Phytochemical Analysis and Antioxidant Activity of Cotinus coggygria Scop. from Armenian Flora

1Institute of Pharmacy YSU, 0025, 1 Alek Manukyan st., Yerevan, RA, ARMENIA.  
2Yerevan State Medical University after M.Heratsi, Department of Pharmacognosy, 0025, 2 Koryun st., Yerevan, RA, ARMENIA.  
3Analytical laboratory after Academician Emil Gabrielyan, branch of “Scientific center of drug and medical technology expertise” CJSC, 0051, Komitas Ave., 49/4, Yerevan, RA, ARMENIA.  
4Scientific and Production Center «Armbiotechnology» of National Academy of Sciences RA, 0056, 14 Gyurjyan Str., Yerevan, RA, ARMENIA.  

ABSTRACT  
Armenian flora is stood out by the variety of its medicinal and edible plants. Here small plants as well as trees and shrubs are met. Cotinus coggygria of Armenian flora is used in folk medicine. The aim of this investigation was to determine the chemical composition of Armenian flora’s smoke trees leaves’ essential oil and antiradical activity of ethanol extract of leaves and branches of smoke tree. Methods: The aerial parts were extracted by hydrodistillation method, and the composition was analyzed by gas chromatograph interfaced to a mass spectrometer. Antiradical activity of essential oil from leaves and branches was determined by using appropriate methods. Results: The total amount of flavonoids has been determined in the leaves of the smoke tree by micrictin. This analysis revealed that the flavonoid amount in ethanol extract of C. coggygria, growing in Armenia, was 0.94%. In volatile oil of Smoke tree leaves 22 compounds were determined. The results demonstrated that in the essential oil of leaves of Cotinus from Armenian flora predominated Germacrene D, Linalool, formate, α-Terpineol sesquiterpene and diterpene alcohol Thunbergol. Ethanol extracts of leaves and branches of Smoke tree showed antioxidant activity. Conclusion: From the results, it is evident that C. coggygria from Armenia flora contains various bioactive compounds and the extracts of leaves pronounced higher antioxidant activity and recommended as a plant of phytopharmaceutical importance.  
Key words: Cotinus coggygria Scop., Essential oil, Ethanol extract, Antiradical activity, DPPH.

