Juglans regia L.: Source of Bioactive Compounds with Potential Anticancer Activity

Carla Y. Requejo-Rodríguez¹, Elmer M. Roncal-Alayo¹, Carmen R. Silva-Correa^{2,*}, Víctor E. Villarreal-La Torre², William A. Sagástegui-Guarniz², César D. Gamarra-Sánchez², Walter E. Janampa-Castillo³, José E. Alvarez-Trujillo³, Glenda J. Vela-Urbina³, Abhel A. Calderón-Peña⁴, Cinthya L. Aspajo-Villalaz⁴, María E. Cotrina-León⁵, Julio A. Castañeda-Carranza⁵, Deivy Y. Dionicio-Rosado⁶

Carla Y. Requejo-Rodríguez¹, Elmer M. Roncal-Alayo¹, Carmen R. Silva-Correa^{2,*}, Víctor E. Villarreal-La Torre², William A. Sagástegui-Guarniz², César D. Gamarra-Sánchez², Walter E. Janampa-Castillo³, José E. Alvarez-Trujillo³, Glenda J. Vela-Urbina³, Abhel A. Calderón-Peña⁴, Cinthya L. Aspajo-Villalaz⁴, María E. Cotrina-León⁵, Julio A. Castañeda-Carranza⁵, Deivy Y. Dionicio-Rosado⁶

¹Escuela de Posgrado, Universidad Nacional de Trujillo, Perú

²Facultad de Farmacia y Bioquímica, Universidad Nacional de Trujillo, Perú

³Facultad de Educación y Ciencias de la Comunicación, Universidad Nacional de Trujillo, Perú

⁴Facultad de Ciencias Biológicas, Universidad Nacional de Trujillo, Perú.

⁵Facultad de Ciencias Físicas y Matemáticas, Universidad Nacional de Trujillo, Perú.⁶Facultad de Ciencias Sociales y Humanidades, Universidad Nacional Ciro Alegría, Perú

Correspondence

Carmen R. Silva-Correa

Facultad de Farmacia y Bioquímica, Universidad Nacional de Trujillo, PERÚ.

E-mail: csilva@unitru.edu.pe

History

- Submission Date: 01-07-2024;
- Review completed: 13-08-2024;
- Accepted Date: 19-08-2024.

DOI : 10.5530/pj.2024.16.161

Article Available online

http://www.phcogj.com/v16/i5

Copyright

 $\hfill \odot$ 2024 Phcogj.Com. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Phcog j.co



Background: Juglans regia L., commonly known as "walnut", belongs to the Juglandaceae family, with antioxidant, anti-inflammatory, and hypoglycemic medicinal properties. **Objective:** Describe the anticancer potential of the bioactive compounds present in *Juglans regia* L. **Method:** Recent scientific studies were reviewed on the effects of bioactive compounds from *Juglans regia* L. on inhibiting tumor growth and cancer development in several experimental models. To do this, a scientific literature search was carried out, using databases such as PubMed, Scopus, and Science Direct. **Results:** Regarding the selected articles, it was found that some bioactive compounds from *Juglans regia* L. exhibit mechanisms of anticancer action, among which the following stand out: induction of apoptosis, suppression of angiogenesis, and modulation of cell signaling pathways related to cell proliferation and survival. **Conclusion:** It is concluded that *Juglans regia* L. contains active metabolites with potential anticancer effects. **Keywords:** *Juglans regia* L., Cancer, Antitumor, Apoptosis, Angiogenesis.

INTRODUCTION

Cancer is one of the most widespread diseases and one of the main causes of death that is continuously increasing, due to the lack of selectivity and resistance induced by chemotherapy drugs¹. World cancer statistics record 18.1 million new cases of cancer with 9.6 million deaths due to this disease and that the main cause of death is lung cancer followed by breast, colorectal, stomach, and liver².

Plant secondary metabolites are important sources of molecules with potential for chemotherapy. These bioactive compounds have been shown to increase anticancer activities and reduce serious side effects of antitumor drugs³.

Juglans regia L. (*J. regia*) commonly known as "walnut", belongs to the Juglandaceae family, frequently cultivated in China, the United States, Eastern Europe, and Iran⁴, this plant species has been used ancestrally to treat various conditions in humans⁵.

The phytoconstituents found in this plant vary depending on the geographical location, climatic conditions, and nature of the soil. Thus, in the leaves of *J. regia*, phytochemicals with pharmacological properties are found, among the most common of which are phenolic acids, tannins, essential fatty acids, ascorbic acid, flavonoids such as quercetin, caffeic acid, paracomaric acid and juglone (5-hydroxy-1,4-naphthoquinone) as the most active compound^{6,7}. These components not only contribute to its antioxidant activity⁸ but have also been shown to have lipid-lowering, antihypertensive⁹, antibacterial^{10,11}, antifungal¹², anti-inflammatory¹³, neuroprotective¹⁴, antithrombotic¹⁵activities, healing activity in incision and excision wounds¹⁶, and hypoglycemic activity¹⁷. Research supports its medicinal properties, highlighting its anti-cancer potential⁶.

