td>
<td>8.60 ± 0.43</td>
</tr>
<tr>
<td>4.</td>
<td>S. typhimurium</td>
<td>16.4 ± 0.82</td>
<td>10.50 ± 0.53</td>
<td>10.50 ± 0.53</td>
</tr>
<tr>
<td>5.</td>
<td>S. aures</td>
<td>12.5 ± 0.63</td>
<td>3.12 ± 0.16</td>
<td>3.12 ± 0.16</td>
</tr>
<tr>
<td>6.</td>
<td>S. mutans</td>
<td>17.00 ± 0.85</td>
<td>1.56 ± 0.08</td>
<td>1.56 ± 0.08</td>
</tr>
</tbody>
</table>

**Acknowledgements**

The corresponding author is grateful to DST, Government of India (SERB/F/5369/2013-14) for partial financial help of the study. The authors are also thankful to the KIIT-TBI, KIIT University, Bhubaneswar, India for GC-MS analysis and CRNN, University of Calcutta, Kolkata, India for SEM study.

**Conflicts of Interest**

None.

**Abbreviations**

- GC-MS: Gas Chromatography-Mass Spectrometry
- DCM: Dichloromethane Fraction
- SEM: Scanning Electron Microscope
- MIC: Minimum Inhibitory Concentration
- MBC: Minimum Bacterial Concentration
- TLC: Thin-Layer Chromatography
- CFU: Colony-Forming Unit
- LDH: Lactate Dehydrogenase

**References**


**Conclusion**

The study suggested that the MP-DCMF contained some potent antibacterial compounds belonging to the class alkaloids, monoterpenoid and phenol. The antibacterial efficacy of these three types of secondary metabolites among which the alkaloids have specific for one or a limited number of molecular targets whereas phenols are multi-target agents modulating the activity of proteins, nucleic acids and bio-membranes in a less specific way. Thus, it can be concluded that the Mandukaparni (H. javanica) which is used in ayurvedic preparations has the potential to treat against bacterial pathogens of gastrointestinal infections, food borne and topical diseases (including dental caries pathogens). This is the first scientific report on the antibacterial potentiality of DCM fraction of the high altitude folklore medicinal herb, H. javanica. So, further isolation, purification and structure elucidation of the active compounds, their acute as well as cytotoxicity tests, QSAR guided chemical study for drug development, and vivid mode of action might explore the better application of this novel repository herb for anti-enteric drug development.
Mandal, et al.: GC-MS Analysis of Anti-Enterobacterial Dichloromethane Fraction of Mandukaparni (Hydrocotyle javanica Thunb.) – A plant from Ayurveda


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**GRAPHICAL ABSTRACT**

Mandukaparni (Hydrocotyle javanica Thunb.) (Family Araliaceae)

- D-carvon
- p-Cresol
- Thymol
- 2-(2-chlorophenyl) Cyclohexanone
- 1H-Isoindoile-1,3 (2H)-dione

GC-MS profiling

Anti-enterobacterial potentiality

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