INTRODUCTION  
Nowadays, from the standpoint of obtaining biologically active raw materials, wild plants are noticeable because of the interest in their mass-growing habitats and significant raw material reserves. Most notable are pharmaceutical raw materials containing polyphenolic compounds that have a broad spectrum of pharmacological activity (used as a coagulant, anti-inflammatory, antibacterial, etc.). From this point of view, the plants belonging to the family Anacardiaceae are quite unique. Some plants in this family are widely cultivated as edible plants, some are poisonous, others serve as medicines and a source of wood.  
Cotinus coggygria (Cotinus coggygria Scop., Synonymous with Rhus cotinus L.) belongs to the family Anacardiaceae, growing up to 5 - 12 m high, spherical or canopy foliage, 18 cm thick, leafy shrub covered with brownish bark. This plant is usually either considered as large shrubs or small trees. It has glaucous, simple, ovate or obovate leaves, 3–8 cm long. The flowers are pentameric, pale yellow or yellow-green, hermaphrodite or some of them abortive, with long peduncles, in terminal loose inflorescences. This plant grows in the Caucasus, Crimea, northern Ukraine, Central Asia, Georgia, usually in dry places, often on mineral and sandy slopes, at 1500 m above sea level.  
According V. Koleva and E. Simeonov(2014) Cotinus coggygria is wide spread in Southern Europe, the Balkans, and Southwestern and Central Asia. Extracts from the leaves, twigs, wood, and inflorescences of the plant are used in the ethnomedicine of Eastern and Southeastern Europe and China as anti diarrhoetic, anti-inflammatory and anti-paradentosis.1 Cotinus coggygria, also known as the “smoke tree”, is one of the two species constituting a small genus of the family Anacardiaceae, viz., C. coggygria Scop. (syn.: Rhus cotinus L.) and Cotinus obovatus Raf., the American smoke tree. It has a wide distribution from southern Europe, the Mediterranean, Moldova and the Caucasus to central China and the Himalayas.2  
Due to the results of a mass spectrometric study of essential oil obtained from the leaves of Cotinus coggygria Scop growing in Turkey, 42 active components were found, representing 99.6% of the total components detected. The major constituents were identified as limonene (48.5%), (Z ) (–)-β-ocimene (27.9%) and (E) -β-ocimene (9.7%).3 Similar research was done on raw materials cultivated from Smoke trees growing in Serbia and Greece. The quantitative contents were different, and in all cases the predominant ingredient was limonene. In this study, the Smoke tree leaves were standardised based on limonene, while in scientific sources it is based on myrcene.  
The leaves of Smoke trees growing in Georgia were studied. Antioxidant activity of phenolic fraction obtained from the leaves extract was confirmed.4 A
total of 30 compounds were identified in essential oil of Rhus cotinus the dominant components were monoterpenes (65,9%), which were beta-pinene, camphene, limonene, alpha-pinene and p-cymene followed by sesquiterpene hydrocarbons(20,6%). The oil exhibited antioxidant activity by inhibiting betta- carotene bleaching (56.4±1.88%), and by scavenging DPPH free radical (IC50 =720±0.10 µg M1). In another study the antimicrobial activity of the young shoots of the smoke tree was investigated. The acetone extract and the derived ethyl acetate fraction effectively inhibited the growth of Gram-positive and Gram-negative bacteria (MIC 25-200 µg/ml), while the chloroform fraction showed pronounced activity against the yeast Candida albicans (MIC 3.12 µg/ml). The ethyl acetate fraction exhibited a significant ferric-reducing ability (10.7 mmol Fe(2+)/ g extract), a very high DPPH radical scavenging activity (SC50 =1.7 µg/ml) and inhibition of lipid peroxidation (IC50 = 41.8 µg/ml). High amounts of total phenolics (929.8 mg/g), tannins (833.8 mg/g) and flavonoids (35.5 mg/g) were determined in the ethyl acetate fraction, which also exerted significant anti-inflammatory (76.7%) and cytotoxic effects (IC50 = 15.6 µg/ml). In the study by the author Saini Shagun et al. (2016) 19 compounds were identified in the essential oil of Cotinus. The compounds characterizing major peaks were a-Phellandrene (7,83%), a-Mircene (5,36%), (+)-2-Bornanone (14,52%), Caryophyllene (7,65%), (+)-epi-Bycyclesquiphellandrene (5,59%), e-Celene (9,56%), Globulol (7,95%). The composition of monoterpene hydrocarbons was observed dominant over sesquiterpene hydrocarbons. The author Sanja Matic’ et al. reported that the Anacardiaceae Lindl. family comprises of many species which are used in nutrition and in traditional folk medicine for the treatment of several human diseases. Cotinus coggygria Scop. commonly known as ‘smoke tree’, is a commercial ornamental plant with high medicinal usages, belongs to the family Anacardiaceae.

The plant has been extensively investigated in a broad range of studies to provide scientific evidence for folklore claims or to find new therapeutic uses. Numerous activities namely antioxidative, antibacterial, antifungal, antiviral, anticancer, antigenotoxic, hepatoprotective and anti-inflammatory have been demonstrated for all parts of these plants by in vivo and in vitro studies. Essential oils and extracts showed various pharmacological and biological properties which make them an effective remedy for various kinds of illnesses. Considering data from the literature, it could be demonstrated that C. coggygria possesses diverse bioactive properties and immense utilization in medicine, health care, cosmetics and as health supplements.

Cotinus coggygria is an important herbal medicine in Armenia. From this point of view, it is quite interesting that Cotinus coggygria typical to the Caucasus region especially growing in the flora of Armenia, the pharmacognostic study of which can be promising in terms of locally sourced raw materials and production. Smoke trees are found in Armenia in the low and middle mountainous areas of Tavush and Syunik regions. It grows on rocky slopes, in bushes and forest edges. So, the aim of this research was to study the phytochemical and comparative antioxidant activity of the leaves and branches of smoke tree growing in Armenia.