The purpose of this review is to provide an overview of the anticancer potential of the bioactive compounds of *J. regia* L. based on the scientific literature found to date and to promote further studies that contribute to finding compounds that contribute to the treatment of cancer.

METHODOLOGY

An exhaustive review of the scientific literature was carried out, the review studies have been compiled from scientific articles published in databases such as Science Direct, Scopus, and PubMed. Original scientific documents were included that covered descriptive and experimental studies related to the potential anticancer activity of the bioactive compounds of *Juglans regia* L. Original articles in preclinical and clinical phases were considered. The keywords in the scientific literature search were obtained from DeCS in Spanish and MeSH in English, which allowed a quick selection of scientific articles related to the thematic field.

Bioactive Compounds of Juglans regia L.

Juglans regia L. has found diverse applications in both the cosmetic and pharmaceutical industries due to its bioactive compounds present in its fruits, peels, seeds, bark, flowers, and leaves¹⁸. *J. regia* L. serves as a significant dietary source of protein, fiber, essential fatty acids, vitamins, and minerals. Additionally, it provides a diverse array of flavonoids, phenolic acids, and related polyphenols¹⁹.

Cite this article: Requejo-Rodríguez CY, Roncal-Alayo EM, Silva-Correa CR, Villarreal-La Torre VE, Sagástegui-Guarniz WA, Gamarra-Sánchez CD, Janampa-Castillo WE, et al. *Juglans regia* L.: Source of Bioactive Compounds with Potential Anticancer Activity. Pharmacogn J. 2024;16(5): 998-1003. Walnuts serve as a significant fat source (approximately 74 g/100 g) with a high caloric value of 720 kcal/100 g. Beyond fat content, walnuts also contain valuable polyunsaturated fatty acids and plant proteins rich in essential amino acids and minerals. Proteins, carbohydrates, crude fiber, and minerals constitute approximately 24%, 14%, 2%, and 1.7% of the nut's weight, respectively. Additionally, walnuts harbor bioactive components such as polyphenols and tocopherols, with hydroxybenzoic acids predominantly found in the polyphenolic profile, particularly gallic acid²⁰. In the tocopherol spectrum, four forms of tocopherols are identified: α , β , γ , and δ -tocotrienol²¹, with α -tocopherol being the one with the highest content and having antioxidant activity, principally in preventing the lipid oxidation process²².

In *J. regia* nut oil, various bioactive compounds have been identified, including tocopherols, sterols, triterpenic and aliphatic alcohols, carotenoids, and volatile compounds. Among the phytosterols, β -sitosterol is the most abundant, followed by campesterol and Δ -5-avenasterol. The major triterpenic alcohol present is cycloartenol, while hexacosanol serves as the primary aliphatic alcohol. Additionally, the volatile compounds detected include pentanal, hexanal, nonanal, 2-decenal, and hexanol²³.

Naphthoquinones, including juglone and hydrojuglone glycosides, have been identified in various tissues of *J. regia*²⁴. Juglona pueden combatir el estrés oxidativo y proteger contra diversas enfermedades y el envejecimiento²⁵. Juglone exhibits antioxidant properties, protecting against oxidative stress and age-related diseases. However, as a quinone, juglone can also act as a pro-oxidant, inhibiting cancer cell growth²⁶. Additionally, research has investigated polymeric micelles as a delivery system for juglone in pancreatic cancer treatment²⁷.

J. regia leaves reported several bioactive compounds such as ascorbic acid, juglone²⁸, phenolic compounds, 3-caffeoylquinic, 3-p-coumaroylquinic, and 4-p-coumaroylquinic acids²⁹. Additionally, flavonoids like epicatechin, syringetin-O-hexoside, myricetin-3-O-glucoside, myricetin-3-O-pentoside, esculetin, taxifolin-pentoside, quercetin-3-O-glucuronide, kaempferol-O-pentoside and kaempferol-O-rhamnoside³⁰. The acetonic extract from *J. regia* flowers reveals gallic acid, α -tocopherol, and substantial quantities of quercetin hyperoside, quercitrin, and isoquercitrin³¹.

Overall, the bioactive compounds identified in *J. regia* have been recognized as potential anticancer agents in the literature. These compounds include ellagic acid³², quercetin, ascorbic acid³³, tocopherol³⁴, rutin³⁵, carotenoids³⁶, and phytosterols³⁷.

Anti-Cancer Evaluation of Juglans regia L.

Extracts from various parts of *Juglans regia* demonstrate anticancer and antitumor properties by inhibiting proliferation and inducing apoptosis in cancer cells. These plant extracts were evaluated both *in vitro* using cancer cell lines and *in vivo* through animal models. The primary studies assessing the anticancer effects of *Juglans regia* are summarized in Table 1.

Anti-Cancer Mechanisms

The described mechanisms offer a comprehensive understanding of how compounds from *Juglans regia* L. could influence various stages and processes in tumor development, establishing a robust foundation for considering their therapeutic potential in cancer treatment.

