Immunostimulatory Activities of Pegagan Embun (Hydrocotyle sibthorpioides Lam.) in White Male Mice

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ABSTRACT

Introduction: Pegagan embun (Hydrocotyle sibthorpioides L.) has many pharmacological activities, such as improving the immune system. Aim: This research aims to study the immunomodulatory effect of pegagan embun herbs ethanol extract (Hydrocotyle sibthorpioides L.) determined by phagocytic activity and capacity of macrophages, total and percentage of leukocytes. Methods: 25 male white mice were divided into 5 equal groups. Negative control group were given Na CMC 0.5%, the extract treated group were given pegagan embun ethanol extract at doses of 10, 50, 200 mg/kgbw, and the positive control group was given Stimuno 50 mg/kgbw orally for 7 days. On the 8th day, the total and percentage of leukocytes were counted through blood sample taken intravenously. The mice were then induced with Staphylococcus aureus suspension. After one hour, the peritoneal fluids was taken to determine the macrophages activity and capacity. The macrophages phagocytic activity and capacity, total and percentage of leukocytes were analyzed by One-Way Anova followed by Duncan Multiple Range Test (p<0.05). Results: The results show significant increase of concentration (p<0.05) towards macrophages phagocytic activity and capacity, and total leukocytes count. Percentage of leukocytes show that lymphocytes increase significantly (p<0.05), meanwhile neutrophils segments decrease significantly (p<0.05). Conclusion: It can be concluded that ethanol extract of pegagan embun herb at doses of 10, 50, 200 mg/kgbw shows immunostimulatory activity.

Key words: Hydrocotyle sibthorpioides Lam., Staphylococcus aureus, Phagocytosis, Macrophage, Leukocytes.

INTRODUCTION

The immune system protects human from foreign substances and pathogens such as viruses, bacteria, parasites, and fungi. There are two types of immune response such as specific and non-specific immune response. Non-specific immune response works rapidly to protect human from microorganisms while specific immune response shows specific response towards specific microbes.1,2

Non-specific immune response protect the body by phagocytizing antigen such as bacteria without regarding its difference from other foreign substances. The most important phagocytic cells in this process is the macrophages. Macrophages come from adult monocytes in the tissues. The two main functions of macrophages are destroying antigens and presenting them to lymphocytes T as macrophages acts as Antigen Presenting Cells (APC).3

An immunostimulant can increase body defense mechanism. Generally, an immunostimulant is defined as a compound that can increase body defense mechanism specifically and non-specifically through cellular or humoral response.4 Certain plants contains compound that show immunomodulatory activity. One of those plants is known as pegagan embun (Hydrocotyle sibthorpioides L.) herbs that is widely used in Chinese traditional medicines to treat immune and liver related disease.5 Pegagan embun has several medicinal properties such as anti-swelling, anti-inflammatory, diuretics, antibiotics, antipiretics, detoxificans, and expectorants. Farong Yu et al. reported that Hydrocotyle sibthorpioides extract shows excellent anti-tumour activities and helps repairing mice’s immunologic functions.6

Based on the above explanations, a research to determine the immunomodulatory activity of Hydrocotyle sibthorpioides extract was conducted. The determined parameters were macrophages phagocytic activity and capacity, total leukocytes count, and percentages of leukocytes in white male mice induced by Staphylococcus aureus.

MATERIALS AND METHODS

Time and place
This study was carried out for 4 months at Research Laboratorium and Immunology and Serology Laboratorium of Faculty of Pharmacy of Andalas University.

Apparatus
The apparatus used in this study were measuring cylinder (Pyrex), erlenmeyer (Pyrex), gavage needle (Terumo), rotary evaporator (Ika), filter paper (Whatman), analytical balance (Ohaus), microscope (Olympus), TLC plate (Merck), centrifuge, WBC pipette (Assistant), mice cage, and surgical scissors.
Materials

The materials used in this study were *pegagan embun* (*Hydrocotyle sibthorpioides* Lam.), stimuno ( Dexa Medica, Batch No.: SLBZ4289), quercetin (Sigma-Aldrich, CAS Number: 117-39-5), aquadest (Andeska Laboratory), carbomethylcellulose (Na CMC), Giemsa stain 0,5% (Merck), ethanol 70% (Andeska Laboratory), *Staphylococcus aureus* (Microbiology Laboratory of Faculty of Pharmacy of Andalas University), nutrient agar (Merck), nutrient broth (Merck), physiologic NaCl (Andeska Laboratory), Turk solution (Sagara Husada Mandiri), and white male mice (Wistar).