MATERIALS AND METHODS

Plant materials and essential oil extraction

Cotinus coggygria leaves and shrubs collected in Yerevan, 40°0'51" N, Longitude: 44°30'48" E Elevation above sea level: 994 m = 3261 ft during the blossoming period (July 2019). The primary processing and drying of a raw material was carried out by WHO instructions (GACP, WHO 2011). The identification of plant was carried out at the Department of Pharmacognosy, Yerevan State Medical University (Armenia). Plant samples were deposited and are available at the Herbarium of the Institute if Botany, National Academy of Sciences of Armenia.Yeravan (Armenia). A voucher specimen number is ERE 193405. EO was extracted from the aerial parts of leaves plants by hydro-distillation method (with Cleveenger –type apparatus) that lastad approximately 4 h. The distilled essential oils were dehydrated with anhydrous sodium sulfate and was stored at +4±1°C in the dark for further use.

Gas chromatography/mass spectrometry (GC-MS) analysis

The EO composition was analyzed at the "Analytical laboratory after academician Emil Gabriyelyan", branch of "scientific center of drug and medical technology expertise” CJSC (Armenia). GC analysis was carried out using a gas chromatograph (BRUKER 450-GC, USA. An OPTIMA-FFAP capillary column (0.25 mkm, 60 m x0.25 mm (ID, MACHEREYNAGEL, Germany) was used for the separation of volatile oil compounds. Helium gas carrier flow-rate 1.0ml/min, volume of injected sample 2mkl, the oven temperature 220°C, temperature gradient 50°C (2min), split-separation -5. Compounds were identified using the NIST Mass spectral. The identification of peaks was tentatively carried out based on library search using NIST -2013.

Ethanol extracts preparation and determination of total flavonoid content by myricetin

Flavonoids have low or moderate solubility in water, the extraction was made using dried plant material to eschew the dilution of the solvent. Aerial parts of leaves plants were at 55 °C for 24h. 1g powdered dried aerial parts of leaves was homogenized in 10 to 15ml 40% ethanol and left over night at 10°C. The extract was centrifuged for 5 min at 5000grm, and the supernatant was isolated. The precipitate was extracted by 4-folds, and the combined supernatant was dried by evaporation at room temperature. The evaporated mass was solved in ethanol, and the extracts in different dilutions were used. The absorbance of the extract was measured (415nm) with the spectrophotometer Genesis 10S UV-Vis (Thermo Scientific, USA), and total flavonoid content of EE was expressed in terms of myrcitin (E1%1cm 380/myrcetin+ AlCl3 l/f).

Total flavonoid content (mg/g)=D x100 x2
380 x m x 2

Where m- mass of the row material
D- optic density

The total flavonoid content in the ethanol extract of leaves of cotinus was determined using aluminium chloride colorimetric assay.

DPPH (free radical-scavenging) assay

Scavenging free radical potentials were determined in ethanol solution of DPPH. In the research, the methanol, a stable radical of DPPH ([2,2-diphenyl-1-picrylhydrazyl, C14H11N3O2, M = (394.33) (Sigma Aldrich GmbH)) and the dry methanol extracts of the leaves and shrubs of C. coggygria were used.

The antioxidant activity determination was carried out using the spectrophotometric method, in which the natural antioxidant interacts with the stable chromogen radical DPPH. The antioxidant activity was determined after the interaction of the methanol solution of DPPH with the test solution and was calculated using a calibration graph of the DPPH (the dependence of the optical density from the DPPH concentration). The absorbance of the samples was determined at the wavelength 515 nm.

The optical density definition of the testing solutions was performed by the device Helios Comp Thermoelectron (England). The measurements were carried out in five replicates. The optical density of the DPPH was recorded after 1, 5 and 20 minutes.


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The scavenging effect in percentage was defined according to this formula:

\[
\text{Antioxidant Activity (\%) = \left( \frac{(C_c - C_{\text{ext}})}{C_c} \right) \times 100\%}
\]

where \( C_c \) is the concentration control absorbance, \( C_{\text{ext}} \) is the concentration corresponds to the absorbance in the presence of extracts.

Antioxidant activity was calculated as the effective concentration at which the DPPH radicals were inhibited by 50% \( (\text{IC}_{50}) \).

Statistical analysis

Statistical analysis was made by SPSS ® for Windows (Version 16.0, Chicago, IL, USA). The results were presented as a mean ± standard error of mean; \( p<0.05 \) was regarded as statistically significant.