Juglone

Juglone inhibits the growth of malignant cells, including HCT-15 cells derived from human colon carcinoma, primarily by blocking the S phase of the cell cycle. Additionally, it induces apoptosis in human leukemia cells (HL-60), human gastric cancer cells (SGC-7901), and SKOV3 ovarian cancer cells through mitochondria-dependent

apoptosis pathways and an elevated Bax/Bcl-2 ratio. Furthermore, Juglone significantly inhibits proliferation and induces apoptosis in human bladder carcinoma cell lines (TCC-SUB and RT-4), while in the human breast cancer cell line (MCF-7), it leads to elevated levels of reactive oxygen species (ROS), reduced Bcl-2 expression, increased Bax expression, decreased mitochondrial membrane potential, elevated intracellular Ca²⁺ concentration, rupture of the outer mitochondrial membrane, cytochrome c release, and caspase-3 activation²⁵. Research also suggests that juglone significantly inhibits cell proliferation and activates apoptosis in colon cancer. Additionally, juglone negatively regulates the Wnt/ β -catenin pathway^{50,51}.

Juglone was evaluated on intestinal carcinogenesis in rats by dietary exposure during the initiation phase. Data suggest that juglone could be a promising chemopreventive agent for human intestinal neoplasia. Juglone inhibits tumor progression in mice, triggering oxidative stress leading to apoptosis and cell cycle arrest, suppression of hypoxia-inducible factor 1alpha, and disengagement of glycolytic metabolism.²⁵.

Ellagic Acid

Ellagic acid may exert an antitumor effect through diverse molecular mechanisms, including inhibition of proliferation, induction of apoptosis, suppression of metastasis and invasion, autophagy induction, and modulation of tumor metabolic reprogramming³². Studies demonstrate that ellagic acid reduces cell viability and suppresses tumors in breast cancer. Specifically, it inhibits cell growth and migration by arresting the cell cycle and suppressing metastasis⁵², activates pro-apoptotic pathways, and positively regulates p53³². Additionally, ellagic acid stimulates apoptosis via the TGF- β /Smad3 pathway and regulates CDK6 during the G0/G1 phase⁵³. Furthermore, it reduces cell motility and invasion by binding to and degrading ACTN4 through the ubiquitin-proteasome pathway. Moreover, ellagic acid decreases angiogenesis by reducing VEGFR-2 activity⁵⁴ and inhibits proliferation, migration, and invasion by targeting Wnt/ β -catenin and PI3K/Akt signaling pathways⁵⁵.

Quercetin

In vitro investigations have revealed distinct effects of quercetin on normal prostate cells compared to prostate cancer cells, emphasizing its specific cytotoxic impact on cancer cells. The anticancer mechanism involves Bax dissociation from Bcl-xL⁵³ and modulation of PI3K/Akt/ mTOR, Wnt/ β -catenin, and MAPK/ERK1/2 pathways. Quercetin promotes loss of cell viability, apoptosis, and autophagy in cancer by reducing the stabilization of β -catenin and HIF-1. Additionally, it activates caspase-3 and inhibits phosphorylation of Akt, mTOR, and ERK. Furthermore, quercetin prevents metastasis by reducing VEGF and MMP secretion⁵⁶.

Similarly, quercetin demonstrates inhibitory effects on cell proliferation and angiogenesis in animal models of prostate cancer, along with a reduction in tumor size⁵⁷. However, the clinical application of this agent faces several challenges related to its bioavailability, absorption, metabolism, stability, and rapid elimination from circulation. Addressing these issues is essential to enhance the clinical utility of quercetin⁵³.

Tocopherol

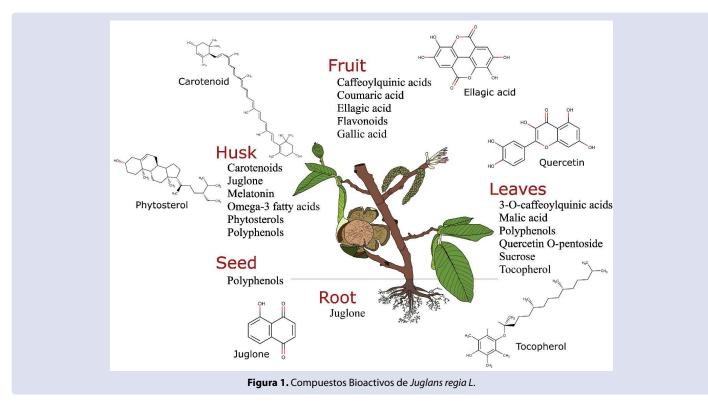
In animal models of liver cancer, tocopherol (vitamin E) has demonstrated the ability to suppress angiogenesis, induce apoptosis, and reduce metastasis formation⁵⁸.

Carotenoids

Carotenoids have demonstrated protective properties against oxidative stress and reduced cell proliferation in breast cancer studies⁵⁹. Although *in vitro* and *in vivo* research has yielded promising results regarding

Table 1. Studies evaluating the anticancer activity of Juglans regia L.