Extract preparation

1 kg of dried *pegagan embun* were finely grinded (Mesh 20) and macerated with ethanol 70%. An amount of fine dried powder and solvent were added into a mcerator with a ratio of 1:10. The mixture was stirred occasionally for 6 hours, left for 18 hours, and then filtered. The above processes were repeated two times with the same ratio of dried powder and solvent. The macerates were collected and concentrated by rotary evaporator until a crude extract was obtained.\(^7\) The extract was tested before use. The tested parameters were organoleptic, phytochemical screening, and TLC profile.

Animal model

The animal used in this research was 25 naive white male mice, 2-3 months old that weigh 20-30 g. The test animal were grouped into 5 groups consisted of 5 animals each. The negative control group was given Na CMC 5% suspension, group 2,3, and 4 were given *pegagan embun* extracts with doses of 10, 50, and 200 mg/kgbw while positive control group was given stimuno with doses of 50 mg/kgbw. The extract was given orally for 7 days.

Bacteria culture

*Staphylococcus aureus* R. (SA) was cultured into slanted nutrient agar (NA) and then innoculated into a new NA using the innoculating needle. Then the bacteria was incubated for 24 hours at 37°C. *Staphylococcus aureus* R. was then moved into the nutrient broth (NB) and incubated at 37°C for 24 hours. It was then centrifuged at 2500 rpm for 25 minutes until a pellet was formed and suspended with physiological NaCl.\(^6\)

Total leucocyte count

Fresh blood was taken using WBC pipette until the number 0.5 and turk solution was taken until the number 11 then shook for 3 minutes. The first 2 drops of the solution in the pipette were removed. Then, 1 drop of the solution was dropped on the hemocytometer. Leave for 2 minutes for the leukocytes to be sedimented. The white blood cells were counted at the 4 corner of the counting chamber.\(^9\)

\[
\text{Total leucocyte count} = \frac{\text{Total of leucocytes}}{0,4}
\]

Percentage of leucocytes

On the 8th day, blood smear was made on the object glass and left to dry. Methanol was dropped on the dried blood smear and left to dry for 5 minutes. Giemsa stain was added and left to dry for 20 minutes. The blood smear was then rinsed with distilled water and left to dry. Immersion oil was added and examined the smear under the microscope. Count the number of eosinophils, segmented neutrophils, banded neutrophils, lymphocytes, and monocytes at 1000x magnification.\(^7\)

Macrophages phagocytic activity and capacity test

The mice were acclimatized for 7 days then administered with *Hydrocotyle sibthorpioides* extract suspension for 7 days. On the 8th day, blood sample was collected and the animals were administered with 0.5 ml *Staphylococcus aureus* R. suspension intraperitoneally. After 1 hour, the animals were sacrificed and peritoneal fluid sample was collected with a micropipette. The peritoneal fluid was smeared on the object glass, fixated with absolute methanol for 5 minutes, stained with Giemsa, left for 20 minutes, rinsed with distilled water, and left to dry. Immersion oil was added before examining the smear under the microscope with 1000x magnification. Macrophages phagocytic activity was determined based on the percentage of phagocyte that carried out phagocytosis out of 100 phagocytes. Macrophages phagocytic capacity was determined based on the number of phagocytosed *Staphylococcus aureus* R by 50 active phagocytes.\(^10,11\)

Data analysis

The macrophages phagocytic activity and capacity, total leucocyte count, and percentage of leukocytes were analyzed statistically by one-way ANOVA and continued with Duncan’s Multiple Range Test.

RESULTS AND DISCUSSIONS

The herb that was used in this study is *pegagan embun*. The herb was identified as *Hydrocotyle sibthorpioides* Lam, comes from a family of Araliaceae Lam. by Anda Herbarium, Department of Biology of Faculty of Mathematics and Natural Sciences of Andalas University.