RESULTS AND DISCUSSION

Essential oil composition

The GC/MS analysis of oil has revealed that the essential oil of Cotinus coggygria growing in Armenian flora consists of many complex compounds. According to GC-Ms dates, in the volatile oil of Smoke tree leaves 22 compounds were identified. Regarding the composition of the studied EOs, significant differences in relation to the major components as well as the content of the different terpene classes was observed. Sesquiterpenes and diterpenes were most present classes in the oil of C. coggygria. The compounds characterizing major peaks were Germacrene D, Linalool, formate, â –Terpineol and Thunbergol (10.65%, 10.04%, 8.78 % and 6.87% resp.). The latter accompanies the volatile oil, which testified about olea-resina mixture. In oil of C. coggygria was revealed another sesquiterpenes, among which Cycloheptane, 4-methylene-1-methyl-2-(2-methyl-1-propen-1-yl)-1-vinyl-(3.81%), â-Humulene (2.68%), Caryophyllene (2.65%), followed by â-Muurolene (2.06%) et al. (Table 1, Figure 1).

The total flavonoid content was expressed using the specific density for myrcetin, and the result was expressed as myrcetin specific density. Our studies have confirmed that the flavonoid amount in ethanol extract of C. coggygria, growing in Armenia, was 0.94%.

Free Radical Scavenging Activity (% DPPH inhibition)

The results of the study showed that all the samples of the raw material (smoke tree leaves and twigs) are manifested a positive antioxidant activity, determined by the rate of the DPPH inhibition.

Figure 2 shows the relationship between the optical density (D) and concentration (C) of the experimented extracts for corresponding concentrations 2mg, 4mg and 8mg; the results of which are given in Tables 2 and 3.

Table 1: Phytocomponents identified in the essential oil of leaves of Smoke trees (Cotinus coggygria Scop.), growing in Armenian flora by GC-MS.

<table>
<thead>
<tr>
<th>Compound name</th>
<th>Rt</th>
<th>Formula</th>
<th>Mr</th>
<th>Area %</th>
</tr>
</thead>
<tbody>
<tr>
<td>1. 1,6-Octadien-3-ol, 3,7-dimethyl-formate</td>
<td>2.16333</td>
<td>C_{11}H_{18}O_{2}</td>
<td>182</td>
<td>10.04</td>
</tr>
<tr>
<td>Linalool, formate Synonym: Linalool, formate</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>2. 3-Cyclohexene-1-methanol, â,â 4-trimethyl-</td>
<td>2.45333</td>
<td>C_{11}H_{18}O</td>
<td>154</td>
<td>8.78</td>
</tr>
<tr>
<td>Synonym: â -Terpineol</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>3. Bicyclo[3.1.1]hept-3-en-2-one, 4,6,6-trimethyl-</td>
<td>2.57</td>
<td>C_{10}H_{18}O</td>
<td>150</td>
<td>0.14</td>
</tr>
<tr>
<td>Synonym: Berbenone Verbenone</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>4. n-Valeric acid cis-3-hexenyl ester</td>
<td>2.68667</td>
<td>C_{11}H_{20}O_{2}</td>
<td>184</td>
<td>1.08</td>
</tr>
<tr>
<td>Synonym: cis-3-Hexenyl valerate</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>5. 1,5-Dimethyl-1-vinyl-4-hexenyl butyrate</td>
<td>2.957</td>
<td>C_{14}H_{24}O_{2}</td>
<td>224</td>
<td>1.71</td>
</tr>
<tr>
<td>Synonym: Linalyl butyrate</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>6. Bicyclo[2.2.1]heptan-2-ol, 1,7,7-trimethyl-, acetate, endo-</td>
<td>3.507</td>
<td>C_{12}H_{20}O_{2}</td>
<td>196</td>
<td>4.44</td>
</tr>
<tr>
<td>Synonym: Bornyl acetate</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>7. 2-Oxopentanoic acid, 2' -acetylphenyl ester</td>
<td>3.69</td>
<td>C_{13}H_{14}O_{4}</td>
<td>234</td>
<td>0.31</td>
</tr>
<tr>
<td>Synonyms: no synonyms.</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>8. (E)-2-Hexenyl tiglate</td>
<td>4.41</td>
<td>C_{11}H_{18}O_{2}</td>
<td>182</td>
<td>0.54</td>
</tr>
<tr>
<td>Synonyms: no synonyms.</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>9. 3-Cyclohexene-1-methanol, â,â,â 4-trimethyl-, acetate</td>
<td>4.57</td>
<td>C_{11}H_{18}O_{2}</td>
<td>196</td>
<td>4.3</td>
</tr>
<tr>
<td>Synonyms: â -Terpineol acetate</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>10. â-Cubebene</td>
<td>5.01</td>
<td>C_{15}H_{24}</td>
<td>204</td>
<td>0.96</td>
</tr>
<tr>
<td>Synonyms: (-)-â-Cubebene</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>11. Bicyclo[3.1.1]heptane-2-methanol, 6,6-dimethyl, [15-(1'2,2,5a)]- Synonyms: cis-Myrtanol</td>
<td>5.16</td>
<td>C_{15}H_{26}O</td>
<td>154</td>
<td>1.75</td>
</tr>
</tbody>
</table>
12. Caryophyllene
Synonyms: α-Caryophyllene, trans-Caryophyllene
5.837 C_{15}H_{24} 204 2.63