Plant part	Extract	Doses	Bioactive com- pounds	Experimental model	Results	Ref
fruit	ethanol-water in a ratio 80:20	50 and 100 mg/kg body weight	gallic acid, caffeoylquinic acids, coumaric acid and flavonoids	prostate cancer in rats	The administration of Juglans regia extract was associated with a reversal of testosterone induced changes in oxidative stress or inflammatory markers	38
husk	ethanolic extract	1 and 10 µM juglone	juglone	HL-60, human leukemia cell line	Apoptosis of HL-60 was detected at 10 μM juglone	6
husk	Extractions were carried out in a Soxhlet apparatus with three different solvents: methanol, chloroform, and n-hexane.	100 mg/mL	omega-3 fatty acids, phytosterols, polyphenols, carotenoids, and melatonin	PC-3 Human Prostate Cancer Cell	The methanol (IC50 66.72 μ g/mL), n-hexane (IC50 27.29 μ g/mL), and chloroform (IC50 91.14 μ g/mL). Green husk extracts suppressed proliferation and induced apoptosis in a dose- and time- dependent manner.	39
fruit	ethanolic extract	8.75 to 140 μg/ml	ellagic acid derivatives and flavanols	human A172 glioblastoma cell line	Extract could decrease cancer cell proliferation and migration	40
fruit	diet	The equivalent of 80 g (approximately 3 ounces) of walnuts in humans	not detected	the transgenic adenocarcinoma of the mouse prostate model	Walnuts as part of a diet reduce tumour growth and size.	41
leaves	Hexane Extract	5, 50 and 100 μg/mL	not detected	PC-3 Human Prostate Cancer Cell	Inhibits growth of human prostate cancer cells by inducing apoptosis with concomitant alterations in cell cycle phase distribution	42
fruit	Methanolic extract and fractions	1, 10, 100, and 500 μg/mL	phenolic compounds	Human cancer cell lines, such as MCF-7 (estrogen receptor positive breast adenocarcinoma), KB (oral and mouth), HepG-2 (liver), Caco2 (colon), and WRL-68 (liver)	Chloroform and ethyl acetate fractions exhibited a high level of antiproliferation against HepG-2, liver cancer cell line (IC50 = 9 and 15 µg/mL, respectively).	43
root bark	Extractions were carried out in a Soxhlet apparatus with three different solvents; methanol, choloroform and n-hexane.	500μg/mL	not detected	MDA-MB-231 breast cancer cells	Suppressed proliferation and induced apoptosis in a dose and time dependent manner	44
root	chloroform extract	100 μΜ	juglone	Prostate colon (Colo-205 and HCT- 116), breast (T47D), prostate (PC-3 and DU-145), skin (A-431) and lung (NCI-H322 and A549).	Exhibited satisfactory cytotoxic activity against a panel of eight different human cancer cell lines	45
seed, green husk and leaf.	mixed with methanol or petroleum ether	31.25, 62.5, 125, 250 and 500 lg extract/mL.	polyphenols	Human renal cancer cell lines A-498 and 769-P and the colon cancer cell line Caco-2.	Showed concentration dependent growth inhibition toward human kidney and colon cancer cells.	46
leaves	hexane, chloroform, ethyl acetate and methanol fractions	0.25 - 1.5 mg/mL	polyphenols, flavonoids and condensed tannins	Human oral cancer, breast adenocarcinoma and colon adenocarcinoma cell lines.	Walnut chloroform fraction may contain effective compounds which can be used as a chemotherapeutic agent.	47
fruit	methanol, ethanol, and hexane solvents	2.0 μg/ml	not detected	MDA-MB-231 cell line	Showed evident apoptotic activity even if it was not as potent as doxorubicin.	48
Leaves	decoction	different concentrations not reported	Malic acid, sucrose, tocopherol, 3-O-caffeoylquinic acids and quercetin O-pentoside	MCF-7 (breast adenocarcinoma), NCI-H460 (non-small cell lung cancer), HCT-15 (colon carcinoma), HeLa (cervical carcinoma) and HepG2 (hepatocellular carcinoma)	The methanol extract was most potent against cervical carcinoma cell line (HeLa, GI50= 294.87 μg/mL)	49



the impact of carotenoids on breast cancer, clinical trials have not yet provided definitive conclusions 60 .

Phytosterols

Phytosterols can suppress angiogenesis, metastasis, infiltration, and proliferation of cancer cells by regulating the tumor microenvironment through molecular signaling pathways associated with growth factors, chemokines, pro-inflammatory mediators, and pro/anti-apoptotic genes. These compounds have been demonstrated to arrest G_0/G_1 and G_2 cell cycles, promote cell death by upregulating caspases and pro-apoptotic enzymes while downregulating anti-apoptotic enzymes. Additionally, phytosterols enhance immune system function by improving T-cell proliferation and cytotoxicity. Conversely, they also have the potential to inhibit angiogenesis, leading to reduced levels of vascular endothelial growth factor (VEGF) and transforming growth factor-beta (TGF- β)³⁷.

Moreover, they have demonstrated inhibition of cell proliferation and invasion, as well as a reduction in inflammation and oxidative stress by targeting reactive oxygen species (ROS), inhibiting nuclear factor kappa B (NF- κ B), and decreasing cell invasion in prostate cancer⁶¹.