116,126 g of extract was obtained from 1 kg of dried *pegagan embun* with an extraction yield of 11.61%. The extraction yield was in accordance with specification stated by the Herbal Pharmacopoeia (2008). The organoleptic test results show that *pegagan embun* extract has thick blackish green appearance, specific odour, and bitter taste. Phytochemical screening tests show that the ethanol extract contains alkaloid, flavonoid, phenolic, and saponin compounds.

Thin layer chromatography (TLC) profile is a qualitative test to identify the marker compound (quercetin) in *pegagan embun* extract. The stationary phase used was TLC Silica gel 60 F\(_{254}\). The mobile phase used were n-hexane and ethyl acetate with a ratio of 6:4. The TLC profile was examined under UV light with a wavelength of 254 nm. The retention factor (RF) of the extract and pure quercetin was 0.51. This show that the extract contains quercetin because it has the same RF as pure quercetin, which can be seen in Figure 1.

![Figure 1: Thin layer chromatography profile of pegagan embun ethanol extract. (S) pegagan embun extract, (P) quercetin.](image-url)
The results for macrophages phagocytic activity in animals given doses of 10, 50, 200 mg/kgbw of extract, and 50 mg/kgbw of stimuno were shown in Table 1 and Figure 2. Groups that were administered with pegagan embun extract show higher macrophages phagocytic activity compared to group that was administered with Na CMC 0.5%. One-way ANOVA test shows that pegagan embun extract influence macrophages phagocytic activity significantly (p<0.05). Duncan’s multiple range test show that macrophages phagocytic activity increases with increasing doses (p<0.05). Group that was administered 200 mg/kgbw pegagan embun extract show the same macrophages phagocytic activity as group administered with stimuno (p>0.05).

The results for macrophages phagocytic capacity were shown in Table 2 and Figure 3. Groups treated with pegagan embun extract at doses of 10, 50, and 200 mg/kgbw show significant increase in macrophages phagocytic capacity (p<0.05). Groups treated with extract show higher

Table 1: Percentage of peritoneal macrophages phagocytic activity in control groups and male white mice treated with pegagan embun (Hydrocotyle sibthorpioides Lam.) extract.

<table>
<thead>
<tr>
<th>Doses</th>
<th>Percentage of macrophages phagocytic activity (%)</th>
<th>Mean ± SD</th>
</tr>
</thead>
<tbody>
<tr>
<td>Na CMC 0.5 %</td>
<td>51.40 ± 7.70⁴</td>
<td></td>
</tr>
<tr>
<td>10 mg/kgbw</td>
<td>68.00 ± 6.20⁴</td>
<td></td>
</tr>
<tr>
<td>50 mg/kgbw</td>
<td>76.40 ± 3.57⁴</td>
<td></td>
</tr>
<tr>
<td>200 mg/kgbw</td>
<td>88.20 ± 4.96⁴</td>
<td></td>
</tr>
<tr>
<td>Stimuno 50 mg/kgbw</td>
<td>91.60 ± 7.02⁴</td>
<td></td>
</tr>
</tbody>
</table>

Figure 2: Bar graph of dosage versus percentage of peritoneal macrophages phagocytic activity in white male mice.

Figure 3: Bar graph of dosage versus percentage of peritoneal macrophages phagocytic capacity in white male mice.
Table 2: Mean of peritoneal macrophages phagocytic capacity in control groups and male white mice treated with pegagan embun (Hydrocotyle sibthorpioides Lam.) extract.

<table>
<thead>
<tr>
<th>Doses</th>
<th>Mean ± SD</th>
</tr>
</thead>
<tbody>
<tr>
<td>Na CMC 0.5%</td>
<td>72.60 ± 8.08</td>
</tr>
<tr>
<td>10 mg/kgbw</td>
<td>195.80 ± 20.93</td>
</tr>
<tr>
<td>50 mg/kgbw</td>
<td>218.60 ± 15.75</td>
</tr>
<tr>
<td>200 mg/kgbw</td>
<td>234.60 ± 28.91</td>
</tr>
<tr>
<td>Stimuno</td>
<td>274.40 ± 46.77</td>
</tr>
</tbody>
</table>

Table 3: Mean of total leukocyte count in control groups and male white mice treated with pegagan embun (Hydrocotyle sibthorpioides Lam.) extract.