13. β-Cubebene
β-Cuvebene
6.04 C_{15}H_{24} 204 1.17

14. α-Humulene
Synonyms: 1,4,8-Cycloundecatriene, 2,6,6,9-tetramethyl-,
(E,E,E)-
6.55 C_{15}H_{24} 204 2.68

15. Epizonarene
Synonyms: no synonyms
6.887 C_{15}H_{24} 204 0.63

16. 1,6-Cyclodecadiene, 1-methyl-5-methylene-8-(1-
methyleneethyl)-, [E,E,E]-
Synonyms: 1.Germacrene D
7.193 C_{15}H_{24} 204 10.65

17. (+)-Epi-bicyclosesquiphellandrene
Synonyms: 1.1-Isopropyl-4-methyl-7-methylene-1,2,3,4,4a,5,6,7-
octahydronaphthalene
7.33 C_{15}H_{24} 204 0.78

18. Naphthalene, 1,2,4a,5,6,8a-hexahydro-4,7-
dimethyl-1-(1-methyleneethyl)-, (1k,4a,8a)-
Synonyms: α-Muurolene
7.777 C_{15}H_{24} 204 1.26

19. Naphthalene, 1,2,3,4,4a,5,6,8a-octahydro-7-methyl-
4-methylene-1-(1-methyleneethyl)-, (1a,4a,8a)--
Synonyms: ã-Cadinene
17.18 C_{15}H_{24} 204 3.81

20. α-Cadinol
No synonyms 21.687 C_{20}H_{34}O 230 6.87

21. Cycloheptane, 4-methylene-1-methyl-2-(2-methyl-
1-propen-1-yl)-1-vinyl-
no synonyms.
21.687 C_{15}H_{24}O 230 6.87

Table 2: Correlation of the DPPH optical density and concentration.

<table>
<thead>
<tr>
<th>Optical density (D)</th>
<th>Concentration (c)</th>
</tr>
</thead>
<tbody>
<tr>
<td>c = 20.06*D + 0.996</td>
<td></td>
</tr>
<tr>
<td>0.947</td>
<td>20</td>
</tr>
<tr>
<td>0.05</td>
<td>2</td>
</tr>
</tbody>
</table>

Table 3: The relationship between the optical density (D) and concentration (C) [(x 10^{-5}) g/l] of the inhibition DPPH (n=5, $\bar{x}$ – mean, $\sigma_r$ standard error of the mean).

Analytical samples of the raw materials.

<table>
<thead>
<tr>
<th>Extract (mg)</th>
<th>The time of the inhibition DPPH (min) $\bar{x} \pm \sigma_r$</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
</tr>
<tr>
<td>D-4</td>
<td>0.443±0.008</td>
</tr>
<tr>
<td></td>
<td>0.236±0.002</td>
</tr>
<tr>
<td>D-8</td>
<td>0.422±0.008</td>
</tr>
<tr>
<td></td>
<td>0.139±0.003</td>
</tr>
<tr>
<td>C-4</td>
<td>9.886±0.164</td>
</tr>
<tr>
<td></td>
<td>5.74±0.04</td>
</tr>
<tr>
<td>C-8</td>
<td>9.461±0.157</td>
</tr>
<tr>
<td></td>
<td>3.786±0.068</td>
</tr>
<tr>
<td><strong>D-2</strong></td>
<td>0.245±0.002</td>
</tr>
<tr>
<td></td>
<td>0.092±0.002</td>
</tr>
<tr>
<td><strong>D-8</strong></td>
<td>0.118±0.002</td>
</tr>
<tr>
<td></td>
<td>0.08±0.002</td>
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<tr>
<td><strong>C-2</strong></td>
<td>5.905±0.033</td>
</tr>
<tr>
<td></td>
<td>2.835±0.044</td>
</tr>
<tr>
<td><strong>C-8</strong></td>
<td>3.357±0.038</td>
</tr>
<tr>
<td></td>
<td>2.598±0.035</td>
</tr>
</tbody>
</table>