FUTURE PERSPECTIVES

This review highlights the potential of bioactive compounds found in *Juglans regia* L. as promising agents against various types of cancer. The included studies demonstrate the effectiveness of these compounds, including juglone, ellagic acid, quercetin, ascorbic acid, tocopherol, rutin, carotenoids, and phytosterols. These bioactive molecules exhibit inhibition of cell proliferation, induction of apoptosis, suppression of angiogenesis, and modulation of cancer-associated signaling pathways in different cellular models and cancer types. However, due to the scarcity of human studies, conclusive results regarding the safety and efficacy of these compounds in cancer treatment remain a challenge for future research.

CONCLUSIONS

The bioactive compounds found in *Juglans regia* L. hold promise as potential anticancer agents, given their ability to inhibit cell proliferation and induce apoptosis. Additionally, several of these compounds demonstrate efficacy in suppressing angiogenesis and modulating cancer-associated signaling pathways. These findings underscore the importance of further exploring the specific molecular mechanisms underlying the therapeutic potential of bioactive compounds derived from *Juglans regia* L. for cancer treatment. Future research should focus on evaluating optimal doses and potential side effects in various cancer types.

CONFLICTS OF INTEREST

The authors declare no conflict of interest.

REFERENCES

- 1. Jaber SA. The antioxidant and anticancer activity of *Quercus* coccifera plant leaves extracts. *Saudi J Biol Sci.* 2024;31(5). doi:10.1016/j.sjbs.2024.103979
- Chandra S, Gahlot M, Choudhary AN, Palai S, Silva R, Lucena JE, *et al.* Scientific evidences of anticancer potential of medicinal plants. *Food Chemistry Advances.* 2023;2. doi:10.1016/j. focha.2023.100239
- Goyal Y, Koul A, Ranawat P. Ellagic acid modulates cisplatin toxicity in DMH induced colorectal cancer: Studies on membrane alterations. *Biochem Biophys Rep.* 2022;31. doi:10.1016/j.bbrep.2022.101319
- Isaac R, Siddiqui S, Aldosari OF, Kashif Uddin M. Magnetic biochar derived from *Juglans regia* for the adsorption of Cu2+ and Ni2+: Characterization, modelling, optimization, and cost analysis. *Journal of Saudi Chemical Society*. 2023;27(6). doi:10.1016/j. jscs.2023.101749

- Dolatabadi S, Moghadam HN, Mahdavi-Ourtakand M. Evaluating the anti-biofilm and antibacterial effects of *Juglans regia* L. extracts against clinical isolates of *Pseudomonas aeruginosa. Microb Pathog.* 2018;118:285-289. doi:10.1016/j.micpath.2018.03.055
- Soto-Maldonado C, Vergara-Castro M, Jara-Quezada J, Caballero-Valdés E, Müller-Pavez A, Zúñiga-Hansen ME, et al. Polyphenolic extracts of walnut (*Juglans regia*) green husk containing juglone inhibit the growth of HL-60 cells and induce apoptosis. *Electronic Journal of Biotechnology*. 2019;39:1-7. doi:10.1016/j. ejbt.2019.02.001
- Gupta A, Behl T, Panichayupakaranan P. A review of phytochemistry and pharmacology profile of *Juglans regia*. Obes Med. 2019;16. doi:10.1016/j.obmed.2019.100142
- Fernández-Agulló A, Castro-Iglesias A, Freire MS, González-Álvarez J. Optimization of the extraction of bioactive compounds from walnut (*Juglans major* 209 x *juglans regia*) leaves: Antioxidant capacity and phenolic profile. *Antioxidants*. 2020;9(1). doi:10.3390/ antiox9010018
- Singh Jaiswal B, Tailang M. Juglans regia: A review of its traditional uses phytochemistry and pharmacology. Indo American Journal of Pharmaceutical Research. 2017;7(9):390-398. www.iajpr.com
- Sharma P, Ravikumar G, Kalaiselvi M, Gomathi D, Uma C. *In vitro* antibacterial and free radical scavenging activity of green hull of *Juglans regia. J Pharm Anal.* 2013;3(4):298-302. doi:10.1016/j. jpha.2013.01.006
- Acquaviva R, D'Angeli F, Malfa GA, Ronsisvalle S, Garozzo A, Stivala A, et al. Antibacterial and anti-biofilm activities of walnut pellicle extract (*Juglans regia* L.) against coagulase-negative staphylococci. Nat Prod Res. 2021;35(12):2076-2081. doi:10.1080/14786419.2019.1650352
- Ara T, Shafi S, Ghazwani M, Mir JI, Shah AH, Qadri RA, et al. In Vitro Potent Anticancer, Antifungal, and Antioxidant Efficacy of Walnut (*Juglans regia* L.) Genotypes. Agronomy. 2023;13(5). doi:10.3390/ agronomy13051232
- Mobashar A, Shabbir A, Shahzad M, Gobe G. Preclinical Rodent Models of Arthritis and Acute Inflammation Indicate Immunomodulatory and Anti-Inflammatory Properties of Juglans regia Extracts. *Evidence-based Complementary and Alternative Medicine*. 2022;2022. doi:10.1155/2022/1695701
- Sharma P, Verma PK, Sood S, Pankaj NK, Agarwal S, Raina R. Neuroprotective potential of hydroethanolic hull extract of *Juglans regia* L. on isoprenaline induced oxidative damage in brain of Wistar rats. *Toxicol Rep.* 2021;8:223-229. doi:10.1016/j. toxrep.2021.01.006
- Amirou A, Razzok EM, Legssyer A, Ziyyat A, Aziz M, Bnouham M, *et al.* Effects of Walnut Bark Extract on the Human Platelet Aggregation, Adhesion, and Plasmatic Coagulation *in Vitro. Adv Pharmacol Pharm Sci.* 2023;2023. doi:10.1155/2023/5644803
- Al-Nadaf AH, Awadallah A, Thiab S. Superior rat wound-healing activity of green synthesized silver nanoparticles from acetonitrile extract of *Juglans regia* L: Pellicle and leaves. *Heliyon*. 2024;10(2). doi:10.1016/j.heliyon.2024.e24473
- Nurcahyanti ADR, Jap A, Lady J, Prismawan D, Sharopov F, Daoud R, *et al.* Function of selected natural antidiabetic compounds with potential against cancer via modulation of the PI3K/AKT/ mTOR cascade. *Biomedicine and Pharmacotherapy.* 2021;144. doi:10.1016/j.biopha.2021.112138
- Zakavi F, Golpasand Hagh L, Daraeighadikolaei A, Farajzadeh Sheikh A, Daraeighadikolaei A, Leilavi Shooshtari Z. Antibacterial effect of *Juglans regia* bark against oral pathologic bacteria. *Int J Dent.* 2013;2013. doi:10.1155/2013/854765
- Yan M, Chen M, Zhou F, Cai D, Bai H, Wang P, *et al.* Separation and analysis of flavonoid chemical constituents in flowers of *Juglans regia* L. by ultra-high-performance liquid chromatography-hybrid quadrupole time-of-flight mass spectrometry. *J Pharm Biomed Anal.* 2019;164:734-741. doi:10.1016/j.jpba.2018.11.029

