# Antimalaria Activities of Several Active Compounds from Medicinal Plants

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#### ABSTRACT

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The growth and spread of resistance to all first-line antimalarial drugs used in the treatment of malaria today has caused many problems in eradicating malaria in various countries in the world. Researchers have begun to look for candidates for new antimalarial drug compounds derived from natural products that have been scientifically proven to have antimalarial activity. This is done to replace antimalarial drugs that are currently experiencing resistance. Some active compounds that have been successfully isolated from various plants, including curcumin, kaempferol, piperine, andrographolide,  $\alpha$ -mangostin, catechin, luteolin and betulinic acid, have been scientifically tested to have antimalarial activity with different mechanisms of action.

Key words: Malaria, Antimalarial, Curcumin, Kaempferol, Piperine, Andrographolide,  $\alpha$ -Mangostin, Catechin, Luteolin, Betulinic acid.

## **INTRODUCTION**

Malaria is still a health problem in the world, with 229 million cases of malaria worldwide and more than 400,000 people suffering from malaria dying each year.1 The high number of malaria cases is caused by the emergence of various obstacles in eradicating malaria, including the resistance of malaria parasites to available antimalarial drugs, increased Anopheles mosquito immunity to chemicals, and the discovery of side effects from these antimalarial drugs.2 The growing and widespread resistance to all the first-line antimalarial drugs used in the treatment and prevention of malaria has now caused many problems in the malaria prevention programme.<sup>3</sup> Research to obtain new antimalarial drugs, both synthetic drugs and those derived from natural materials, is continuing, one of which is through the exploration of active compounds from natural materials, especially medicinal plants that have traditionally been used by the community to treat malaria in various endemic areas across the world. The aim is to find new antimalarial compounds that have mild side effects with low toxicity, so that they do not harm the patient.<sup>4</sup> This review article will discuss the active compounds and antimalarial activity of several medicinal plant isolates.

## **CURCUMIN**

Curcumin (1,7-bis (4'hydroxy-3-methoxyphenyl) 1,6-heptadiene, 3,5-dion) is a phytochemical component found mainly in *Curcuma longa* and *Curcuma zanthorrhiza*. Curcumin is often used as a medicine and traditional ingredient to treat various diseases in several countries.<sup>5</sup> Curcumin has several pharmacological activities such as anti-inflammatory,<sup>6-7</sup> antioxidant,<sup>8</sup> anticancer,<sup>9</sup> anticoagulant,<sup>10</sup> hepatoprotective<sup>11</sup> and antibacterial.<sup>12</sup> In addition, curcumin has been shown to have antimalarial activity against *Plasmodium* species.<sup>13-14</sup> The antimalarial activity

of curcumin is caused by its ability to inhibit histone acetyltransferase from P. falciparum, causing parasitic cell damage.14 Other studies also report that curcumin can interact with sarco/endoplasmic reticulum Ca2+ATPase (SERCA) in P. falciparum which causes the inhibition of a Ca2+ ion transporter called PfATP6.<sup>15</sup> Meanwhile, from in vitro antimalarial activity testing,  $IC_{50}$  values obtained from curcumin were 5 µM, whereas at concentrations of 50 µM, curcumin was able to reduce the viability of P. falciparum, which caused a decrease in parasitic cell proliferation. Other results also reported that at concentrations of 5 µg/ml and 50 µg/ml, curcumin was able to inhibit 79.6% and 100% growth of P. falciparum.<sup>16</sup> In addition, in antimalarial testing in vivo with a dose of 100 mg/kg, curcumin was able to inhibit the growth of parasites >80%.13 Waknine-Grinberg et al. reported that the administration of curcumin at a dose of 50 mg/kg, twice daily for 6 days after infection in C57Bl/6 mice infected with P. berghei ANKA, can prevent cerebral malaria (CM) and delay death by up to 10 days.17 In addition, it has been shown in vitro that curcumin reduces the production of proinflammatory cytokines IL6, TNF and IL12p40 in PBMC primed with trophozoites/ schizonts stages of P. falciparum and downregulated the expression of VCAM1, ICAM1 and E-selectin in TNF-activated human endothelial cells.18 In addition, other studies report that curcumin can inhibit thrombin-activated platelet adhesion to brain microvascular endothelial cells in vitro19 which is thought to accumulate in brain microvasculature in murine and pediatric CM patients.20

# KAEMPFEROL

Kaempferol (3,4' 5, 5,7-tetrahydroxyflavone) is a natural flavonol, a flavonoid derivative, found in various plants and plant-derived foods such as kale, beans, tea, spinach and broccoli.<sup>21-22</sup> Kaempferol has been reported to have high antioxidant properties, which are able to inhibit oxidative stress resulting from the formation of reactive oxygen species (ROS)