**D-2, **D-4, D-8- the values of the optical densities of the extracts with concentrations of 2 mg, 4 mg and 8 mg, respectively.

**C-2, **C-4, C-8- the values of the concentrations of experimented herbal extracts 2 mg, 4 mg and 8 mg, respectively.

Figure 1: Mass spectrum and structure of phytocomponents identified by GC-MS in the essential oil of leaves Cotinus coggygria.
Table 4: The antioxidant activity of the dry 50% methanol extracts of leaves and branches of C. coggygria (n=5, $\mu$ - mean, $\sigma$ - standard error of the mean).

<table>
<thead>
<tr>
<th>Analytical samples of the raw materials.</th>
<th>Extract (mg)</th>
<th>The time of the inhibition DPPH, min and antioxidant activity (%)</th>
<th>$IC_{50}$ (x 10$^{-5}$) g/l</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>$1$</td>
<td>$5$</td>
</tr>
<tr>
<td>branches of C. coggygria</td>
<td>4</td>
<td>50.57±0.82</td>
<td>71.30±0.23</td>
</tr>
<tr>
<td></td>
<td>8</td>
<td>52.70±0.79</td>
<td>81.07±0.34</td>
</tr>
<tr>
<td>leaves of C. coggygria</td>
<td>2</td>
<td>70.47±0.16</td>
<td>85.83±0.18</td>
</tr>
<tr>
<td></td>
<td>8</td>
<td>83.22±0.19</td>
<td>87.03±0.17</td>
</tr>
</tbody>
</table>

Figure 2: Correlation graph of the DPPH optical density and concentration.

Figure 3: The relationship between the optical density (D) and concentration (C) of the experimented extracts for corresponding concentrations 2mg, 4mg and 8mg. SV4 extract from shrubs - the concentration 4mg; SV8 extract from shrubs - the concentration 8mg; SL2 extract from leaves - the concentration 2mg; SL8 extract from leaves - the concentration 8mg.
The highest values (IC 50 13.95±0.32 and 13.85±0.27 (x 10^-5) g/l) of extracts antioxidant activity. The highest IC50 was recorded in the branch extracts, which showed low the rate of reaction. The Leaves extract has the highest activity. The antioxidant activity, with the indicators differ from each other in the range from 64 ± 0.49 to 13.95 ± 0.32 (x 10^-5) g / l.

The extract of the smoke tree branches showed the lowest activity, the antioxidant activity index of which was 50.57 ± 0.82 in the first 1 minute at a concentration of 4 mg of DPPH.

It is noteworthy that at all concentrations of the extract of the leaves and branches, the same antioxidant activity is observed during the 5th 20th minutes. That is, an unreliable difference is noticed between the measures (p≥0.05). It should be noted that branch extracts have less antioxidant activity than leaf extracts. The concentrations of antioxidants was also calculated during the study (IC50) that inhibited DPPH radicals by 50%. As it is seen from the table IC50 values were in the range from 64 ± 0.49 to 13.95 ± 0.32 (x 10^-5) g / l.

Thus, the study results showed that the extracts of the branches and leaves of smoke trees growing in Armenia have a pronounced antioxidant activity, with the indicators differ from each other in the rate of reaction. The Leaves extract has the highest activity. The highest IC50 was recorded in the branch extracts, which showed low antioxidant activity.

The highest values (IC_50 13.95±0.32 and 13.85±0.27 (x 10^-5) g/l) of extracts were received from the branches C. coggymria for the concentrations 4mg and 8 mg respectively had the smallest of antioxidant activity.