- Pycia K, Kapusta I, Jaworska G, Jankowska A. Antioxidant properties, profile of polyphenolic compounds and tocopherol content in various walnut (*Juglans regia* L.) varieties. *European Food Research and Technology*. 2019;245(3):607-616. doi:10.1007/ s00217-018-3184-3
- Amaral JS, Alves MR, Seabra RM, Oliveira BPP. Vitamin E composition of walnuts (*Juglans regia* L.): A 3-year comparative study of different cultivars. *J Agric Food Chem.* 2005;53(13):5467-5472. doi:10.1021/jf050342u
- Ara I, Shinwari MMA, Rashed SA, Bakir MA. Evaluation of Antimicrobial Properties of Two Different Extracts of *Juglans regia* Tree Bark and Search for Their Compounds Using Gas Chromatohraphy-Mass Spectrum. *Int J Biol.* 2013;5(2). doi:10.5539/ ijb.v5n2p92
- Abdallah IB, Tilii N, Martinez-Force E, Rubio AG, Perez-Camino MC, Albouchi A, *et al.* Content of carotenoids, tocopherols, sterols, triterpenic and aliphatic alcohols, and volatile compounds in six walnuts (*Juglans regia* L.) varieties. *Food Chem.* 2015;173:972-978. doi:10.1016/j.foodchem.2014.10.095
- Medic A, Zamljen T, Hudina M, Solar A, Veberic R. Seasonal variations of naphthoquinone contents (juglone and hydrojuglone glycosides) in *Juglans regia* L. *Sci Hortic*. 2022;300. doi:10.1016/j. scienta.2022.111065
- 25. Ahmad T, Suzuki YJ. Juglone in oxidative stress and cell signaling. *Antioxidants*. 2019;8(4). doi:10.3390/antiox8040091
- Catanzaro E, Greco G, Potenza L, Calcabrini C, Fimognari C. Natural products to fight cancer: A focus on *Juglans regia. Toxins (Basel)*. 2018;10(11). doi:10.3390/toxins10110469
- Shah VM, Rizvi S, Smith A, Tsuda M, Krieger M, Pelz C, et al. Micelle-Formulated Juglone Effectively Targets Pancreatic Cancer and Remodels the Tumor Microenvironment. *Pharmaceutics*. 2023;15(12). doi:10.3390/pharmaceutics15122651
- Schwindl S, Kraus B, Heilmann J. Secondary metabolites from the leaves of *Juglans regia* L. *Biochem Syst Ecol.* 2019;83:130-136. doi:10.1016/j.bse.2019.01.014
- Amaral JS, Seabra RM, Andrade PB, Valentão P, Pereira JA, Ferreres F. Phenolic profile in the quality control of walnut (*Juglans regia* L.) leaves. *Food Chem.* 2004;88(3):373-379. doi:10.1016/j. foodchem.2004.01.055
- Zhao MH, Jiang ZT, Liu T, Li R. Flavonoids in *Juglans regia* L. leaves and evaluation of in vitro antioxidant activity via intracellular and chemical methods. *Scientific World Journal*. 2014;2014. doi:10.1155/2014/303878
- Pop A, Fizeşan I, Vlase L, Rusu ME, Cherfan J, Babota M, et al. Enhanced Recovery of Phenolic and Tocopherolic Compounds from Walnut (*Juglans Regia* L.) Male Flowers Based on Process Optimization of Ultrasonic Assisted-Extraction: Phytochemical Profile and Biological Activities. *Antioxidants*. 2021;10(1):1-25. doi:10.3390/antiox1004
- Lu G, Wang X, Cheng M, Wang S, Ma K. The multifaceted mechanisms of ellagic acid in the treatment of tumors: State-of-theart. *Biomedicine and Pharmacotherapy*. 2023;165. doi:10.1016/j. biopha.2023.115132
- Maekawa T, Miyake T, Tani M, Uemoto S. Diverse antitumor effects of ascorbic acid on cancer cells and the tumor microenvironment. *Front Oncol.* 2022;12. doi:10.3389/fonc.2022.981547
- Bustamante M, Mitcham E. Impact of low oxygen storage on post-storage quality of pasteurized walnut (*Juglans regia* L.) kernels. *Postharvest Biol Technol.* 2023;205. doi:10.1016/j. postharvbio.2023.112542
- Ben Sghaier M, Pagano A, Mousslim M, Ammari Y, Kovacic H, Luis J. Rutin inhibits proliferation, attenuates superoxide production and decreases adhesion and migration of human cancerous cells. *Biomedicine and Pharmacotherapy*. 2016;84:1972-1978. doi:10.1016/j.biopha.2016.11.001