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that occur during malaria infections.<sup>23</sup> ROS and oxidative stress are also produced by monocytes and neutrophils which are activated during malarial infection, which cause haemoglobin to degrade,<sup>24-25</sup> thus there is a correlation between antioxidant activity and antimalarial activity of kaempferol.<sup>26</sup> In addition, the antimalarial activity of kaempferol is also caused by its ability to inhibit glycogen synthase kinase-3β (GSK3β) from malaria parasites. GSK3ß is reported to play an important role in the host response to malaria parasitic infections.<sup>27-28</sup> By inhibiting GSK3ß in mammals, it can also significantly inhibit the activity of recombinant P. falciparum (PfGSK3B) protein.29 Somsak et al., reported that at a dose of 20 mg/kg, kaempferol was able to inhibit P. berghei by 52.89%, 40.80%, and 36.63% for each chemosuppressive, chemoprophylactic, and curative trial.<sup>30</sup> Meanwhile, Barliana et al., reported that kaempferol was able to inhibit the growth of P. falciparum which is resistant to chloroquine in vitro after 24 hours of therapy with  $IC_{_{50}}\,106\,\mu M.^{_{31}}$  In addition, ka<br/>empferol at a dose of 2,000 mg/kg daily for 30 days does not produce signs of toxicity in the form of hepatotoxic, nephrotoxic, haematotoxic, or death in test animals.<sup>30</sup>

## PIPERINE

Piperine (1-[5-(1,3-benzodioxol-5-yl)-1-oxo-2,4-pentadienyl]piperidine) is the main amide compound isolated from the *Piper chaba* Hunt fruit,<sup>32</sup> *Piper nigrum* L. (black pepper) and *Piper longum* L. (long pepper).<sup>33</sup> Piperine has been shown to have several pharmacological activities such as antioxidants,<sup>34</sup> anticancer,<sup>35</sup> antidepressive,<sup>36</sup> hepatoprotective,<sup>37</sup> anti-asthmatic,<sup>38</sup> antipyretic,<sup>33</sup> anti-inflammatory,<sup>39</sup> analgesic, and anticonvulsant.<sup>40</sup> In addition, piperine was also reported to have antimalarial activity in vitro against *P.falciparum* 3D7 strains (chloroquine-sensitive) and K1 (chloroquine-resistant) with IC<sub>50</sub> against 3D7 and K1 *P.falciparum* respectively at 111.5  $\mu$ M and 59  $\mu$ M.<sup>32</sup> The antimalarial activity of piperine is due to its ability to inhibit the modulation of expression of all P. falciparum resistance genes,32 including the P. falciparum multidrug resistance protein 1 gene (pfmrp1), the pfmrp1 protein coding gene that is a member of the ABC transporter superfamily located on chromosome 1,41 where this protein has been reported to be involved in the export of folate from malaria parasites into red blood cells,42 P. falciparum multidrug resistance gene 1 (*pfmdr*1) is a homologous P-glycoprotein coding gene located in the parasite digestive vacuole membrane, which is thought to play a role in modulation of antimalarial drug resistance <sup>43</sup> and *P*. falciparum chloroquine resistance transporter (pfcrt), a protein-coding gene that is localised to the parasitic digestive vacuole membrane and contains 10 domains that line the membrane in the erythrocytic stage.44 Piperine might be a promising candidate for the development of new antimalarial drugs, due to its antimalarial potential and low risk of resistance in modulating the P. falciparum resistance gene that causes the development of resistance.<sup>32</sup>

## ANDROGRAPHOLIDE

Andrographolide (3-[2-[Decahydro-6-hydroxy-5-(hydroxymethyl)-5,8a-dimethyl-2-methylene-1-napthalenyl]ethylidene]dihydro-4hydroxy-2(3H)-furanone) is a lactone diterpene compound isolated from the *Andrographis paniculata* (Burm. F) Nees plant, commonly known as the "*king of bitters*".<sup>45</sup> Andrographolide has been reported to have several pharmacological activities such as anti-inflammatory, antitumor, antidiabetic, and hepatoprotective.<sup>46</sup> In addition, andrographolide has been shown to have antimalarial activity in vitro and in vivo.<sup>47</sup> The antimalarial activity of andrographolide is due to its ability to inhibit nuclear kappa B (NF- $\kappa$ B) transcription factors induced when erythrocytes are infected with *P. falciparum* <sup>48</sup> by blocking the binding of NF- $\kappa$ B oligonucleotides to nuclear proteins <sup>49</sup> where this pathway is an inflammatory pathway in the human cerebral



**Figure 1**. Parasite targets for intervention. **A**) Heme detoxification pathways in the intra-erythrocytic cycle and redox homeostasis in infected red blood cell: α-mangostin inhibit heme degradation in the parasite food vacuole. **B**) The electron transport chain (ETC) of mitochondria involve in the bio-activation of catechin induces the membrane depolarization and disrupt the normal function of mitochondria, through the production of ROS. Furthermore, andrographolide inhibits thioredoxin reductase an enzyme maintains the redox equilibrium in plasmodium species. **C**) Inactivation of SERCA which is responsible for protein synthesis in parasite by curcumin and betulinic acid. **D**) Inhibition of type II fatty acid biosynthesis pathway in *P. falciparum* by luteolin.