<table>
<thead>
<tr>
<th>Doses</th>
<th>Mean ± SD</th>
</tr>
</thead>
<tbody>
<tr>
<td>Na CMC 0.5%</td>
<td>4370 ± 496.99</td>
</tr>
<tr>
<td>10 mg/kgbb</td>
<td>8540 ± 482.70</td>
</tr>
<tr>
<td>50 mg/kgbb</td>
<td>8810 ± 207.36</td>
</tr>
<tr>
<td>200 mg/kgbb</td>
<td>9460 ± 403.73</td>
</tr>
<tr>
<td>Stimuno</td>
<td>10020 ± 668.58</td>
</tr>
</tbody>
</table>

Table 4: Percentage of leukocytes in control groups and male white mice treated with pegagan embun (Hydrocotyle sibthorpioides Lam.) extract.

<table>
<thead>
<tr>
<th>Doses</th>
<th>Segmented neutrophils</th>
<th>Banded neutrophils</th>
<th>Eosinophils</th>
<th>Lymphocytes</th>
<th>Monocytes</th>
</tr>
</thead>
<tbody>
<tr>
<td>Na CMC 0.5%</td>
<td>45.40 ± 3.36a</td>
<td>3.60 ± 1.51</td>
<td>1.80 ± 0.83</td>
<td>45.80 ± 2.16a</td>
<td>3.40 ± 2.30</td>
</tr>
<tr>
<td>10 mg/kgbw</td>
<td>41.20 ± 1.92b</td>
<td>4.40 ± 1.51</td>
<td>2.20 ± 0.83</td>
<td>47.80 ± 3.11b</td>
<td>4.40 ± 1.34</td>
</tr>
<tr>
<td>50 mg/kgbw</td>
<td>40.00 ± 1.58bc</td>
<td>3.60 ± 1.51</td>
<td>2.20 ± 0.83</td>
<td>50.00 ± 2.23c</td>
<td>4.20 ± 0.83</td>
</tr>
<tr>
<td>200 mg/kgbw</td>
<td>35.20 ± 1.64c</td>
<td>4.00 ± 1.58</td>
<td>2.20 ± 1.30</td>
<td>53.80 ± 2.28c</td>
<td>4.80 ± 2.04</td>
</tr>
<tr>
<td>Stimuno</td>
<td>37.60 ± 2.50d</td>
<td>1.80 ± 0.83</td>
<td>1.80 ± 0.83</td>
<td>55.60 ± 1.14d</td>
<td>3.20 ± 1.09</td>
</tr>
</tbody>
</table>
Figure 4: Peritoneal macrophages of white male mice. (A) macrophage, (B), activated macrophage, (C) Staphylococcus aureus.

Figure 5: Bar graph of dosage versus total leukocyte count in white male mice.

Figure 6: Bar graph of dosage versus percentage of leukocytes in white male mice.
ACKNOWLEDGMENT

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CONFLICTS OF INTEREST

The authors declare that there is no conflict of interest.

REFERENCES

SUMMARY

Pegagan embun (Hydrocotyle sibthorpioides L.) is known for its immunostimulatory effect. The aim of this research is to study the effect of pegagan embun ethanol extract on macrophages phagocytic activity and capacity, total leukocyte count, and percentage of leukocytes. 25 male white mice were divided into 5 equal groups. Negative control group were given Na CMC 0.5%, the extract treated groups were given ethanol extract of pegagan embun herbs at doses of 10, 50, 200 mg/kgbw, and the positive control group was given Stimuno 50 mg/kgbw orally for 7 days. On the 8th day, the total and percentage of leukocytes were counted through the blood sample taken intravenously. The mice were then induced with Staphylococcus aureus suspension. The results show significant increase of concentration (p<0.05) towards macrophages phagocytic activity and capacity, and total leukocytes count. Percentage of leukocytes show that lymphocytes increases significantly (p<0.05), meanwhile neutrophils segments decreases significantly (p<0.05). It can be concluded that ethanol extract of pegagan embun herb at doses of 10, 50, 200 mg/kgbw shows immunostimulatory activity through increasing macrophages phagocytic activity and capacity, increasing the number of lymphocytes, and decreasing the number of segmented neutrophils. Pegagan embun extract at doses of 200 mg/kgbw show similar immunostimulatory effect as 50 mg/kgbw stimuno.
Afriwardi, et al.: Immunostimulatory Activities of Pegagan Embun (Hydrocotyle sibthorpioides Lam.) in White Male Mice

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