- Giani M, Montoyo-Pujol YG, Peiró G, Martínez-Espinosa RM. Halophilic carotenoids and breast cancer: From salt marshes to biomedicine. *Mar Drugs*. 2021;19(11). doi:10.3390/md19110594
- Bakrim S, El Omari N, Khan EJ, Khalid A, Abdalla AN, Chook JB, et al. Phytosterols activating nuclear receptors are involving in steroid hormone-dependent cancers: Myth or fact? *Biomedicine and Pharmacotherapy*. 2023;169. doi:10.1016/j.biopha.2023.115783
- Stefanucci A, Marinaccio L, Llorent-Martínez EJ, Zengin G, Bender O, Dogan R, *et al.* Assessment of the *in-vitro* toxicity and invivo therapeutic capabilities of *Juglans regia* on human prostate cancer and prostatic hyperplasia in rats. *Food Biosci.* 2024;57. doi:10.1016/j.fbio.2023.103539
- Alshatwi AA, Hasan TN, Shafi G, Syed NA, Al-Assaf AH, Alamri MS, et al. Validation of the antiproliferative effects of organic extracts from the green husk of *Juglans regia* L. on PC-3 human prostate cancer cells by assessment of apoptosis-related genes. *Evidencebased Complementary and Alternative Medicine*. 2012;2012. doi:10.1155/2012/103026
- Genovese C, Cambria MT, D'angeli F, Addamo AP, Malfa GA, Siracusa L, *et al.* The double effect of walnut septum extract (*Juglans regia* L.) counteracts A172 glioblastoma cell survival and bacterial growth. *Int J Oncol.* 2020;57(1):1129-1144.
- Davis PA, Vasu VT, Gohil K, Kim H, Khan IH, Cross CE, *et al.* A highfat diet containing whole walnuts (*Juglans regia*) reduces tumour size and growth along with plasma insulin-like growth factor 1 in the transgenic adenocarcinoma of the mouse prostate model. *British Journal of Nutrition.* 2012;108(10):1764-1772. doi:10.1017/ S0007114511007288
- Li W, Li DY, Wang HD, Zheng ZJ, Hu J, Li ZZ. Juglans regia hexane extract exerts antitumor effect, apoptosis induction and cell circle arrest in prostate cancer cells in vitro. *Tropical Journal of Pharmaceutical Research*. 2015;14(3):399-405. doi:10.4314/tjpr.v14i3.7
- Negi AS, Luqman S, Srivastava S, Krishna V, Gupta N, Darokar MP. Antiproliferative and antioxidant activities of *Juglans regia* fruit extracts. *Pharm Biol.* 2011;49(6):669-673. doi:10.3109/13880209.2 010.537666
- Hasan T, Grace L, Shafi G, Al-Hazzani A, Alshatwi A. Antiproliferative Effects of Organic Extracts from Root Bark of *Juglans regia* L. (RBJR) on MDA-MB-231 Human Breast Cancer Cells: Role of Bcl-2/Bax, Caspases and Tp53. *Asian Pacific Journal of Cancer Prevention,*. 2011;12(1):525-530.
- Zhang XB, Zou CL, Duan YX, Wu F, Li G. Activity guided isolation and modification of juglone from *Juglans regia* as potent cytotoxic agent against lung cancer cell lines. *BMC Complement Altern Med.* 2015;15(1). doi:10.1186/s12906-015-0920-0
- Carvalho M, Ferreira PJ, Mendes VS, et al. Human cancer cell antiproliferative and antioxidant activities of *Juglans regia* L. *Food* and *Chemical Toxicology*. 2010;48(1):441-447. doi:10.1016/j. fct.2009.10.043
- Salimi M, Majd A, Sepahdar Z, *et al.* Cytotoxicity effects of various Juglans regia (walnut) leaf extracts in human cancer cell lines. *Pharm Biol.* 2012;50(11):1416-1422. doi:10.3109/13880209.2012. 682118
- Kadıoğlu Dalkılıç L, Dalkılıç S, Uygur L. Investigation of apoptotic, cytotoxic, and antioxidant effects of *Juglans regia* against MDA-MB-231 and A549 cell lines. *International Journal of Plant Based Pharmaceuticals*. 2023;3(3):62-67. doi:10.29228/ijpbp.17