Active Compounds	Chemical Structures	Types of Malaria Parasites	Study Design	Mechanism of Action	References
Curcumin	HO O CH <sub>3</sub> OOOCH <sub>3</sub> OH	<i>1. P. falciparum</i> strain 3D7 and D10. <i>2. P. falciparum</i> strain DD2 and 7G8. <i>3. P. berghei</i> strain ANKA.	In vitro and in vivo	Interacts with sarco/ endoplasmic reticulum $Ca^{2+}ATPase$ (SERCA) in <i>P. falciparum</i> which causes inhibition of $Ca^{2+}$ ion transporter.	15
Kaempferol	НО ОН ОН ОН	1. P. falciparum strain 3D7. 2. P. berghei strain ANKA.	In vitro and in vivo	Inhibits oxidative stress resulting from the formation of reactive oxygen species (ROS).	23
Piperine		1. P. falciparum strain 3D7. 2. P. falciparum strain K1.	In vitro	Inhibits the modulation of expression of all <i>P. falciparum</i> resistance genes.	32
Andrographolide	HOW HO	1. P. falciparum strain 3D7. 2. P. berghei strain ANKA.	In vitro and in vivo	Reduction in GSH concentration and the activity of the thioredoxin reductase (TrxR) enzyme.	50
α-Mangostin	OH O HO HO OHO OH OH OH OH	<ol> <li>P. falciparum strain K1.</li> <li>P. falciparum strain FCR3.</li> <li>P. falciparum strain 3D7.</li> <li>P. berghei strain ANKA.</li> </ol>	In vitro and in vivo	Inhibits haemoglobin degradation process that occurs in the food vacuole of <i>P. falciparum</i> .	53, 58, 59
Catechin	но он он он	1. P. falciparum strain DD2. 2. P. falciparum strain 3D7.	In vitro	Induces oxidative stress to <i>P. falciparum</i> .	62,63
Luteolin	HO OH OH	1. P. falciparum strain 3D7. 2. P. falciparum strain 7G8.	In vitro	Inhibits FabG, FabZ and FabI enzymes which involves in parasite fatty acid biosynthesis.	69
Betulinic acid	но Н	1. P. falciparum strain K1. 2. P. falciparum strain T9-96. 3. P. falciparum strain 3D7.	In vitro	Ability to bind the $PfATP6$ protein and sarco/ endoplasmic reticulum $Ca^{2+}ATPase$ (SERCA) in mammals.	75-77

#### Table 1. Active compounds and working mechanisms as antimalaria.

endothelium.47-48 Meanwhile, Risdawati reported that the antimalarial andrographolide activity was caused by this compound being able to disrupt the parasitic antioxidant defence system as evidenced by the reduction in GSH concentration and the activity of the thioredoxin reductase (TrxR) enzyme.<sup>50</sup> Kusumawardhani, et al., reported that the antimalarial activity of standardised A. paniculata leaf extracts (andrographolide levels; 10.82 ± 0.37%) against P. berghei in vivo obtained ED<sub>50</sub> of 12.2223 mg of standardised A. paniculata /KgBW leaf extract, equivalent to 1,320 mg of andrographolide compounds.<sup>51</sup> Meanwhile, Sachdeva reported that andrographolide at a dose of 0.1 mg/ml was able to inhibit the growth of malaria parasites in vitro by 53.9%.<sup>52</sup> In addition, Mishra et al., reported that andrographolide showed high malaria parasitic inhibitory activity in rat models in vivo when given with curcumin or artesunate via the intraperitoneal route, and without showing any symptoms of toxicity after administration of the therapy.47

## **α-MANGOSTIN**

 $\alpha$ -mangostin (1,3,6-trihidroksi-7-metoksi-2,8-bis (3metil-2-butenil)-9H-xanten-9-on) is the main xanthone compound found in the hull of the *Garcinia mangostana* L fruit.<sup>53</sup> This compound has been reported to have pharmacological activities including anti-inflammatory, anti-tumor, cardioprotective, antidiabetic, antibacterial, antifungal, antiparasitic, antioxidant and anti-obesity agents.<sup>54</sup> In addition,  $\alpha$ -mangostin is also reported to have antimalarial activity.<sup>53</sup> Antimalarial activity is suspected of  $\alpha$ -mangostin which is a hydroxyxanthones derivative due to its ability to form complexes that are soluble with heme compounds so that it inhibits the formation of parasitic hemozoin, a process in which the malaria parasite protects itself against the effects of heme toxins released after digesting haemoglobin.<sup>55-56</sup> The effect was caused by this compound having an alkyl group located in positions 3 and 6 which interacts with nitrogen atoms contained in heme propionate compounds thus forming ionic bonds which cause an increase in antiplasmodial activity.<sup>56-57</sup> This is in accordance with the research of Kuncoro, et al., which reported that with α-mangostin able to inhibit the development of trophozoite stages into schizon stages in P. falciparum, where at the trophozoite stage an increase in haemoglobin uptake, with inhibition of haemoglobin uptake, would result in inhibition haemoglobin degradation process that occurs in the food vacuole of P. falciparum.58 Mahabusarakam et al., reported that active α-mangostin inhibits the growth of *P. falciparum* strain K1 (chloroquine and pyrimethamine-resistant) in vitro with IC<sub>50</sub> of 17 µM.53 This is in accordance with the results of research by Upegui et *al.*, who reported that  $\alpha$ -mangostin was more active in inhibiting the growth of P. falciparum strain FCR3 (chloroquine-resistant) in vitro with IC<sub>50</sub> of 0.2±0.01  $\mu$ M compared to  $\delta$ -mangostin with IC<sub>50</sub> is 121.2  $\pm$  1.0  $\mu$ M, but  $\delta$ -mangostin is more active in inhibiting the growth of *P. falciparum* strain 3D7 (chloroquine-sensitive) in vitro with  $IC_{50}$  of 12.40  $\pm$  1.0  $\mu M$  compared to a-mangostin with IC  $_{_{50}}$  of 36.10  $\pm$  4.9  $\mu M.^{_{59}}$ In addition, Upegui et al., also reported that at doses of 100 mg/kg/ day twice daily for 7 days,  $\alpha$ -mangostin and  $\delta$ -mangostin were able to reduce the growth of P. berghei respectively by 80.7% and 79.9 %.59 These findings can provide a basis for further research on a-mangostin as a potential compound to be developed into antimalarial drugs.<sup>59</sup>