CONCLUSIONS

Analyses of the volatile constituents from the leaves of Cotinus coggymria collected from Yerevan indicate Germacrene D, Linalool formate, α-Terpineol sesquiterpenes and diterpene alcohol Thunbergol which are the predominant components of Cotinus coggymria. This work provides the first report of the analysis of essential oils of Cotinus coggymria from Armenia. At the same time, the results of the research showed that the leaves and shrubs might be further applied as a new natural antioxidant ingredient in the medicine.

ACKNOWLEDGMENTS

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CONFLICTS OF INTEREST

The authors declare no conflicts of interest.

REFERENCES


**GRAPHICAL ABSTRACT**

**ABOUT AUTHORS**

Naira B. Chichoyan, head of the Department of Pharmacognosy, Yerevan State Medical University, Doctor of Pharmaceutical Science, Associate Professor. Supervisor of a research project “The pharmacognostic research of wild-growing and cultivating medicinal plant species of Republic of Armenia and Artsakh flora.” Author of more than 80 scientific works, 3 manuals and a monograph.

Naira K. Shaboyan, 2014-2019- attended courses at University of Basel, Faculty of Natural Sciences, Department of Pharmacology, Switzerland. 2020-up to this day works at Galenica Pharmaceutical Company (Galenica AG Pharmaceuticals Bern, Switzerland). The research project is “The pharmacognostic research of Armenian flora.”

Armenuhi V. Moghrovyan, Senior lecturer at the Department of Pharmacognosy, YSMU, after Mkhitar Heatsi, PhD. The research project is “The pharmacognostic research of Armenian flora.” Particularly the scope of scientific interest is “The pharmacognostic and biological activity analysis of Oregano ordinary (Herba Origani vulgaris) growing wildly in Armenian flora.”

Karine H. Dumanyan, Associate Professor at the Department of Pharmacognosy, YSMU, after Mkhitar Heatsi, PhD. The research project is “The pharmacognostic research of Armenian flora.” Within the framework of the research project she implements the identification of endemic plants. More than 30 years teaches Botany in Pharmacy Faculty.


Nelli H. Ghukasyan, lecturer at the Pharmacognosy Department, YSMU, after Mkhitar Heatsi. The research project is “The pharmacognostic research of Armenian flora.” Also, has an interest on the phytochemical analysis of bulbous-rooted chervil growing wildly in different regions of Armenia. More than 10 years of multilingual teaching.

Astghik A. Altunyan, lecturer at the Department of Pharmacognosy, YSMU, after Mkhitar Heratsi. The research project is “The pharmacognostic research of Armenian flora.” Efficient, hard-working, knowledgeable and technically skilled employee who works well both individually and in a team environment.

Naira I. Arshakyan, assistant at the Department of Pharmacognosy, YSMU, after Mkhitar Heratsi. The research project is “The pharmacognostic research of Armenian flora.” Author of some scientific articles.

Arshaluys M. Ghazaryan, graduated from YSMU, after Mkhitir Heratsi with honours in 2016 and started working as an assistant at the Department of Pharmacognosy. She is a resident of the Department.

Greta R. Ulikhanyan, Lecturer at the Department of «Medical physics » of the Yerevan State Medical University after. M. Heratsi. Scientific interests - application physico-chemical methods for standardization raw materials of plants using in the medico-pharmaceutical practice.

Aghavni V. Ginosyan, PhD in Chemical engineering eMBA in Healthcare Management, Director of Analytical Laboratory branch of «Scientific Center of Drug and Medical Technology Expertise after Academician E.Gabrielyan» JSC. Lecturer, tutor on Analytical Method Validation, Stability Testing and Quality Management Systems. Yerevan State University, Institute of Pharmacy, Department of Pharmacy.

Ani S. Dadayan, Secretary of Scientist’s Board of Institute of Pharmacy YSU, Assistant of Department of Pharmtechnology and Pharmacy Economics & Management, PhD in Chemistry. Scope of scientific interests - Syntheses of new modified chiral auxiliaries and their amino and dehydroamino acid complexes and investigation in asymmetric biomimetic reactions for the syntheses of α- and β- substituted non-proteinogenic (R)- and (S)- α - mino acids. Production of Galenic and New-galenic preparations and herbal origin oils.