- Santos A, Barros L, Calhelha RC, Dueñas M, Carvalho AM, Santos-Buelga C, *et al.* Leaves and decoction of *Juglans regia* L.: Different performances regarding bioactive compounds and *in vitro* antioxidant and antitumor effects. *Ind Crops Prod.* 2013;51:430-436. doi:10.1016/j.indcrop.2013.10.003
- Mallavadhani UV, Prasad CV, Shrivastava S, Naidu VGM. Synthesis and anticancer activity of some novel 5,6-fused hybrids of juglone based 1,4-naphthoquinones. *Eur J Med Chem.* 2014;83:84-91. doi:10.1016/j.ejmech.2014.06.012
- Zhang W, Liu A, Li Y, Zhao X, Lv S, Zhu W, *et al.* Anticancer activity and mechanism of juglone on human cervical carcinoma hela cells. *Can J Physiol Pharmacol.* 2012;90(11):1553-1558. doi:10.1139/ y2012-134
- Yousuf M, Shamsi A, Khan P, Shahbaaz M, AlAjmi MF, Hussain A, et al. Ellagic acid controls cell proliferation and induces apoptosis in breast cancer cells via inhibition of cyclin-dependent kinase 6. Int J Mol Sci. 2020;21(10). doi:10.3390/ijms21103526
- Ghafouri-Fard S, Shabestari FA, Vaezi S, Abak A, Shoorei H, Karimi A, *et al.* Emerging impact of quercetin in the treatment of prostate cancer. *Biomedicine and Pharmacotherapy.* 2021;138. doi:10.1016/j.biopha.2021.111548
- Golmohammadi M, Zamanian MY, Jalal SM, Noraldeen SA, Ramírez-Coronel AA, Oudaha K, *et al.* A comprehensive review on Ellagic acid in breast cancer treatment: From cellular effects to molecular mechanisms of action. *Food Sci Nutr.* 2023;11(12):7458-7468. doi:10.1002/fsn3.3699
- 55. Meng X, Cui Z, Shi H, Ma X, Li W, Liu X, *et al.* Ellagic acid inhibits cell proliferation, migration, and invasion of anaplastic thyroid cancer cells via the Wnt/β-catenin and PI3K/ Akt pathways. *Acta Biochim Pol.* 2023;70(1):109-115. doi:10.18388/abp.2020_6317
- Reyes-Farias M, Carrasco-Pozo C. The anti-cancer effect of quercetin: Molecular implications in cancer metabolism. *Int J Mol Sci.* 2019;20(13). doi:10.3390/ijms20123177
- Pellegrino M, Bevacqua E, Frattaruolo L, Cappello AR, Aquaro S, Tucci P. Enhancing the Anticancer and Anti-Inflammatory Properties of Curcumin in Combination with Quercetin, for the Prevention and Treatment of Prostate Cancer. *Biomedicines*. 2023;11(7). doi:10.3390/biomedicines11072023
- Factor VM, Laskowska D, Jensen MR, Woitach JT, Popescu NC, Thorgeirsson SS. Vitamin E Reduces Chromosomal Damage and Inhibits Hepatic Tumor Formation in a Transgenic Mouse Model. Proc Natl Acad Sci. 2000;97(5):2196-2201. doi: 10.1073/ pnas.040428797.
- Giani M, Montoyo-Pujol YG, Peiró G, Martínez-Espinosa RM. Haloarchaeal carotenoids exert an *in vitro* antiproliferative effect on human breast cancer cell lines. *Sci Rep.* 2023;13(1). doi:10.1038/ s41598-023-34419-x
- Kim JA, Jang JH, Lee SY. An updated comprehensive review on vitamin a and carotenoids in breast cancer: Mechanisms, genetics, assessment, current evidence, and future clinical implications. *Nutrients*. 2021;13(9). doi:10.3390/nu13093162
- Karim S, Akhter MH, Burzangi AS, Alkreathy H, Alharthy B, Kotta S, et al. Phytosterol-Loaded Surface-Tailored Bioactive-Polymer Nanoparticles for Cancer Treatment: Optimization, *In Vitro* Cell Viability, Antioxidant Activity, and Stability Studies. *Gels.* 2022;8(4). doi:10.3390/gels8040219

Cite this article: Requejo-Rodríguez CY, Roncal-Alayo EM, Silva-Correa CR, Villarreal-La Torre VE, Sagástegui-Guarniz WA, Gamarra-Sánchez CD, Janampa-Castillo WE, et al. *Juglans regia* L.: Source of Bioactive Compounds with Potential Anticancer Activity. Pharmacogn J. 2024;16(5): 998-1003.