#### CATECHIN

Catechin (2R,3S)-2-(3,4-dihydroxyphenyl)-3,4-dihydro-2H-chromene-3,5,7-triol) is the main bioactive compound found in green tea (Camellia sinensis).60 Catechin is reported to have several pharmacological activities including antioxidant, anti-microbial, anti-inflammatory, anti-viral, anti-cancer and anti-allergic properties.<sup>61</sup> In addition, this compound is also reported to have antimalarial activity against P. falciparum.62-63 The antimalarial activity of catechin is due to its ability to induce oxidative stress to *P. falciparum*.<sup>63</sup> Goyal *et al.*, reported that Plasmodium is very susceptible to oxidative stress, where an increase in oxidative stress will cause a redox imbalance.<sup>64</sup> Meanwhile, the redox system plays an important role in the survival of Plasmodium in the host cell, by disturbing the redox balance will affect the survival of Plasmodium during the intraerythrocytic stage.<sup>65</sup> Budiman et al., reported that active catechin inhibited the growth of P. falciparum with  $IC_{50}$  of 0.734  $\mu$ M,<sup>62</sup> this is in accordance with the research of Abdulah et al., who reported that catechin could inhibit the growth of P. falciparum strain DD2 (chloroquine-resistant) with  $IC_{_{50}}$  198  $\mu M$  after 24 hours of treatment.  $^{63}$ Although this is a preliminary study, the results of this study can provide information for further research on catechin as a potential compound as an antimalarial drug.63

## LUTEOLIN

Luteolin (3',4',5,7-tetrahydroxyflavone) is a common flavonoid that exists in many types of plants, including herbs, vegetables and medicinal plants.<sup>66</sup> Luteolin is reported to have several pharmacological activities including anti-inflammatory, antioxidant, antidiabetic, antimicrobial, and anticancer.<sup>67</sup> In addition, luteolin was also reported to have antimalarial activity against P. falciparum strain 3D7 (chloroquinesensitive) and P. falciparum strain 7G8 (chloroquine-resistant) with  $IC_{50}$  of 11 ± 1  $\mu$ M and 12 ± 1  $\mu$ M, respectively.<sup>68</sup> The antimalarial activity of luteolin is due to its ability to inhibit the development of parasitic growth at the stage of young trophozoites, so the parasite cannot complete the full intraerythrocytic cycle.68 This compound is also reported to Inhibits FabG, FabZ and FabI enzymes which are involved in parasite fatty acid biosynthesis.<sup>69</sup> In vitro experiments show that luteolin can inhibit the activation of the nuclear factor kappa B (NF-KB) induced by lipopolysaccharide (LPS) and members of the family of mitogen-activated protein kinase (MAPK) ERK, p38, and JNK.<sup>70-71</sup> NF-KB and MAPK are the two main pathways involved in the activation of macrophages in epithelial cells and stromal tissue in

response to inflammatory mediators such as  $\text{TNF}\alpha$  and  $\text{ILs.}^{72}$  It is also an inflammatory pathway in the human cerebral endothelium.<sup>47-48</sup> The results of these studies can make the initial hypothesis that leuteolin can be used as therapy in CM patients, but further research must still be done to see the effectiveness of leuteolin in the treatment of malaria, especially CM.

#### **BETULINIC ACID**

Betulinic acid ((3β)-3-Hydroxy-lup-20(29)-en-28-oic acid) is a naturally occurring pentacyclic triterpenoid found in the skin of several plant species, especially white birch (Betula pubescens).73 Betulinic acid has been shown to have several pharmacological activities such antidiabetic, anti-dyslipidemia, anti-inflammatory, antiviral, as anticancer, parasiticidal and anti-infectious.74 In addition, this compound was also reported to have antimalarial activity against P. falciparum strain K1 (chloroquine-resistant) and P. falciparum strain T9-96 (chloroquine-sensitive) with IC<sub>50</sub> of 19.6 mg/ml and 25.9 mg/ml, respectively.75 This is in accordance with the results of research by Silva et al., who reported that active betulinic acid inhibits the growth of P. falciparum strain 3D7 (chloroquine-sensitive) with  $IC_{50}$  of 18 ± 0.17 µM.76 Antimalarial activity of betulinic acid is thought to be caused by its ability to bind the PfATP6 protein and sarco/endoplasmic reticulum Ca<sup>2+</sup>ATPase (SERCA) in mammals so that it interferes with selectivity between parasites and hosts.77 The development of betulinic acid as an antimalarial for the future is very necessary, given that the antimalarial mechanism of this compound is similar to the drug artemisinin by inhibiting the sarco/endoplasmic reticulum Ca2+ATPase (SERCA) ortholog PfATP6 in P. falciparum, where PfATP6 is the only SERCA type Ca2+ATPase sequence in the parasitic genome.78-79

# CONCLUSION

All of these isolates have been scientifically proven to have antimalarial activity, where each compound has a different mechanism of action in inhibiting the growth of malaria parasites. However, further research must be done to see the effectiveness of these compounds as antimalarials, so that they can be used in the latest malaria treatment, which has now shown resistance to first-line malaria treatment.

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#### CONFLICTS OF INTEREST

The authors declare that there is no conflict of interest regarding the publication of this article.

## ABBREVIATIONS

**SERCA:** sarco/endoplasmic reticulum Ca<sup>2+</sup>ATPase; **CM**: cerebral malaria; **IL-6**: interleukin-6; **TNF**: tumor necrosis factor; **IL-12p40**: interleukin-12p40; **PBMC**: peripheral blood mononuclear cell; **VCAM-1**: vascular cell adhesion molecule 1; **ICAM-1**: intercellular adhesion molecule 1; **ROS**: reactive oxygen species; **GSK3β**: glycogen synthase kinase-3β; **PfGSK3β**: *Plasmodium falciparum* glycogen synthase kinase-3β; **Pfmrp1**: *P. falciparum* multidrug resistance protein 1; **Pfmdr1**: *P. falciparum* multidrug resistance gene 1; **Pfcrt**: *P. falciparum* chloroquine resistance transporter; **NF-kB**: nuclear factor-kappaB; **GSH**: glutathione; **TrxR**: thioredoxin reductase enzyme; **LPS**: lipopolysaccharide; **MAPK**: mitogen-activated protein kinase; **ERK**: extracellular signal-regulated kinases; **JNK**: c-Jun N-terminal kinases; **Hb**: haemoglobin; **FP IX**: ferroprotoporphyrin IX (Heme); **GSH**: glutathione; **SOD**: superoxide dismutase; **NPP**: new permeation pathway; **ACP**: acyl carrier protein; **FabD**: malonylCoA-ACP

transacylase; FabH:  $\beta$ -ketoacyl-ACP synthase III; FabG:  $\beta$ -ketoacyl-ACP reductase; FabA or FabZ:  $\beta$ -hydroxyacyl-ACP dehydratase; FabI: enoyl- ACP reductase; FabB and/or FabF:  $\beta$ -ketoacyl-ACP synthase I and II; CUR: curcumin; KAEMP: kaempferol; PIR: piperine; ADG: andrographolide;  $\alpha$ -MG:  $\alpha$ -mangostin; CAT: catechin; LUT: luteolin; BA: betulinic acid.

# REFERENCES

- WHO. World Malaria Report 2020. World Health Organization 2020. [cited 2021 Jun 17]. Available from: https://www.mmv.org/ newsroom/publications/world-malaria-report-2020?gclid=EAlalQob ChMIz6ycsoOn7AIVmiQrCh1wPAw0EAAYAiAAEgKcEfD\_BwE
- Utami WS, Nuri, Armiyanti Y. Effect *Tithonia diversifolia* (Hemsley) A. Gray) ethanol extract as antimalarial on mice strain Balb/c before and after infected by *Plasmodium berghei*. J Med Plant. 2012;1(5):56-66.
- Burke E, Deasy J, Hasson R, McCormack R, Randhawa, V, Walsh, P. Antimalarial drug from nature. J Trinity Student Med. 2003:1-8.
- Mustofa. Molekul antimalaria alami: potensi dan tantangan pengembangannya sebagai obat baru untuk malaria. Trad Med J. 2003;8(26):8-18.
- Da'l M, Fajria A, Utami W. Sintesis senyawa analog kurkumin 3,5-BIS-(4'-hidroksi-3'-metoksi-benzilidin)-4-piperidin (monohidrat hidroklorida) dengan katalis HCl. Pharmacon. 2010;11(1):33-8.
- Chainani-Wu N. Safety and anti-inflammatory activity of curcumin: a component of tumeric (*Curcuma longa*). J Altern Complement Med. 2003;9(1):161-68.
- Edwards RL, Luis PB, Varuzza1 PV, Joseph AI, Presley SH, Chaturvedi R, *et al.* The anti-inflammatory activity of curcumin is mediated by its oxidative metabolites. J Biol Chem. 2017;292(52):1-22.
- Ak T, Gulcin I. Antioxidant and radical scavenging properties of curcumin. Chem Biol Interact. 2008;174:27-37.
- Allegra A, Innao V, Russo S, Gerace D, Alonci A, Musolino C. Anticancer activity of curcumin and its analogues: preclinical and clinical studies. Cancer Invest. 2016;35(1):1-22.
- 10. Dong-Chan K, Sae-Kwang K, Jong-Sup B. Anticoagulant activities of curcumin and its derivative. BMB Rep. 2012;45(4):221-26.
- Samuhasaneeto S, Thong-Ngam D, Kulaputana O, Suyasunanont D, Klaikeaw N. Curcumin decreased oxidative stress, inhibited NF-KB activation, and improved liver pathology in ethanol-induced liver injury in rats. J Biomed Biotechnol. 2009:1-8.
- 12. Akbik D, Ghadiri M, Chrzanowski W, Rohanizadeh R. Curcuminas a wound healing agent. Life Sci. 2014;116(1):1-7.
- Reddy RC, Vatsala PG, Keshamouni VG, Padmanaban G, Rangarajan P. Curcumin for malaria therapy. Biochem Biophys Res Commun. 2005;326:472-74.
- Cui L, Miao J. Cytotoxic effect of curcumin on malaria parasite *Plasmodium falciparum*: inhibition of histone acetylation and generation of reactive oxygen species. Antimicrob Agents Chemother. 2007;51:488-94.
- Ji HF, Shen L. Interactions of curcumin with the PfATP6 model and the implications for its antimalarial mechanism. Bioorg Med Chem Lett. 2009;19:2453-55.
- Dohutia C, Chetia D, Gogoi K, Bhattacharyya DR, Sarma K. Molecular docking, synthesis and in vitro antimalarial evaluation of certain novel curcumin analogues. Braz J Pharm Sci. 2017;53(4):1-14.
- Waknine-Grinberg JH, McQuillan JA, Hunt N, Ginsburg H, Golenser J. Modulation of cerebral malaria by fasudil and other immunemodifying compounds. Exp Parasitol. 2010;125:141-6.

- Mimche P, Taramelli D, Vivas L. The plant-based immunomodulator curcumin as a potential candidate for the development of an adjunctive therapy for cerebral malaria. Malar J. 2011;10(1):1-9.
- Zhang L, Gu ZL, Qin ZH, Liang ZQ. Effect of curcumin on the adhesion of platelets to brain microvascular endothelial cells in vitro. Acta Pharmacol Sin. 2008;29:800-7.
- Faille D, El-Assaad F, Alessi MC, Fusai T, Combes V, Grau GE. Platelet-endothelial cell interactions in cerebral malaria: the end of a cordial understanding. Thromb Haemost. 2009;102:1093-102.
- Calderón-Montaño JM, Burgos-Morón E, Pérez-Guerrero C, LópezLázaro MA. Review on the dietary flavonoid kaempferol. Mini Rev Med Chem. 2011;11(4):298-344.
- Holland T, Agarwal P, Wang Y, Leurgans SE, Bennett DA, Booth SL, et al. Dietary flavonols and risk of alzheimer dementia. Neurology. 2020;94(16):1-8.
- 23. Wallqvist A, Fang X, Tewari SG, Ye P, Reifman J. Metabolic host responses to malarial infection during the intraerythrocytic developmental cycle. BMC Syst Biol. 2016;10(1):1-18.
- Omoregie ES, Pal A. Antiplasmodial, antioxidant and immunomodulatory activities of ethanol extract of *Vernonia amygdalina* del. leaf in swiss mice. Avicenna J Phytomed. 2016;6(2):236-47.
- 25. Tripathy S, Roy S. Redox sensing and signaling by malaria parasite in vertebrate host. J Basic Microbiol. 2015;55(9):1053-63.
- Pan WH, Xu XY, Shi N, Tsang SW, Zhang HJ. Antimalarial activity of plant metabolites. Int J Mol Sci. 2018;19(1382):1-40.
- 27. Wang H, Kumar A, Lamont RJ, Scott D. GSK3-beta and the control of infectious bacterial diseases. Trends Microbiol. 2014;22(4):208-17.
- Wong SK, Jann MLS, Sudi S, Hassan WRM, Chin LP, Embi N, *et al.* Antimalarial and antiinflammatory effects of Gynura procumbens are mediated by kaempferol via inhibition of glycogen synthase kinase-3β (GSK3β). Sains Malaysiana. 2015;44(10):1489-500.
- Droucheau E, Primot A, Thomas V, Mattei D, Knockaert M, Richardson C, *et al.* Plasmodium falciparum glycogen synthase kinase-3: molecular model, expression, intracellular localisation and selective inhibitors. Biochim Biophys Acta. 2004;1697(1-2):181-96.
- Somsak V, Damkaew A, Onrak P. Antimalarial activity of kaempferol and its combination with chloroquine in *Plasmodium berghei* infection in mice. J Pathog. 2018:1-7.
- Barliana MI, Suradji E, Abdulah R. Antiplasmodial properties of kaempferol-3-o-rhamnoside isolated from the leaves of *Schima wallichii* against chloroquine-resistant *Plasmodium falciparum*. Biomed Rep. 2014;2(4):579-83.
- Thiengsusuk A, Muhamad P, Chaijaroenkul W, Na-Bangchang K. Antimalarial activity of piperine. J Trop Med. 2018:1-7.
- Sabina EP, Nasreen A, Vedi M, Rasool M. Analgesic, antipyretic and ulcerogenic effects of piperine: an active ingredient of pepper. J Pharm Sci Res. 2013;5(10):203-06.
- 34. Mittal R, Gupta R. In vitro Antioxidant activity of piperine. Methods Find Exp Clin Pharmacol. 2000;22(5):271-74.
- Tharmalingam N, Kim SH, Park M, Woo HJ, Kim HW, Yang JY, *et al.* Inhibitory effect of piperine on *Helicobacter pylori* growth and adhesion to gastric adenocarcinoma cells. Infect Agents Cancer. 2014;9(43):1-10.
- Lee SA, Hong SS, Han XH, Hwang JS, Oh GJ, Lee KS, *et al.* Piperine from the fruits of *Piper longum* with inhibitory effect on monoamine oxidase and antidepressant-like activity. Chem Pharm Bull. 2005;53(7):832-35.
- Nirwane AM, Bapat A. Effect of methanolic extract of *Piper nigrum* fruits in ethanol-CCl4 induced hepatotoxicity in wistar rats. Der Pharma Lettre. 2012;4(3):795-802.

- Kim SH, Lee YC. Piperine inhibits eosinophil infiltration and airway hyperresponsiveness by suppressing T cell activity and Th2 cytokine production in the ovalbumin-induced asthma model. J Pharm Pharmacol. 2009;61:353-59.
- Bang JS, Choi HM, Sur BJ, Lim SJ, Kim JY, Yang HI, *et al.* Antiinflammatory and antiarthritic effects of piperine in human interleukin 1β-stimulated fibroblast-like synoviocytes and in rat arthritis models. Arthritis Res Ther. 2009;11(2):1-9.
- Bukhari IA, Alhumayyd MS, Mahesar AL, Gilani AH. The analgesic and anticonvulsant effects of piperine in mice. J Physiol Pharmacol. 2013;64(6):789-94.
- 41. Antony HA, Parija S. Antimalarial drug resistance: an overview. Trop Parasitol. 2016;6(1):30-41.
- Rijpma SR, Van Der Velden M, Bilosetal A. MRP1 mediates folate transport and antifolate sensitivity in *Plasmodium falciparum*. FEBS Lett. 2016;590(4):482-92.
- Huang B, Wang Q, Dengetal C. Prevalence of crt and mdr-1 mutations in *Plasmodium falciparum* isolates from grande comore island after withdrawal of chloroquine. Malar J. 2016;15(1):1-9.
- 44. Martin RE, Kirk, K. The malaria parasite's chloroquine resistance transporter is a member of the drug/metabolite transporter superfamily. Mol Bio Evol. 2004;21(10):1938-49.
- Jayakumar T, Hsieh CY, Lee JJ, Sheu J. Experimental and clinical pharmacology of *Andrographis paniculata* and its major bioactive phytoconstituent andrographolide. Evid based Compl Alternat Med. 2013:1-16.
- Jarukamjorn K, Nemoto N. Pharmacological aspects of Andrographis paniculata on health and its major diterpenoid constituent andrographolide. J Health Sci. 2008;54(4):370-81.
- Mishra K, Dash AP, Dey NA. Andrographolide: a novel antimalarial diterpene lactone compound from *Andrographis paniculata* and its interaction with curcumin and artesunate. J Trop Med. 2011:1-6.
- Tripathi AK, Sha W, Shulaev V, Stins MF, Sullivan Jr D. Plasmodium falciparum-infected erythrocytes induce NFκB regulated inflammatory pathways in human cerebral endothelium. Blood. 2009;114(19):4243-52.
- Bao Z, Guan S, Cheng C, Wu S, Wong SH, Kemeny DM, *et al.* A novel antiinflammatory role for andrographolide in asthma via inhibition of the nuclear factor-κb pathway. Am J Respir Crit Care Med. 2009;179(8):657-65.
- Risdawati. The mechanism of action of andrographolide a compound of sambiloto (*Andrographis paniculata* Nees.) as an antimalarial drug study on oxidative stress of *Plasmodium berghei* ANKA. Disertasi, Program Doktoral. Depok: Universitas Indonesia; 2014.
- Kusumawardhani D, Widyawaruyanti A, Kusumawati I. Efek antimalaria ekstrak sambiloto terstandar (parameter kadar andrografolida) pada mencit terinfeksi *Plasmodium berghei*. Airlangga J Pharm. 2005;5(1):25-9.
- Sachdeva M. Analysis of in vitro antimalarial activity of andrographolide and 5-hydroxy-7,8-dimethoxyflavone isolated from *Andrographis paniculata* against *Plasmodium berghei* parasite. Pharm Sci Monitor. 2011;2(4):104-16.
- Mahabusarakam W, Kuaha K, Wilairat P, Taylor W. Prenylated xanthones as potential antiplasmodial substances. Planta Med. 2006;72(10):912-16.
- Ibrahim MY, Hashim NM, Mariod AA, Mohan S, Abdulla MA, Abdelwahab SI, *et al.* α-mangostin from *Garcinia mangostana* Linn: an updated review of its pharmacological properties. Arabian J Chem. 2016;9:317-29.
- Ignatushchenko MV, Winter RW, Baechinger HP, Hinrichs DJ, Riscoe M. Xanthones as antimalarial agents: studies of a possible mode of action. FEBS Lett. 1997;409:67-73.

- Riscoe M, Kelly JX, Winter R. Xanthones as Antimalarial agents: discovery, mode of action and optimization. Curr Med Chem. 2005;12:2539-49.
- Kelly JX, Winter R, Peyton DH, Hinrichs DJ, Riscoe M. Optimization of xanthones for antimalarial activity: the 3,6-bisdiethylaminoalkoxyxanthone series. Antimicrob Agents Chemother. 2002;46:144-50.
- Kuncoro H, Widyawaruyanti A, Ersam T. α-mangostin effect on inhibition development stadium and globin accumulation against *Plasmodium falciparum*. Pharmacog J. 2018;10(4):783-88.
- Upegui Y, Robledo SM, Romero JFG, Quiñones W, Archbold R, Torres F, *et al.* In vivo antimalarial activity of α-mangostin and the new xanthone δ-mangostin. Phytother Res. 2015;29(8):1195-201.
- Sen G, Sarkar N, Nath M, Maity S. Bioactive components of tea. Arch Food Nutr Sci. 2020;4:001-09.
- 61. Bae J, Kim N, Shin Y, Kim SY, Kim Y. Activity of catechins and their applications. Biomed Dermatol. 2020;4(8):1-10.
- Budiman I, Tjokropranoto R, Widowati W, Rahardja F, Maesaroh M, Fauziah N. Antioxidant and anti-malarial properties of catechins. Br J Med Med Res. 2015;5(7):895-902.
- Abdulah R, Suradji EW, Subarnas A, Supratman U, Sugijanto M, Diantini A, et al. Catechin isolated from Garcinia celebica leaves inhibit Plasmodium falciparum growth through the induction of oxidative stress. Pharmacogn Mag. 2017;13(2):301-5.
- Goyal M, Alam A, Bandyopadhyay U. Redox regulation in malaria: current concepts and pharmacotherapeutic implications. Curr Med Chem. 2012;19:1475-503.
- Mohring F, Pretzel J, Jortzik E, Becker K. The redox systems of *Plasmodium falciparum* and *Plasmodium vivax*: comparison, in silico analysis and inhibitor studies. Curr Med Chem. 2014;21:1728-56.
- Lin Y, Shi R, Wang X, Shen H. Luteolin, a flavonoid with potentials for cancer prevention and therapy. Curr Cancer Drug Targets. 2008;8(7):634-46.
- Dong H, Yang X, He J, Cai S, Xiao K, Zhu L. Enhanced antioxidant activity, antibacterial activity and hypoglycemic effect of luteolin by complexation with manganese (II) and its inhibition kinetics on xanthine oxidase. RSC Adv. 2017;7:53385-95.
- Lehane AM, Saliba K. Common dietary flavonoids inhibit the growth of the intraerythrocytic malaria parasite. BMC Res Notes. 2008;1(26):1-5.
- Deniz T, Gabriela L, Reto B, Peter R, Leonardo S, Remo P. Inhibition of *Plasmodium falciparum* fatty acid biosynthesis: evaluation of FabG, FabZ, and FabI as drug targets for flavonoids. J Med Chem. 2006;49:3345-53.
- Chen CY, Peng WH, Tsai KD, Hsu S. Luteolin suppresses inflammation-associated gene expression by blocking NF-kappaB and AP-1 activation pathway in mouse alveolar macrophages. Life Sci. 2007;81:1602-14.
- Xagorari A, Roussos C, Papapetropoulos A. Inhibition of LPSstimulated pathways in macrophages by the flavonoid luteolin. Br J Pharmacol. 2002;136:1058-64.
- 72. Karin M. Mitogen activated protein kinases as targets for development of novel anti-inflammatory drugs. Ann Rheum Dis. 2004;63(2):62-4.
- Tan Y, Yu R, Pezzuto JM. Betulinic acid-induced programmed cell death in human melanoma cells involves mitogen-activated protein kinase activation. Clin Cancer Res. 2003;9(7):2866-75.
- 74. Ríos J, Máñez S. New pharmacological opportunities for betulinic acid. Planta Med. 2018;84(01):8-19.
- Steele JCP, Warhurst DC, Kirby GC, Simmonds MS. In vitro and in vivo evaluation of betulinic acid as an antimalarial. Phytother Res. 1999;13:115-19.

- Silva GNS, Maria NRG, Schuck DC, Cruz LN, de Moraes MS, Nakabashi M, *et al.* Two series of new semisynthetic triterpene derivatives: differences in anti-malarial activity, cytotoxicity and mechanism of action. Malar J. 2013;12(89):1-7.
- Diedrich D, Wildner AC, Silveira TF, Silva GNS, Santos Fd, do Canto VP, *et al.* SERCA plays a crucial role in the toxicity of a betulinic acid derivative with potential antimalarial activity. Chem-Biol Interact. 2018;287:70-7.
- Eckstein-Ludwig U, Webb RJ, van Goethem IDA, East JM, Lee AG, Kimura M, et al. Artemisinins target the SERCA of *Plasmodium falciparum*. Nature. 2003;424:957-61.
- Jung M, Kim H, Nam KY, No K. Three-dimensional structure of *Plasmodium falciparum* Ca<sup>2+</sup>ATPase (PfATP6) and docking of artemisinin derivatives to PfATP6. Bioorg Med Chem Lett. 2005;15(12):2994-97.



# **GRAPHICAL ABSTRACT**

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